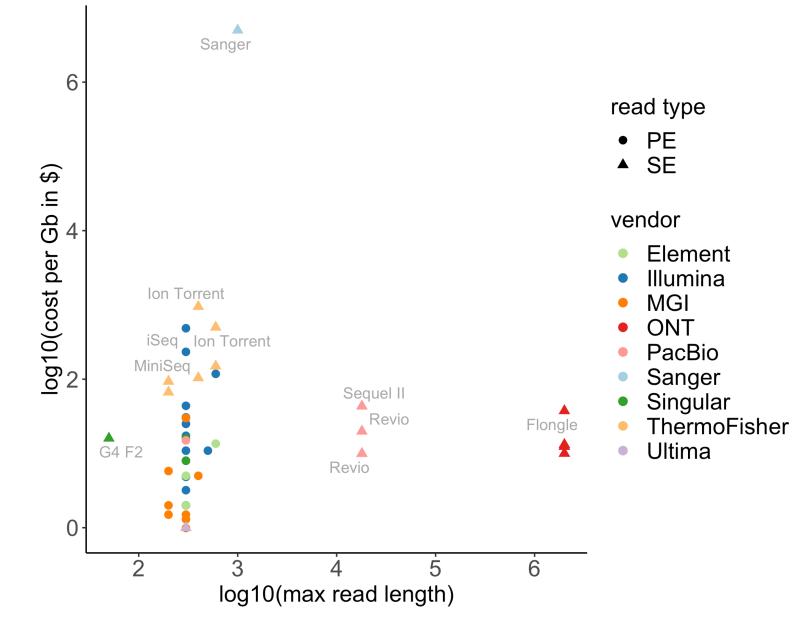
### Long-read sequence analysis

Sequencing technologies

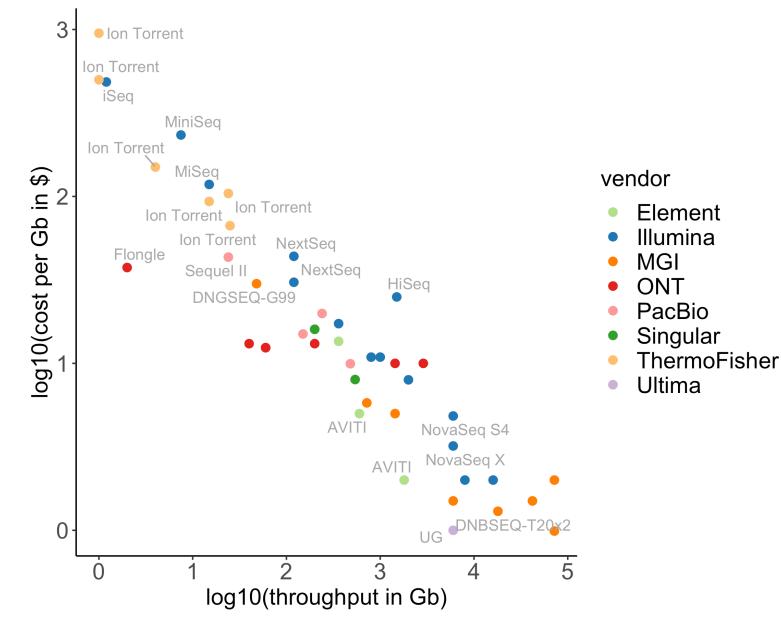
### Question 4

# What is a long read?

- Short read: 50-300 bp, often paired-end (Illumina sequencing)
- Long read: > 1kb, up to 20 Mb:
  - single molecule sequencing or
  - 3<sup>rd</sup> generation sequencing



drawn from: <u>https://docs.google.com/spreadsheets/d/1GMMfhyLK0-</u> <u>q8XkIo3YxlWaZA5vVMuhU1kg41g4xLkXc/</u> Albert Vilella



drawn from: <u>https://docs.google.com/spreadsheets/d/1GMMfhyLK0-</u> <u>q8XkIo3YxIWaZA5vVMuhU1kg41g4xLkXc/</u> Albert Vilella

# Sequencing-by-synthesis

- 2nd generation sequencing
- Massive throughput: up to 500x10<sup>9</sup> bases/run
- Illumina still most used platform today

Element Biosciences Biosciences Biosciences Genomics

# Sequencing-by-synthesis

- 50 300 bp
- Paired-end (or single-end)

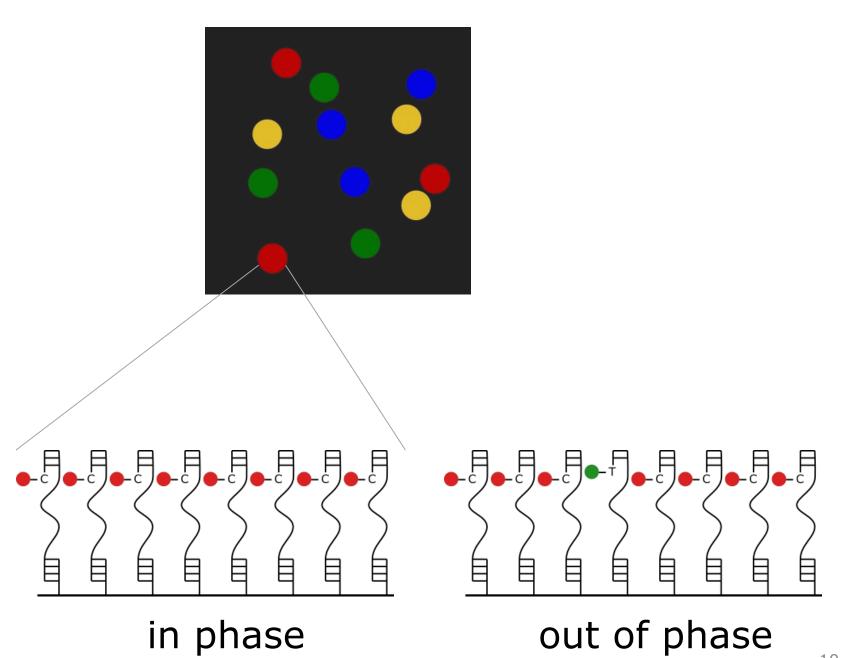
paired-end

Image from: Illumina (2020)

# Question 5

# SBS - limitations

- Maximum read length: 300 bp
- How to reconstruct:
  - Repeats?
  - Isoforms?
  - Structural variation?
  - Haplotypes?
  - Genomes?
- Why not longer read lengths with Illumina?



# Long reads (3rd generation)

- Crux: maximizing signal from a singlemolecule base read-out
- Single molecule, so no out-of-phase signal
- Two frequently used platforms:
  - PacBio SMRT sequencing
  - Oxford Nanopore Technology





# Question 6

# Oxford Nanopore technology

Based on changes in electrical current

-C -G

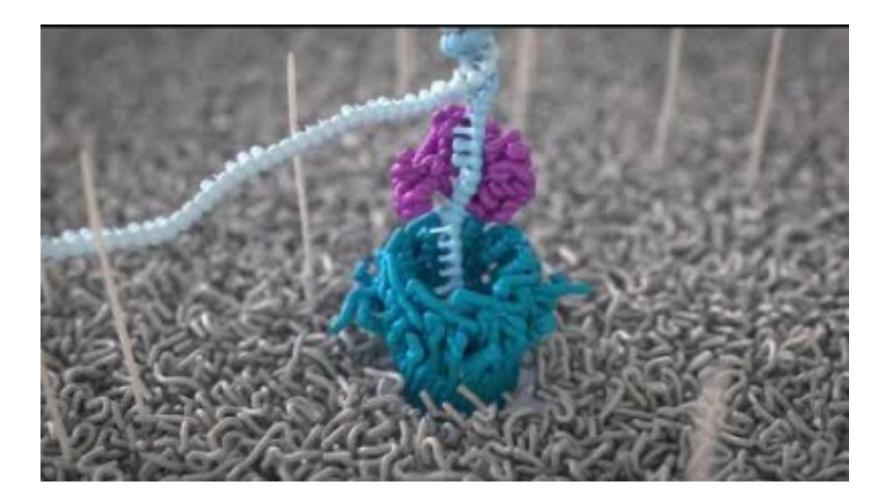
G

G

TAACCAG

- Well-known for its scalability and portability
- 4 bp read at a time
- Up to ~95-99% accuracy
- Errors can be biased





# ONT scalability

1 small flow cell: 1 x 2.8 Gb 1 medium flow cell: 1 x 50 Gb 5 medium flow cells: 5 x 50 Gb

Grid**ION**×5

24-48 big flow cells: 48 x 290 Gb





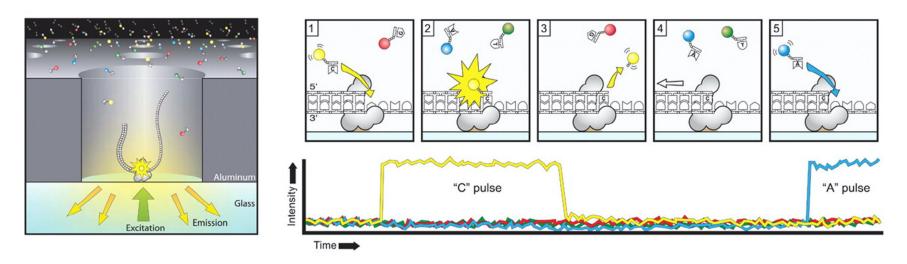
FlongleMinIONGridIONPi

https://nanoporetech.com/products#comparison

# ONT library prep

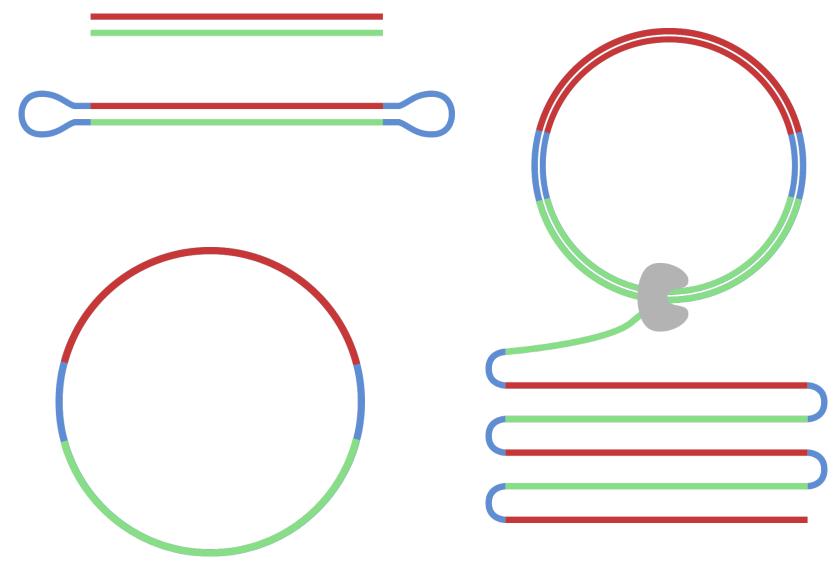
- Standard kit:
  - >1  $\mu$ g HMW DNA
  - Shearing + size selection is optional
  - Multiplexing requires PCR step

### PacBio sequencing



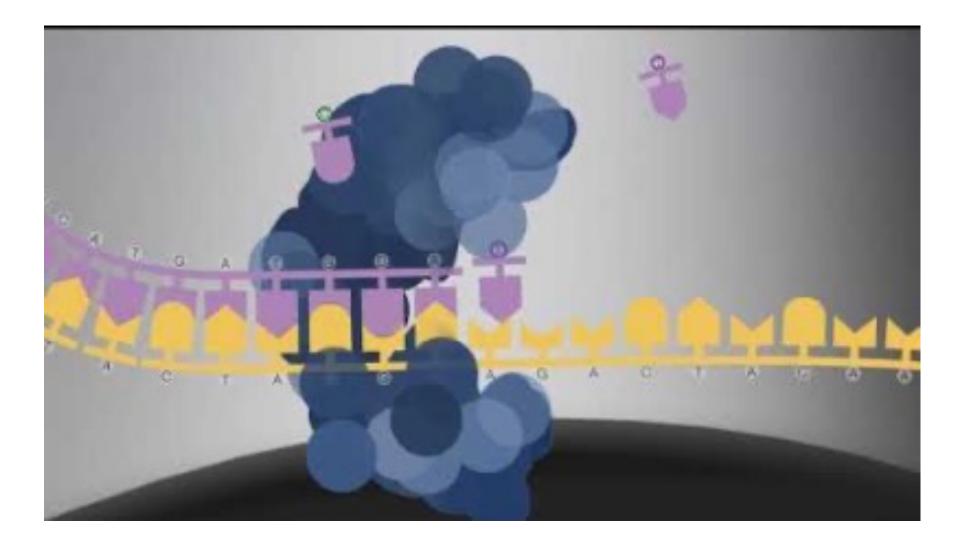
- Polymerase bound to ZMW bottom
- Circular molecules
- Single read out ~90% accuracy
- HiFi: single molecule sequenced multiple times

**Image from:** Rhoads A, Au KF. Genomics Proteomics Bioinformatics 2015;13:278–89



#### Hi-Fi read

Image from: https://doi.org/10.5281/zenodo.4636860



# PacBio Sequel IIe



- 8M ZMW
- ~2M HiFi reads/SMRT cell

### Pacbio Revio



- 25M ZMW
- ~5-6M HiFi reads/SMRT cell
- 2-4 SMRT cells/run

# PacBio library prep

- start with >5  $\mu$ g HMW DNA
- Requires shearing + size selection
- Multiplexing requires PCR

	ONT	PacBio
Read accuracy	~90-95%	> 99.99% (HiFi)
Read length	up to 2 Mb	up to 30-40 kb (HiFi) typically ~15-20 kb
RNA base modifications	Yes (m6A) <sup>1</sup>	No
DNA base modifications	Yes (m5C, m6A) <sup>2</sup>	Yes (m5C, m6A, hm5C) <sup>3</sup>
Throughput (BIF)	10 Tb ~500M reads/run <sup>4</sup>	480 Gb ~25M HiFi reads/run <sup>5</sup>

1. Liu, H., et al (2019). Accurate detection of m6A RNA modifications in native RNA sequences. *Nature Communications*, 10(1), 1–9

2. Liu, Q., et al (2019). Detection of DNA base modifications by deep recurrent neural network on Oxford Nanopore sequencing data. *Nature Communications*, *10*(1).

3. Flusberg, B. A., et al (2010). Direct detection of DNA methylation during singlemolecule, real-time sequencing. *Nature Methods*, 7(6), 461–465

4. 48 flow cells on a PromethIon

5. 4 SMRT cells on a Revio

### Question 7&8