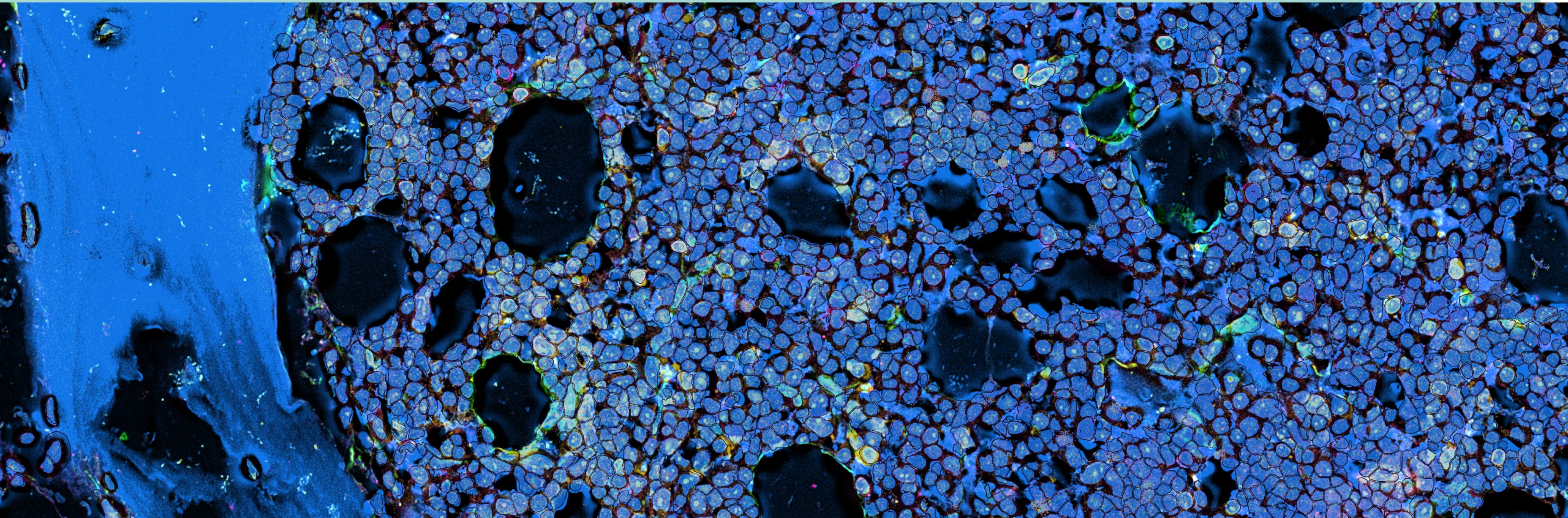


Cell segmentation for spatial transcriptomics (ST)

Ivan Berest, 03.06.2026



Outline

- 1 Intro to the cell segmentation with ST
- 2 Imaging-based ST segmentation methods
- 3 Sequencing-based ST segmentation methods
- 4 Spillover/bleeding challenge
- 5 Analysis example

Why location matters: from bulk to spatial



bulk omics



single-cell omics

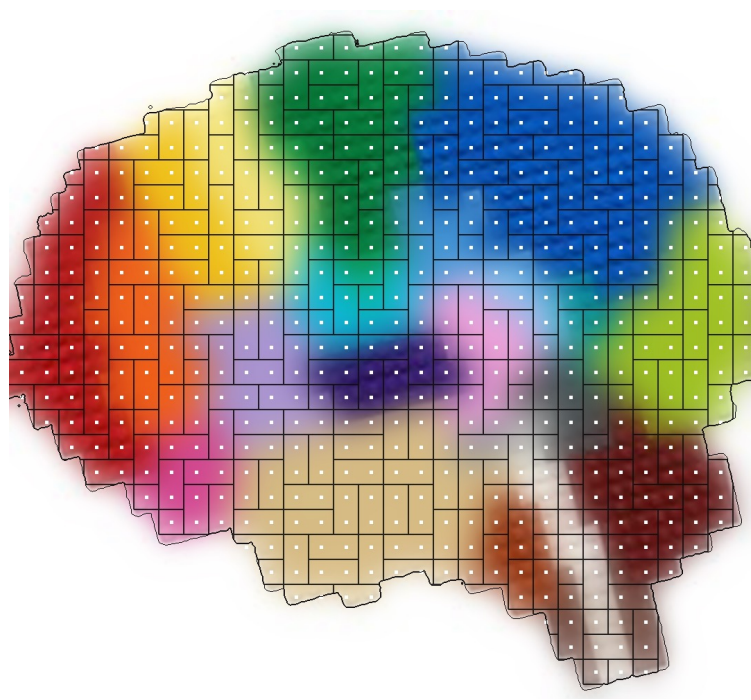


spatial omics

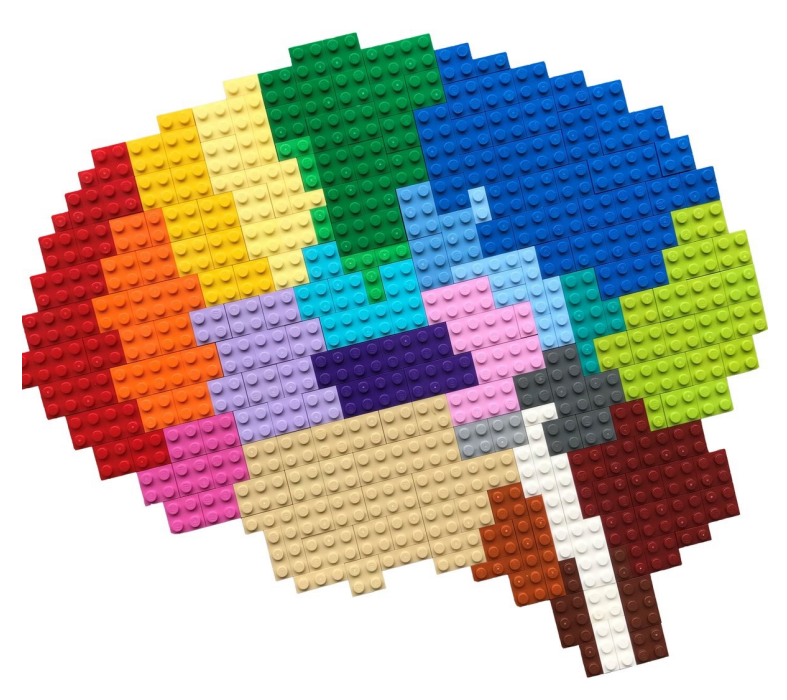
Segmentation is necessary – and never perfect



usual input



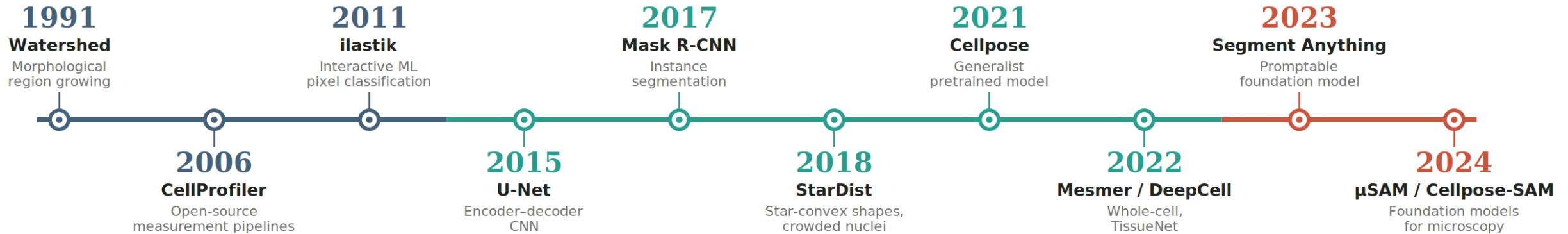
cell segmentation



real scenario

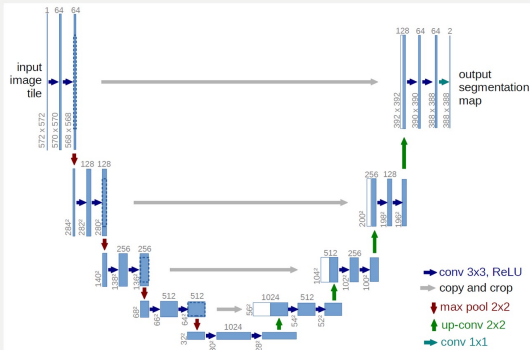
Cell segmentation is not new concept: whole era in imaging!

● Classical ● Deep learning ● Foundation models



U-Net: Convolutional Networks for Biomedical Image Segmentation

The u-net is convolutional network architecture for fast and precise segmentation of images. Up to now it has outperformed the prior best method (a sliding-window convolutional network) on the ISBI challenge for segmentation of neuronal structures in electron microscopic stacks. It has won the Grand Challenge for Computer-Automated Detection of Caries in Bitewing Radiography at ISBI 2015, and it has won the Cell Tracking Challenge at ISBI 2015 on the two most challenging transmitted light microscopy categories (Phase contrast and DIC microscopy) by a large margin (See also our announcement).



U-net architecture (example for 32x32 pixels in the lowest resolution). Each blue box corresponds to a multi-channel feature map. The number of channels is denoted on top of the box. The x-y-size is provided at the lower left edge of the box. White boxes represent copied feature maps. The arrows denote the different operations.

StarDist - Object Detection with Star-convex Shapes

2D:

3D:

Metadata: pypi package 0.9.2, Anaconda.org 0.9.2, Test passing, Test (PyPI) failing, forum 535 topics, downloads 30k/month

2018 Data Science Bowl

Find the nuclei in divergent images to advance medical discovery

Cellpose

docs passing, tests passing, codecov 42%, pypi package 4.1.1, downloads 2M, downloads/month 81k, python 3, license BSD-3-Clause, contributors 70, website up, forum 664 topics, repo size 115.3 MiB, Stars 2.2k, Forks 627

Cellpose-SAM: cell and nucleus segmentation with superhuman generalization. It can be optimized for your own data, applied in 3D, works on images with shot noise, (an)isotropic blur, undersampling, contrast inversions, regardless of channel order and object sizes.

To learn about Cellpose-SAM read the paper or watch the talk. For info on fine-tuning a model, watch this tutorial talk, and see this example video of human-in-the-loop training. For support, please open an issue.

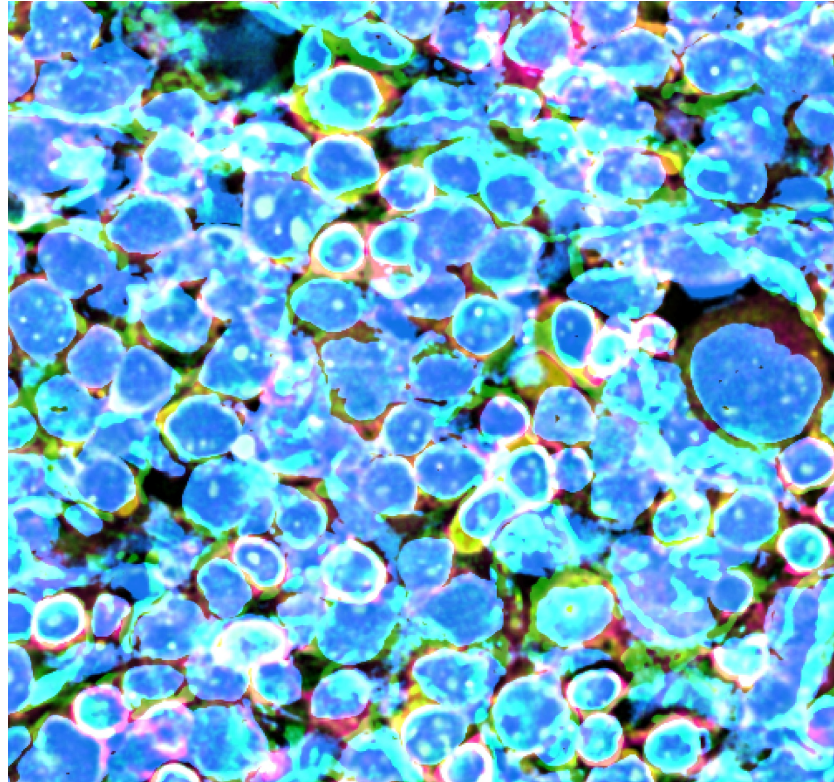
Article | [Open access](#) | Published: 08 December 2025

CellSAM: a foundation model for cell segmentation

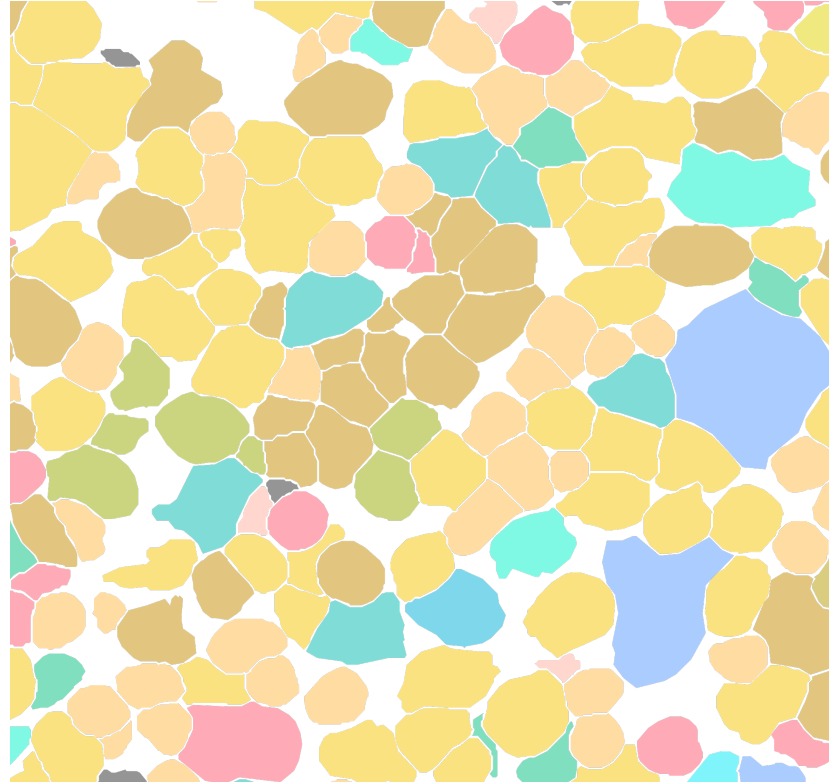
Markus Marks, Uriah Israel, Rohit Dilip, Qilin Li, Changhua Yu, Emily Laubscher, Ahamed Iqbal, Elora Pradhan, Ada Ates, Martin Abt, Caitlin Brown, Edward Pao, Shenyi Li, Alexander Pearson-Goulart, Pietro Perona, Georgia Gkioxari, Ross Barnowski, Yisong Yue & David Van Valen

Nature Methods 22, 2585–2593 (2025) | [Cite this article](#)

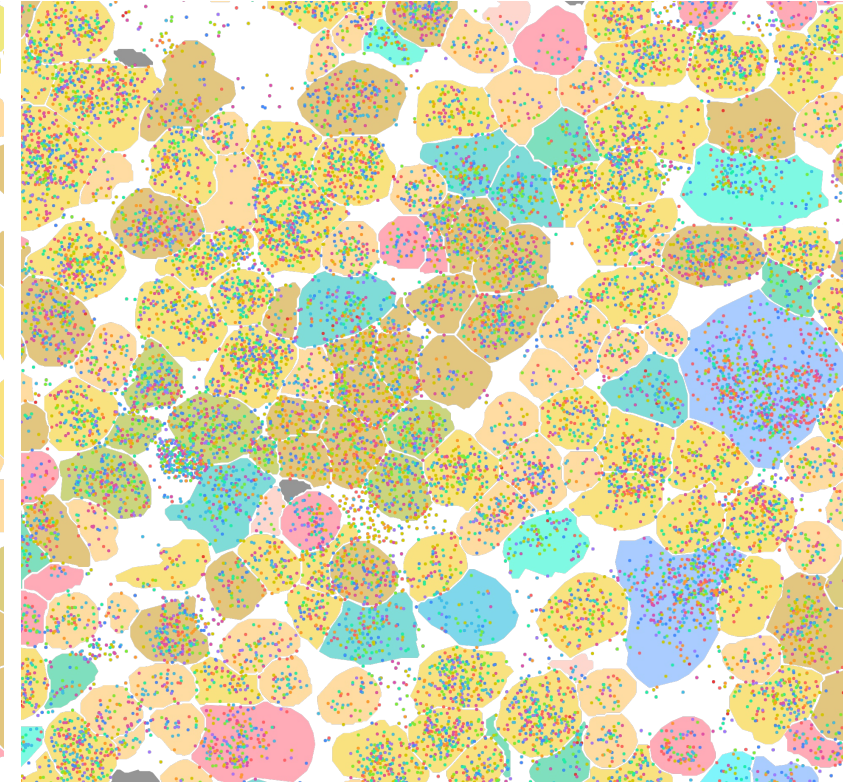
New challenges for cell segmentation with spatial transcriptomics



IF image



segmentation mask



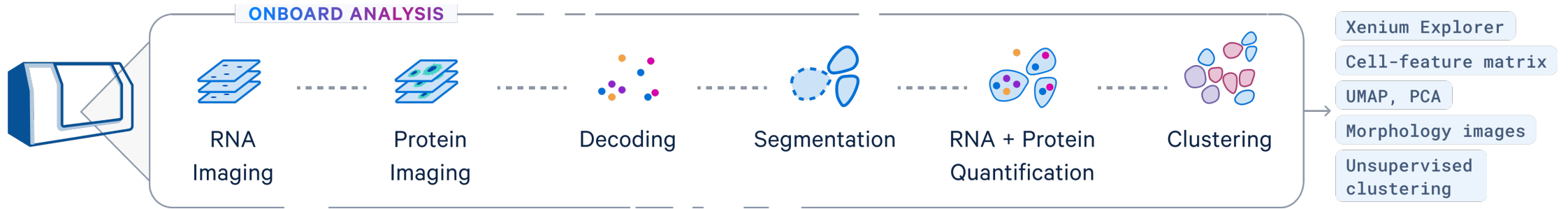
overlaid transcripts

Transcripts for the cells can improve cell segmentation based only on the image?

Outline

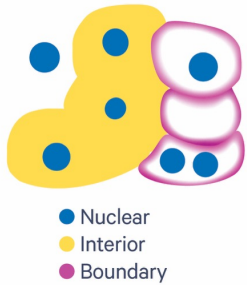
- 1 Intro to the cell segmentation with ST
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-

Xenium Ranger segmentation algorithms



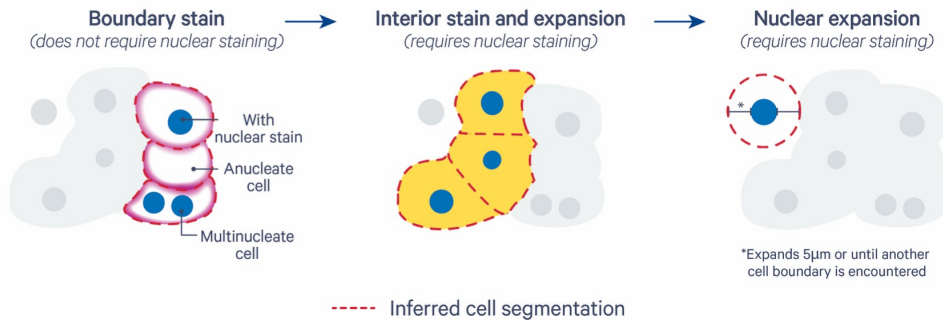
Xenium onboard analysis (XOA) happening directly on the machine

Types of Stains



- DAPI nuclear staining
- ATP1A1/CD45/E-Cadherin boundary staining
- 18S RNA + alphaSMA/vimentin protein interior

Types of Cell Segmentation

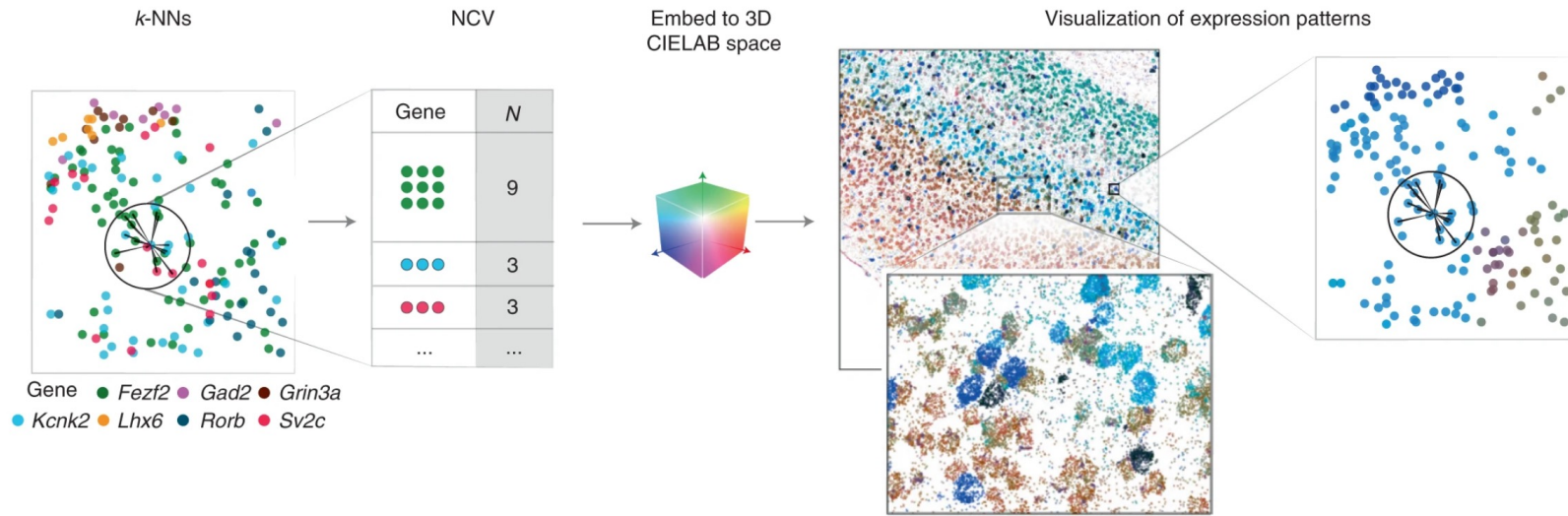


Possible to rerun segmentation later with *xeniumranger resegment* pipeline

Prioritization order:

1. Segment cells based on their cell boundary stain
2. Segment cells based on expansion from the nucleus to the cell interior stain edge
3. Nuclear expansion (default 5 µm)

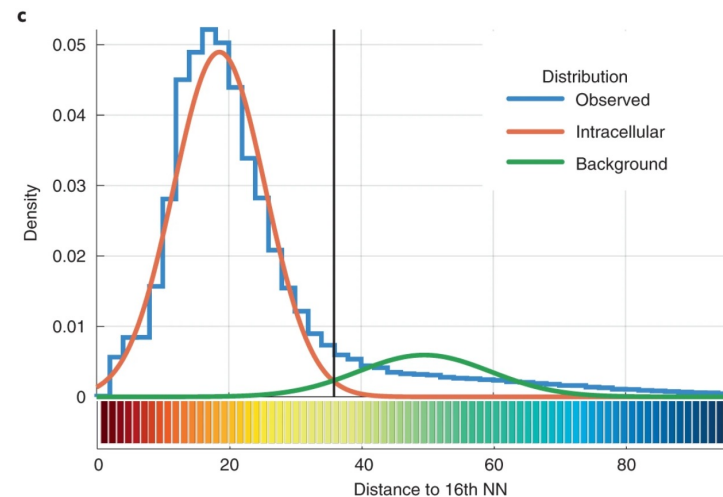
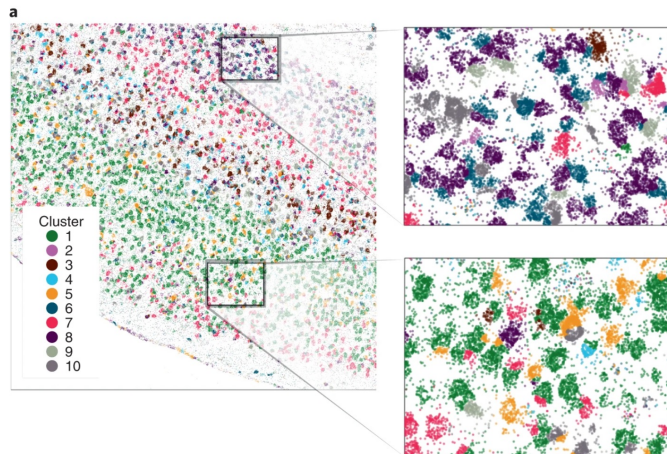
Bayesian Segmentation of Spatial Transcriptomics Data (*Baysor*)



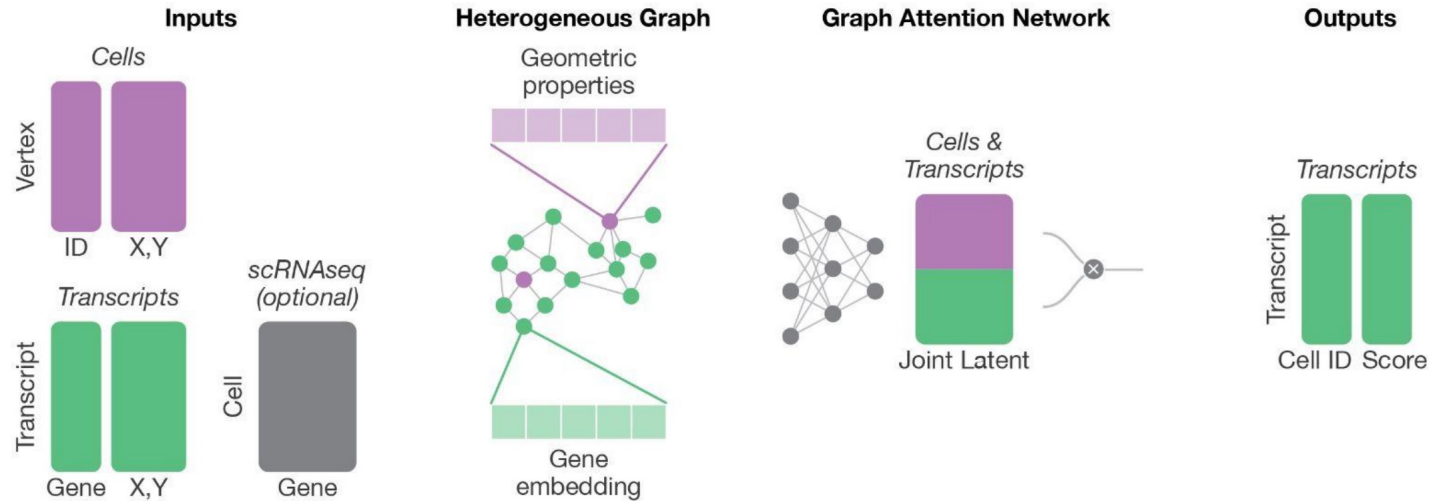
First “big” method that used segmentation-free approach based on transcriptional composition of the cells

Baysor can take advantage of auxiliary stains by incorporating a precalculated segmentation as a probabilistic prior (user can modify weight of the prior)

Really demanding for memory and running time (easy up to 1TB memory, 100s of threads and several days run for 5k Xenium panel) bejsor

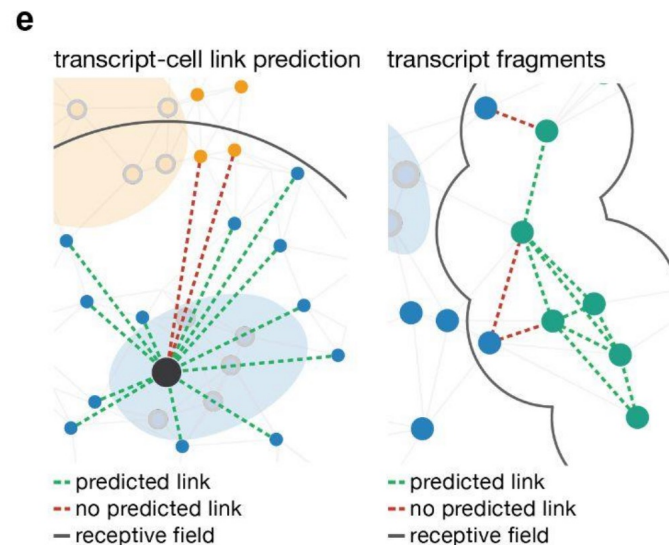
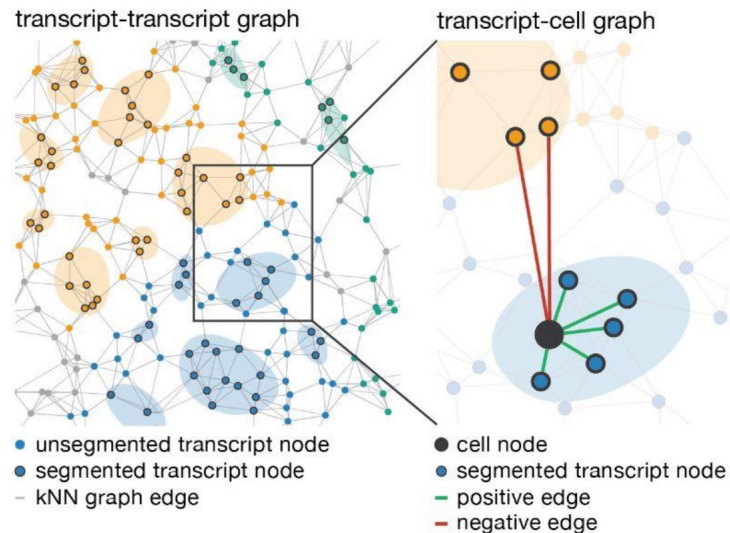


Using graph neural network (GNN) to solve transcripts assignment (*Segger*)



Use as input initial segmentation (i.e. from xeniumranger). Can be supervised by co-expression patterns with additional matching scRNA-seq.

Reframes segmentation problem to the link-prediction problem on a graph (transcripts and cells are nodes). GNN learn meaningful patterns based on transcript colocalization + nuclear/membrane staining



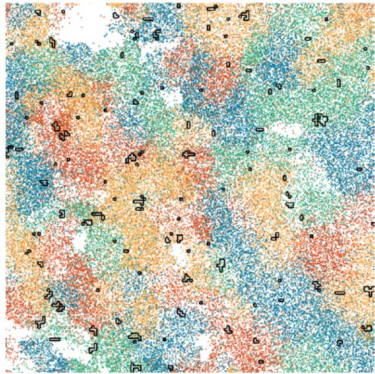
Because of the GNN can be used for the cases with ambiguous border transcripts

Using GPU to run, faster than Baysor, but still takes some time

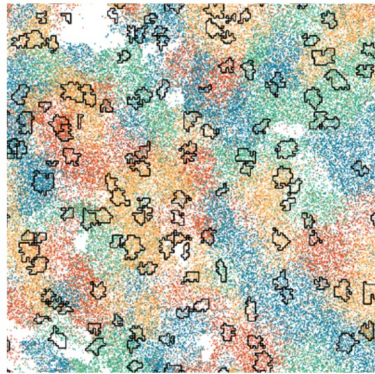
Probabilistic cell segmentation for in situ spatial transcriptomics (*Proseg*)

b Proseg

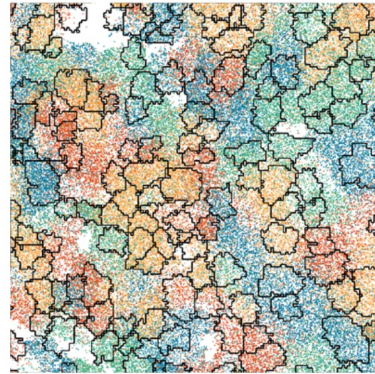
CPM-adapted segment cells by optimizing likelihood of observed transcripts



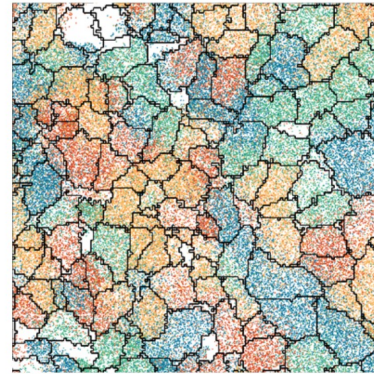
Iteration 1



Iteration 10

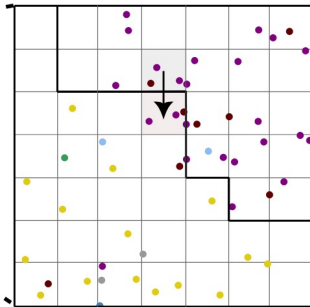
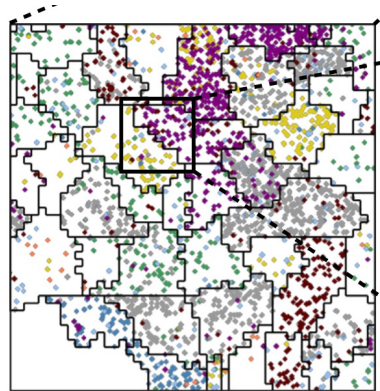


Iteration 100

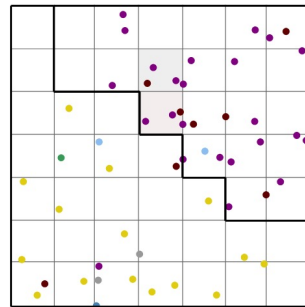


Iteration 500

Gene 1 (red), Gene 2 (blue), Gene 3 (green), Gene 4 (yellow)



Propose copying adjacent cell label



Accept with probability $\min\left(1, \frac{P(\sigma'|\theta)q(\sigma \rightarrow \sigma')}{P(\sigma|\theta)q(\sigma \rightarrow \sigma)}\right)$

- SERPINA1, MET, LAMP3, ERBB3 (tumor 1)
- S100A9, PKM (tumor 2)
- CSF3R, ITGAX (neutrophils)
- COL1A1, FN1, PDGFRB (fibroblasts)
- COL4A1, PLVAP, PECAM1, VWF, ENG (endothelial)
- CD2, CD3E, CD3D, TRAC (T cells)
- CD79A, MZB1, POU2AF1, XBP1 (B cells)
- LYZ, FCGR3A, C1QC, CD14 (macrophages)

What set of cell shapes would best explain where the observed transcripts actually landed?

Initialize cell morphologies from nuclear segmentation and iterated/expanded until best fits all transcripts

Models leaky cells (particular in Xenium data) by adding prior on the distance transcripts migrate from true position

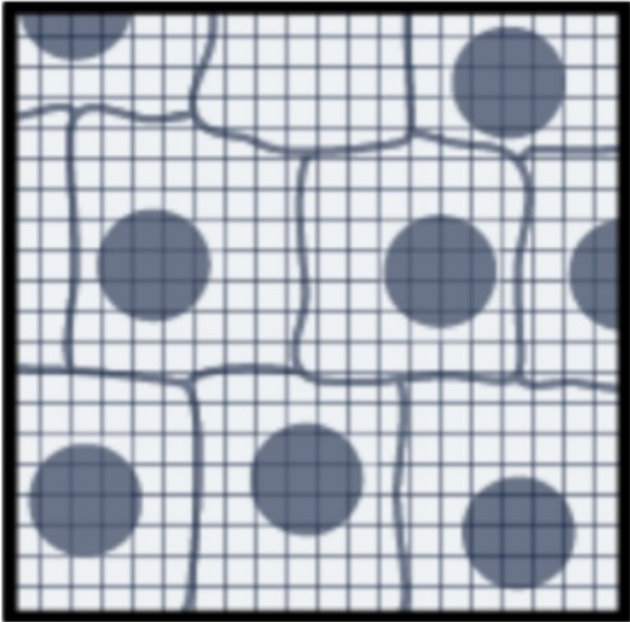
Quite fast, current “gold standard”

Outline

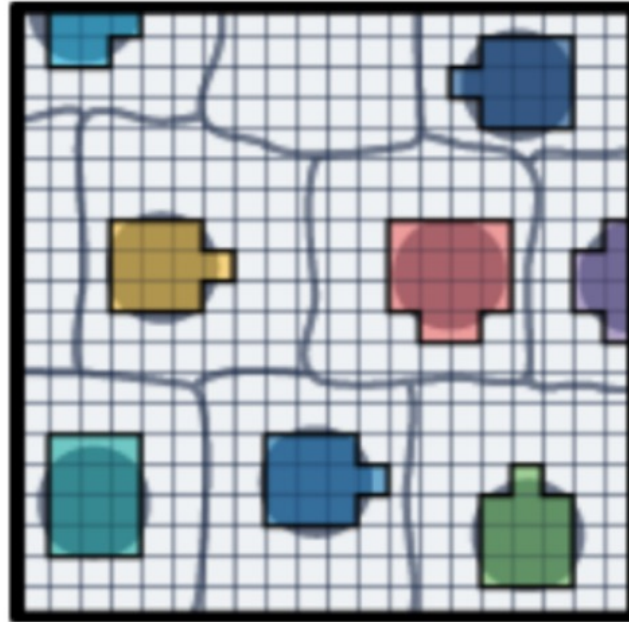
- 1 Intro to the cell segmentation with ST
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-

Cell segmentation from 10x *Space Ranger*

Cells



Nucleus based bins



Expanded bin

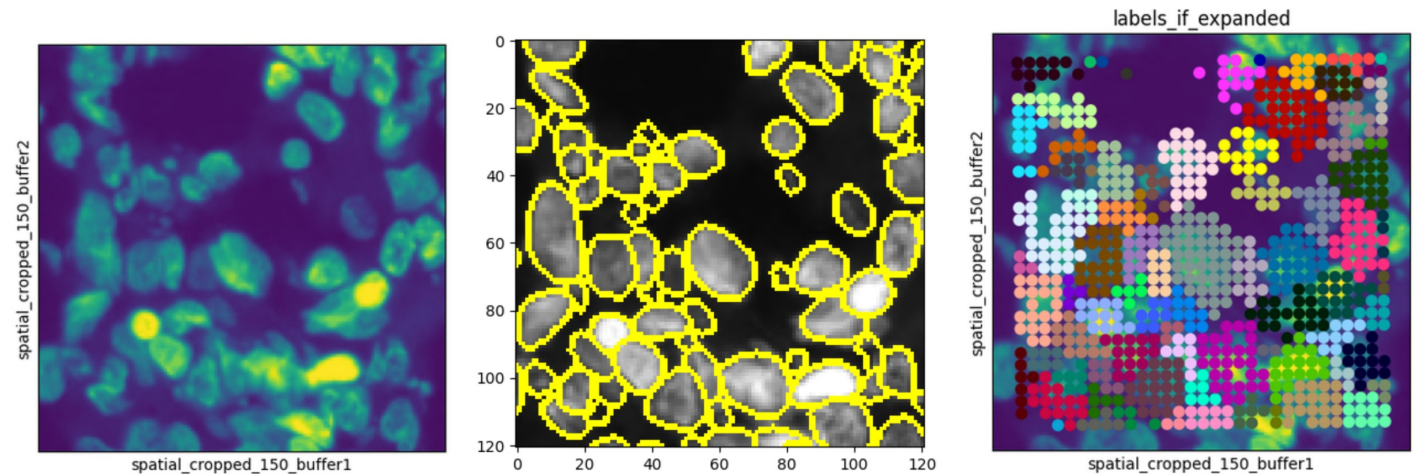
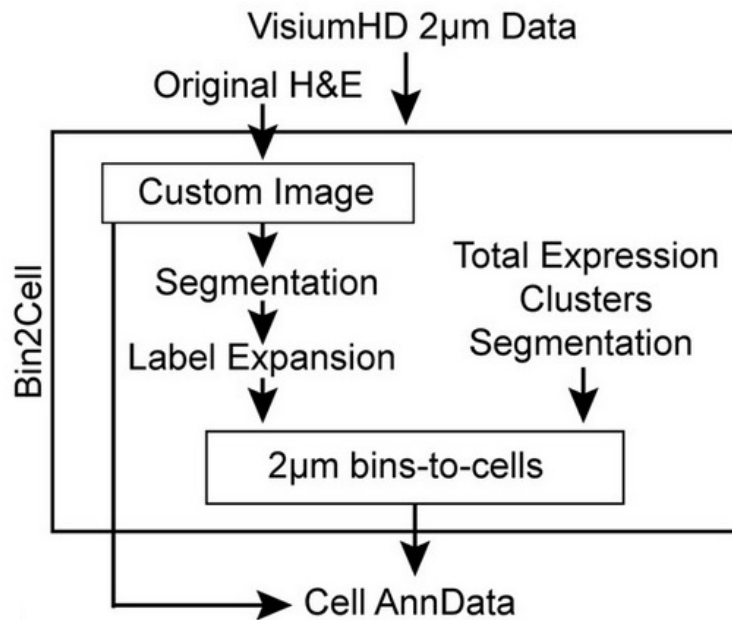


Trained StarDist v6 model on 17k image patches from 150 H&E tissue images for nuclei segmentation

Each spot is assigned to the closest nucleus up until a maximum distance specified by *--nucleus-expansion-distance-micron*

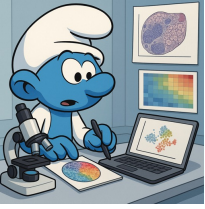
Bin2cell merge VisiumHD bins into cells

- Use cell segmentation algorithm (StarDist) on pre-trained H&E or nuclear marker model
- Label expansion of a fixed distance (default 2 bins)

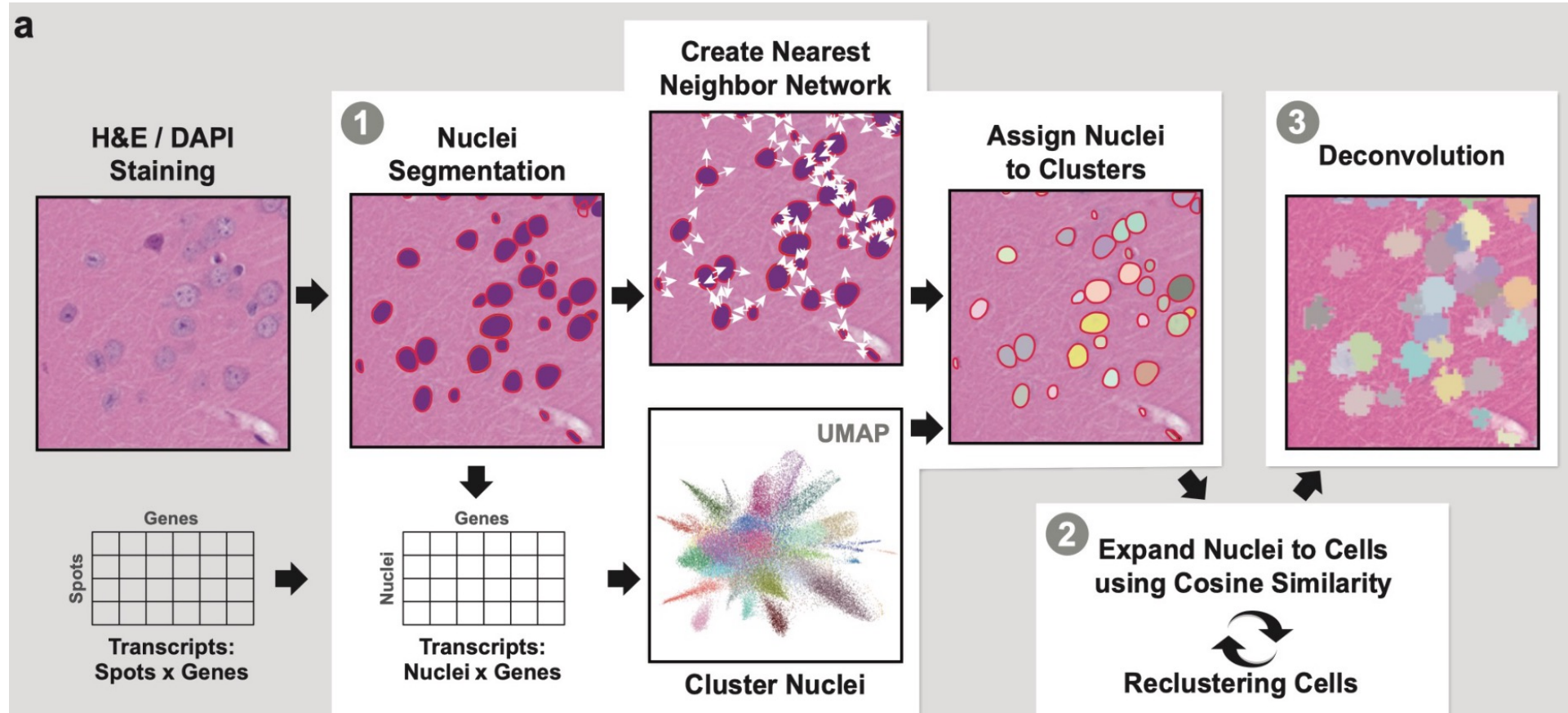


Combine label expanded cell segmentation with segmentation based on GEX counts (fill cells without nucleus)

If bin overlap segmentation masks (cell) add proportional counts of the spot to the respective cells

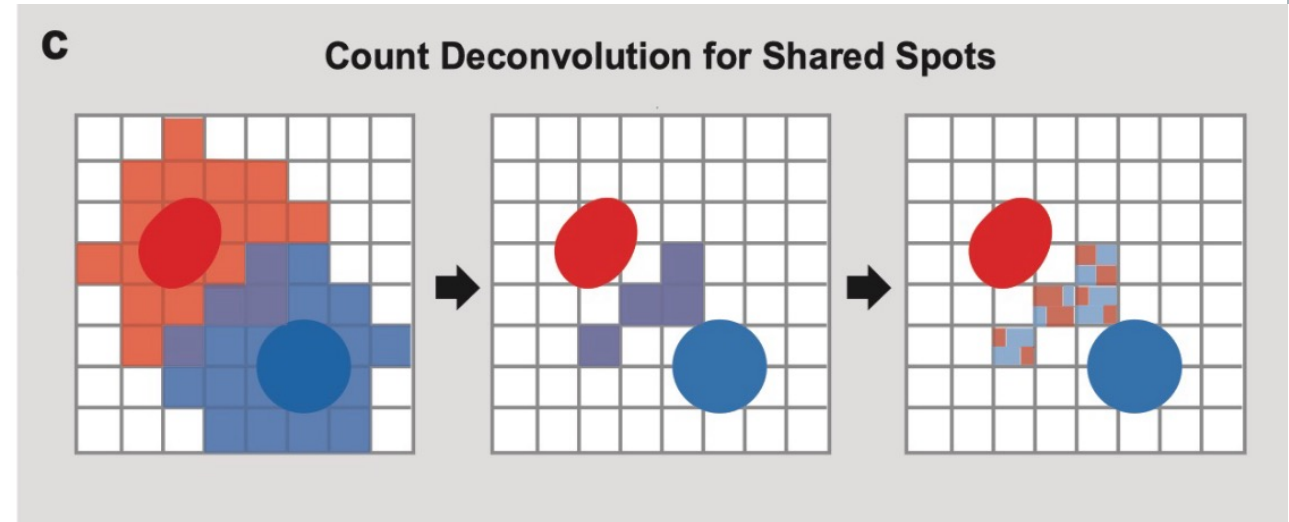
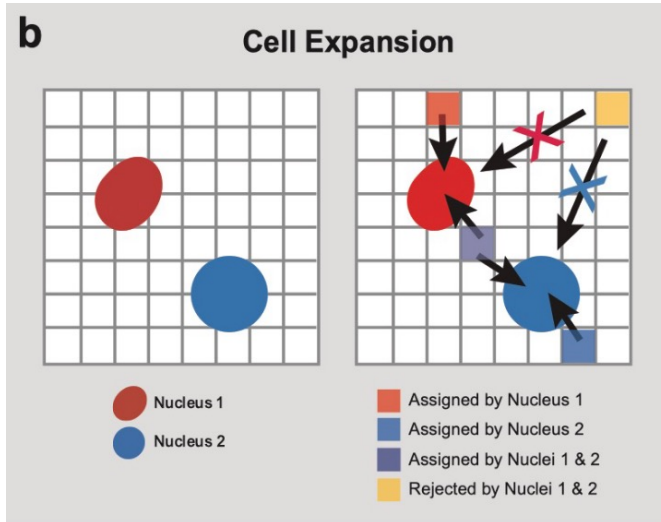
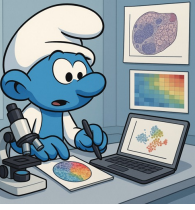


Segmentation and Manifold Unrolling Framework (*SMURF*)



Works with associated H&E or IF stainings + uses prior nuclei segmentation (i.e. StarDist)
Calculate nearest neighbor scores for cells and cluster nuclei data with further assignment to clusters
Link imaging data to spots (50 pixels in one spot)

Segmentation and Manifold Unrolling Framework (*SMURF*)



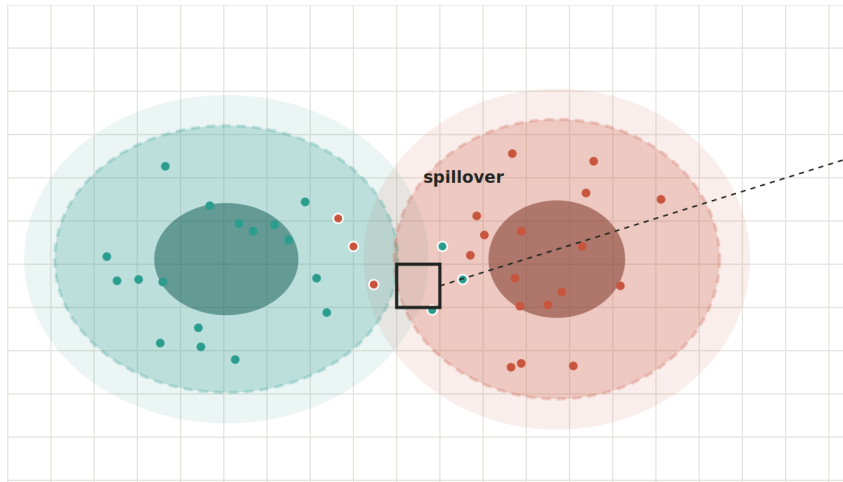
- Adds nearby spots to the nucleus gene expression matrix to increase UMI counts by using concentric expansion
- In each iteration check if the resulted expanded cells increase cosine similarity between cells of the same cluster
- Repeated until clustering is stabilized based on normalized mutual information (NMI)

- Deconvolve counts from shared spots so that every transcript is assigned to at most one cell
- The sum of cell type proportions (weights from clustering) should match the normalized gene expression pattern for the spot

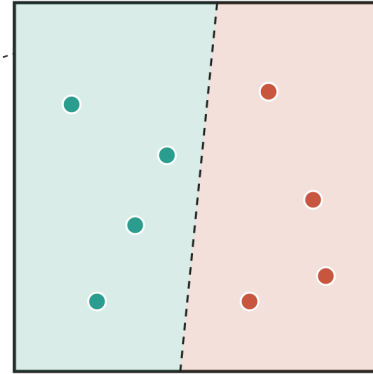
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New challenges for cell segmentation with spatial transcriptomics



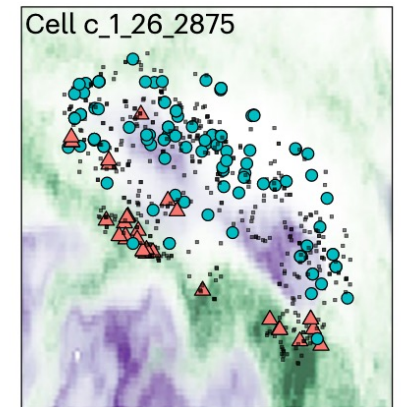
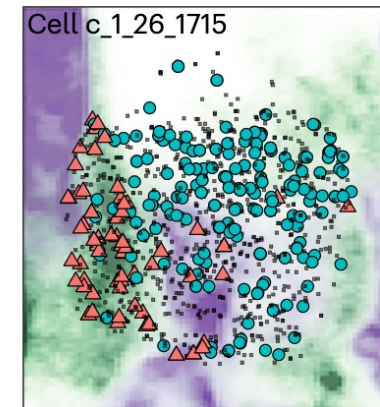
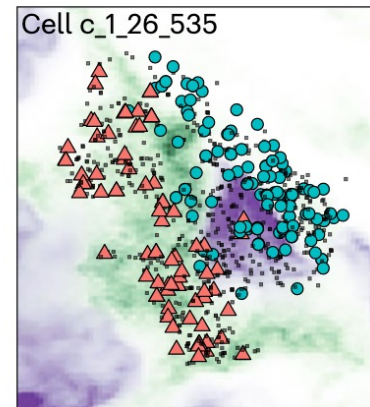
One bin, two cells



→ mixed signal, unreliable cell typing

In Visium HD (sequencing-based ST) source of spillover is inherited from the technique. One bin/spot covering 2 or more cells add noise to this cells, corrupting all downstream analysis (annotation, DE expression)

Spillover effect is evident also in imaging-based ST (i.e. Xenium), where the technical source is diffusion of the RNA molecules to the neighboring cells (i.e. transcriptionally active malignant cells contaminate healthy neighboring cells)



Marker type

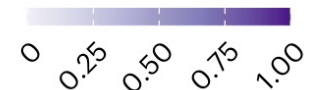
- Fibroblast
- Nonmarker

▲ Malignant

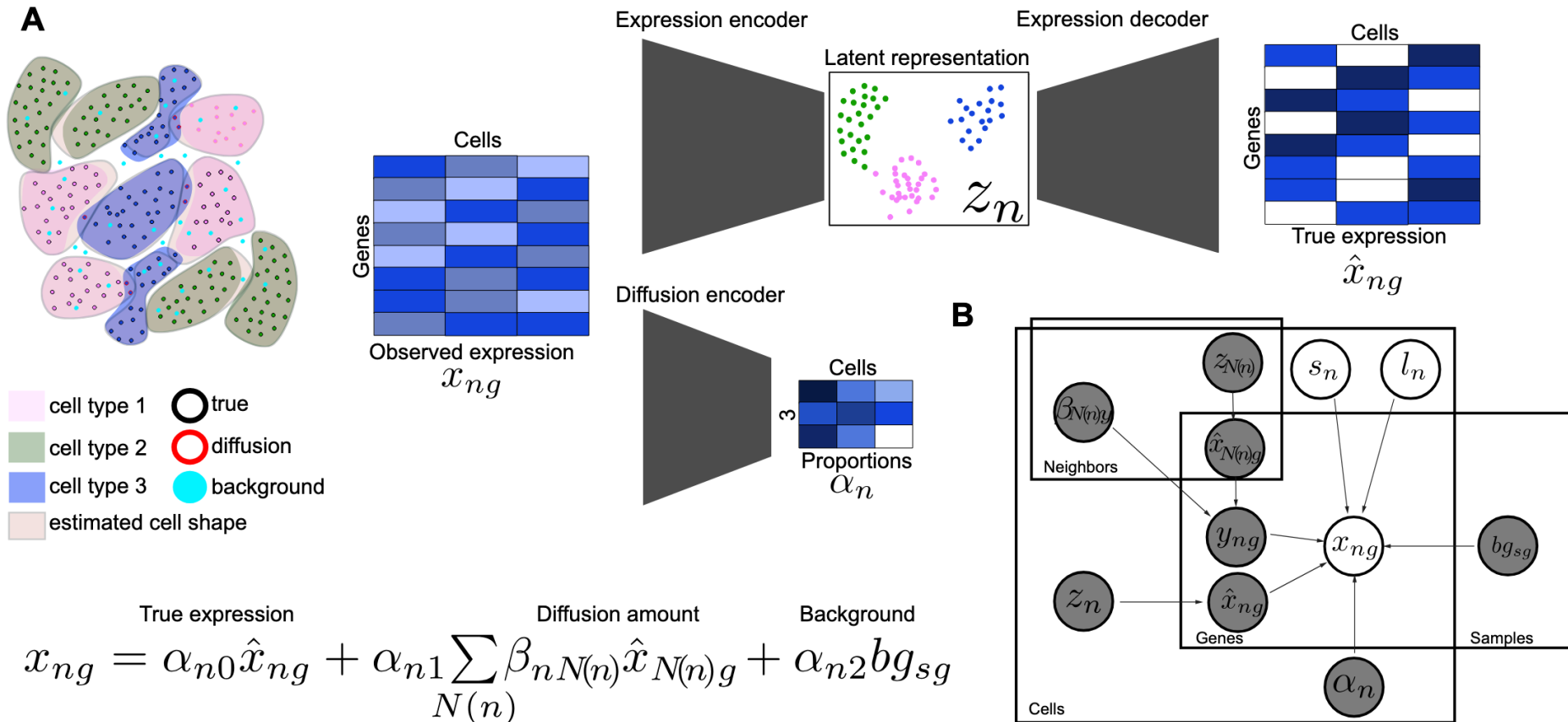
Membrane



DAPI



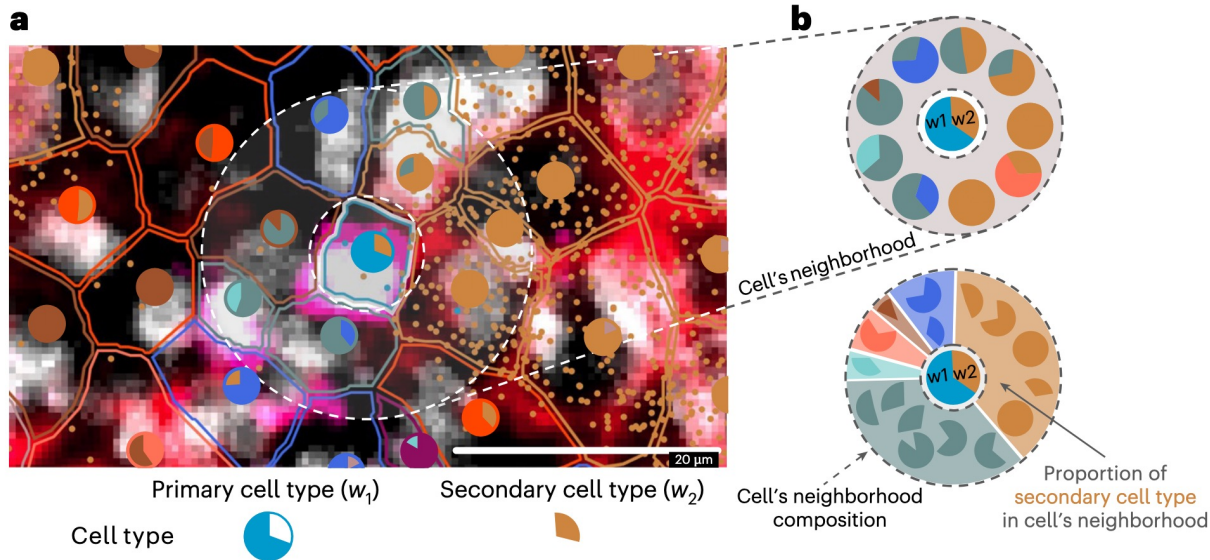
ResolVI - addressing noise and bias in spatial transcriptomics



Uses as input matrix of cell by gene expression, spatial coordinates of each cell and respective tissue slice (sample integration)

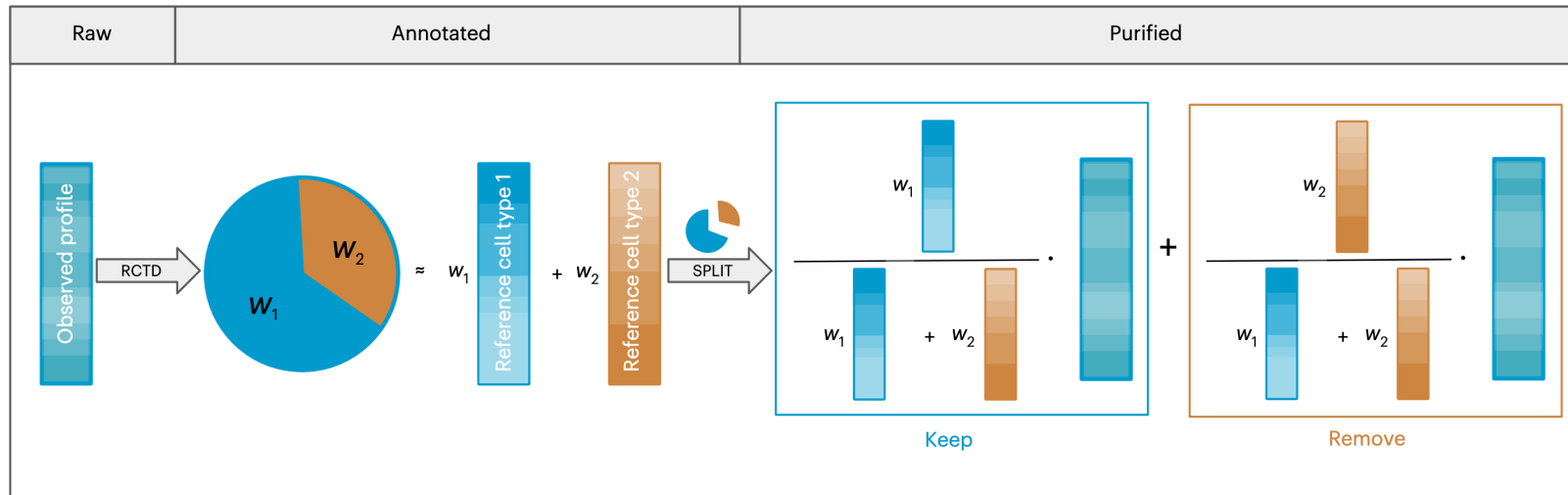
Backbone latent variable model on the variational autoencoder
 Estimates observed expression as a sum of true expression, expression induced by neighboring cells (diffusion) and unspecific background expression

Spatial Purification of Layered Intracellular Transcripts (*SPLIT*)



R package that utilize first RCTD with reference/matching scRNA-seq data for initial decomposition of the cells (similarity of each cell transcriptome profile to the provided celltypes)

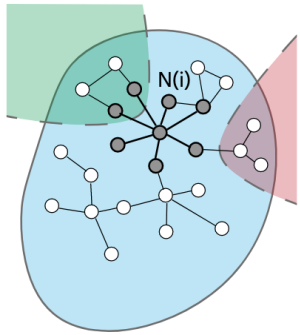
Identify doublet-cells (potential spilling)



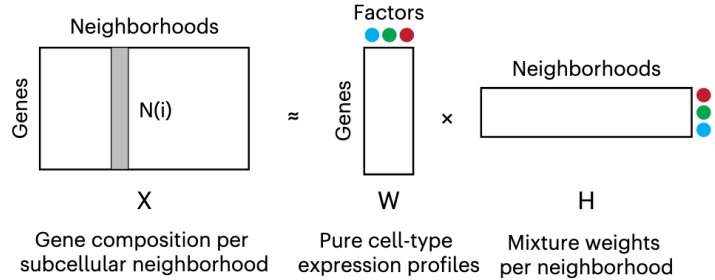
Correct raw counts using doublet-SPLIT approach, which keep for the cells of interest (depending on the mode) only fraction of the read corresponding to the primary celltype relative to the sum of primary and secondary

Correction with segmentation errors with *cellAdmix*

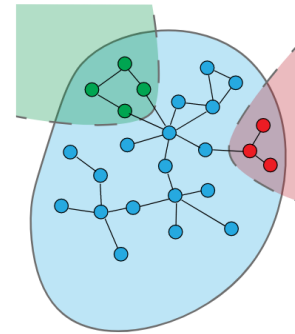
Construct k -NN graph per cell



Recover pure expression profiles using weighted NMF



Label admixture molecules using CRF

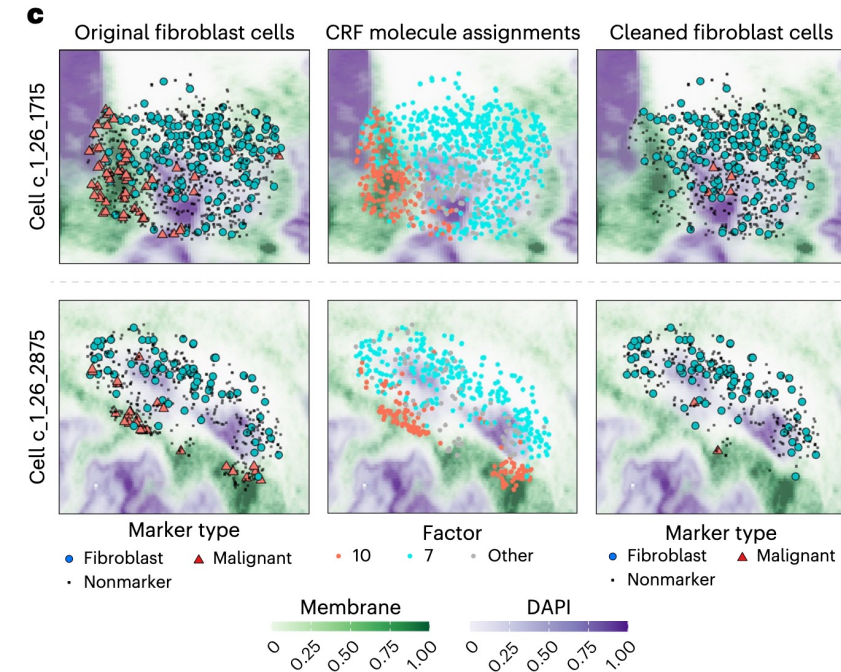
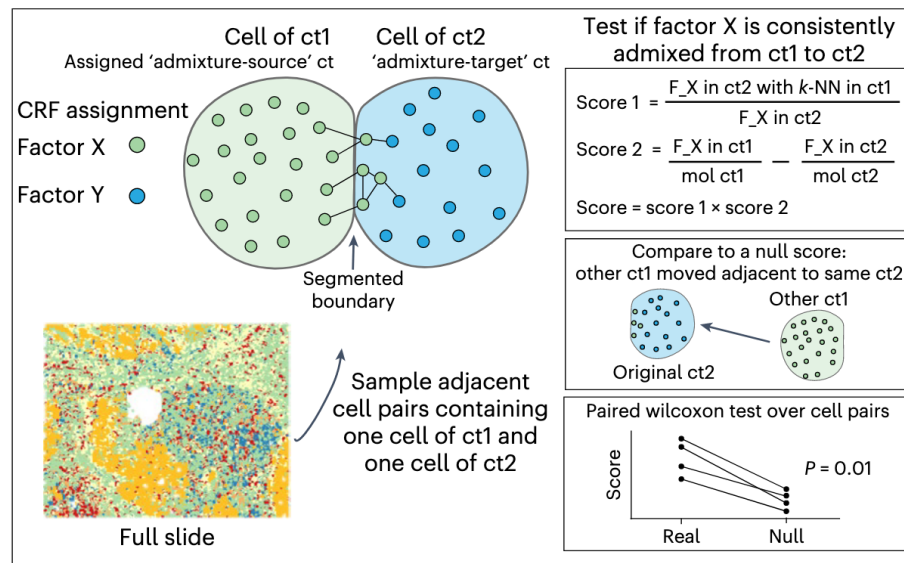


Define admixture score by using Bayesian model to estimate contamination for each gene taking to account cell type adjacency

Compute neighbourhood composition values and factorize with NMF. Conditional random field then assigns individual molecules to the recovered factors.

Via “cell-bridging” molecules whose factor assignment doesn't match their host cell's type are removed

Automated factor annotation using a ‘cell-bridging’ approach



Outline

-
- 1 Intro to the cell segmentation with ST

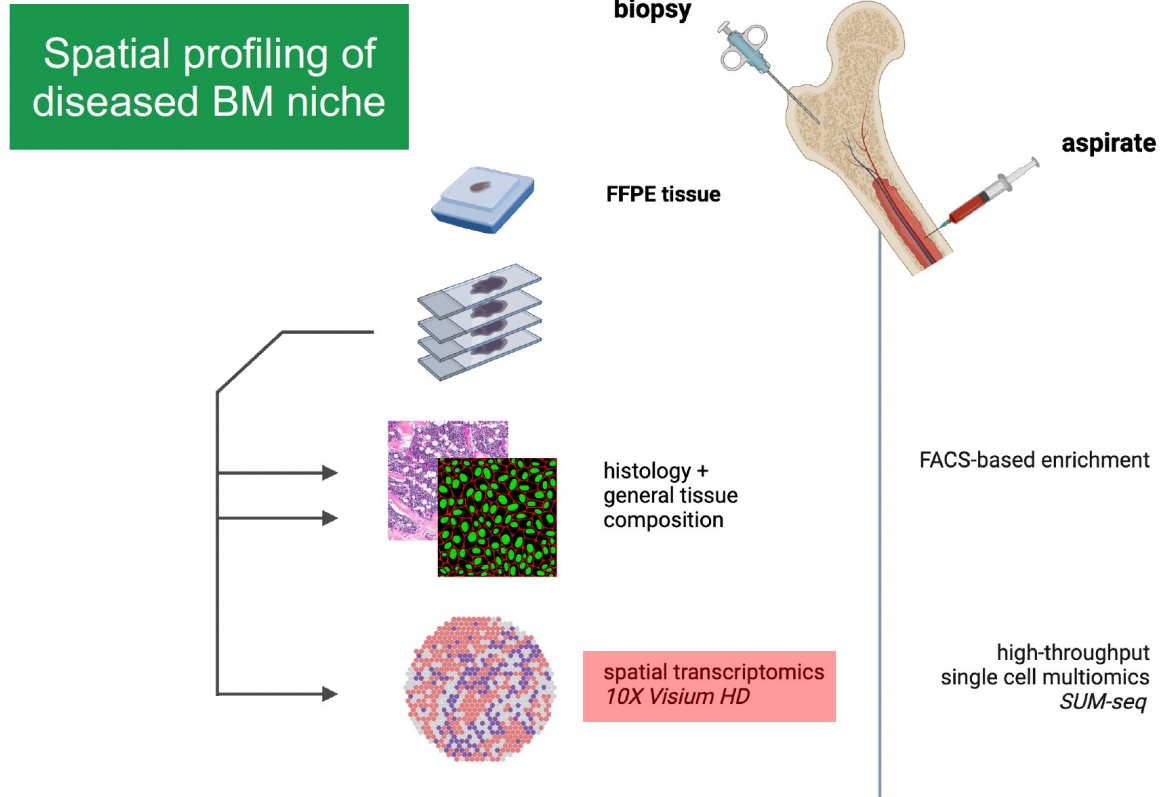
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Project introduction



- Cohort of AML patients (various genetic background):
 - $NPM1_{mut}$ – favorable prognosis
 - $NPM1_{mut}/FLT3ITD_{mut}$ - intermediate
 - $NPM1_{mut}/FLT3ITD_{mut}/DNMT3a_{mut}$ – poor prognosis
 - only $FLT3ITD_{mut}$ – poor prognosis
- Link spatial transcriptomics (FFPE) and multiomics (scATAC/scRNA) sequencing (aspirates) from the same sample
- Focus on the interactions between stromal cells and malignant cells in bone marrow



Universitätsklinikum
Carl Gustav Carus



Patient-derived AML samples used in this analysis

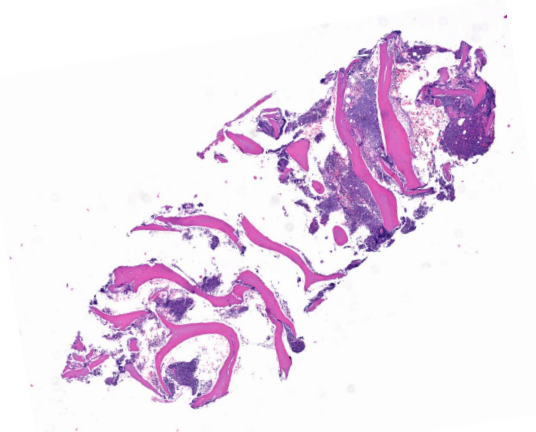
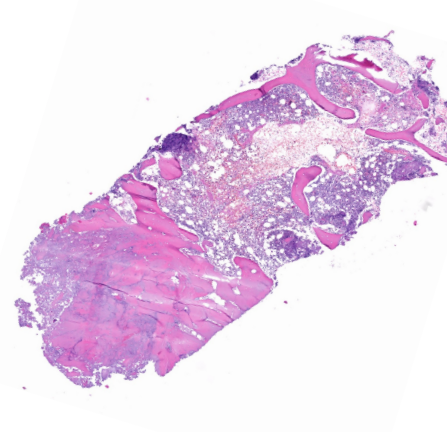
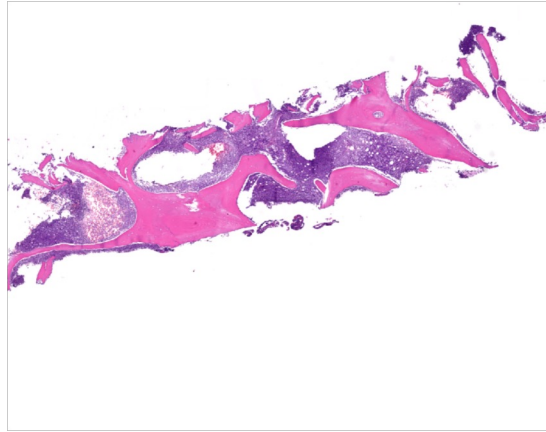
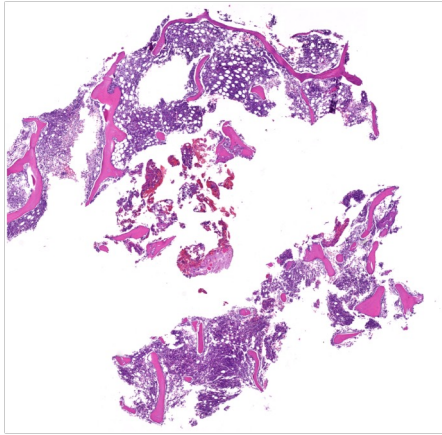
NPM1_{mut}

NPM1_{mut}/FLT3ITD_{mut}/DNMT3A_{mut}

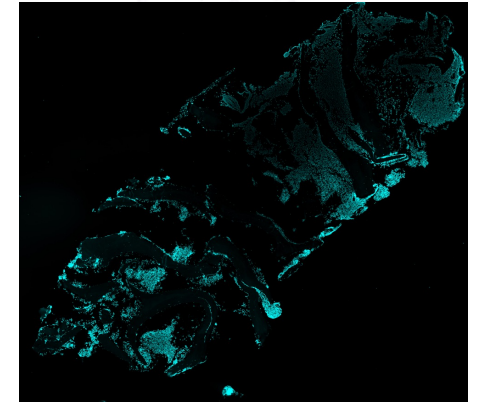
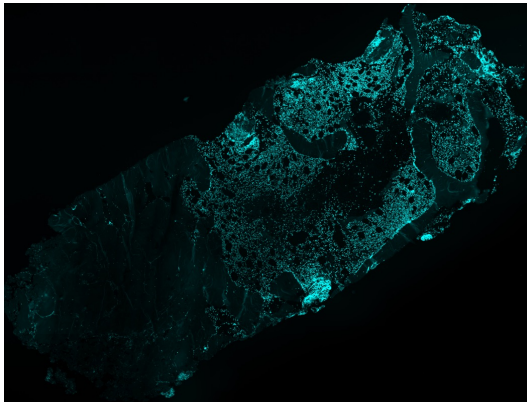
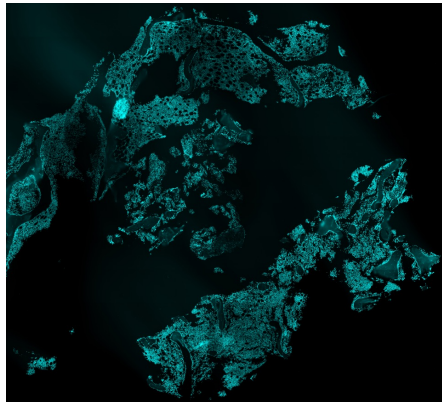
NPM1_{mut}

NPM1_{mut}/FLT3ITD_{mut}

H&E serial



DAPI

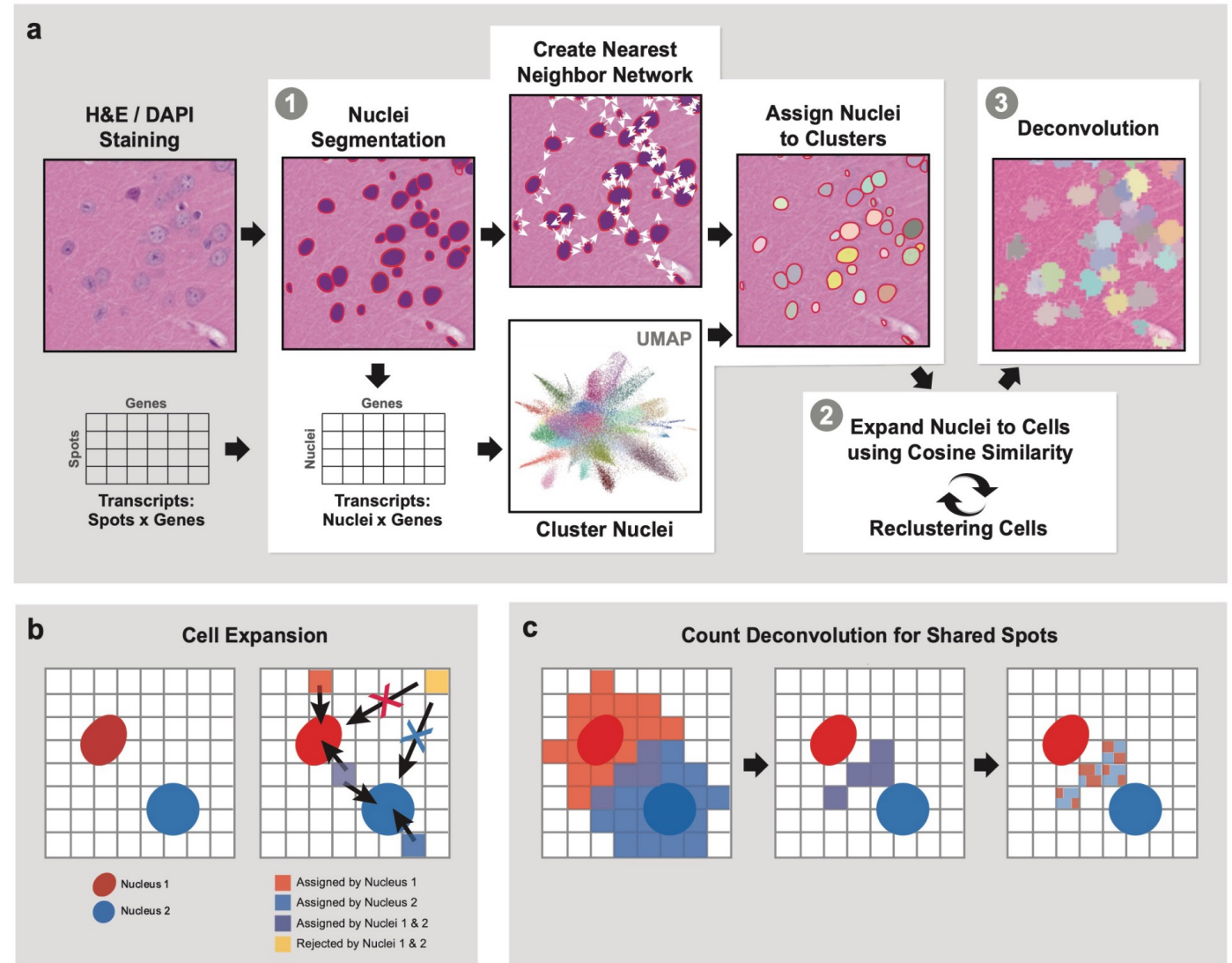
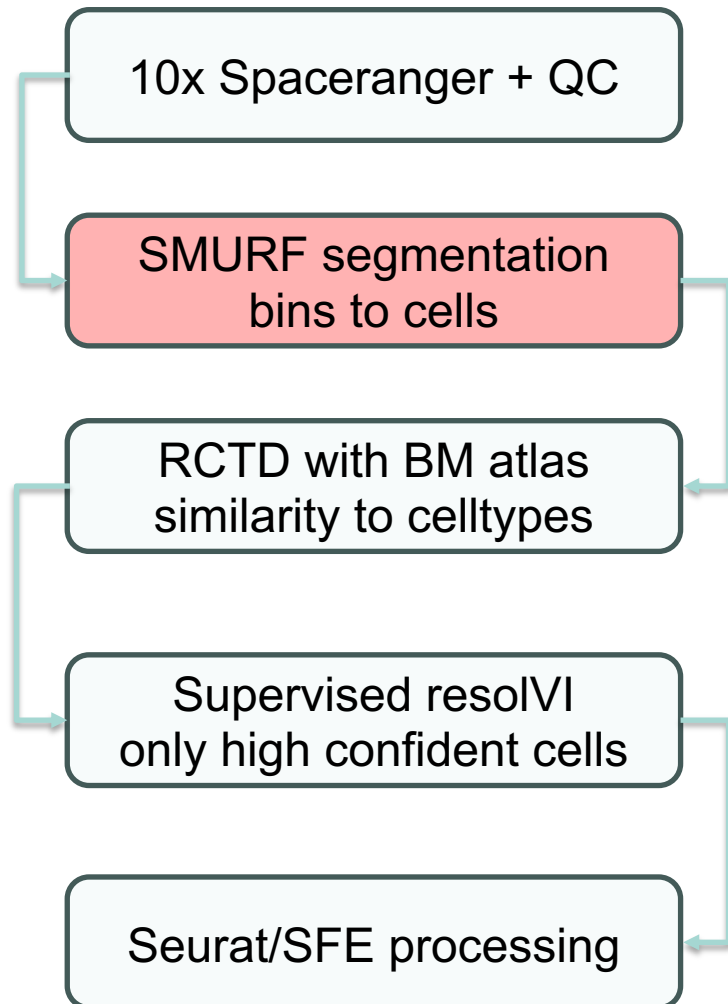


56422

55572

62031

57593





10x Spaceranger + QC

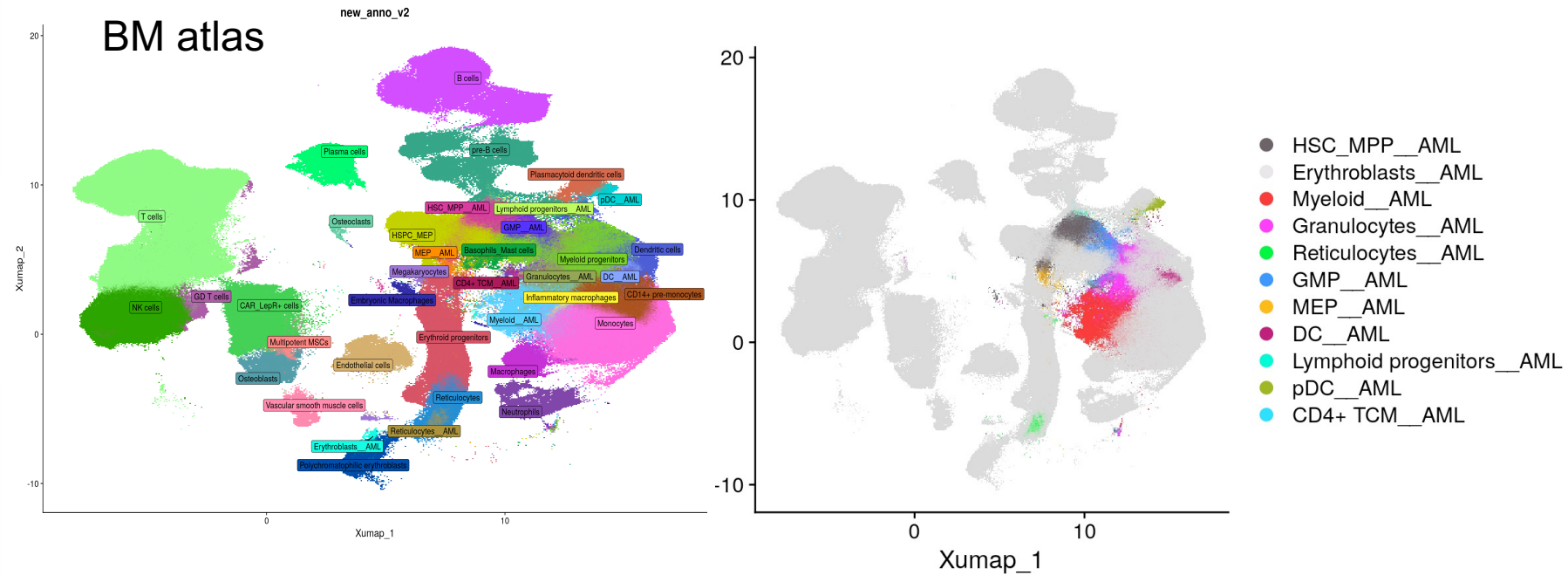
SMURF segmentation
bins to cells

RCTD with BM atlas
similarity to celltypes

Supervised resolVI
only high confident cells

Seurat/SFE processing

Analysis is based on the BM atlas with data from 46 datasets
250 donors with 1.8 million of cells



Using reference atlas transcriptional profiles per celltype to unmix ST data

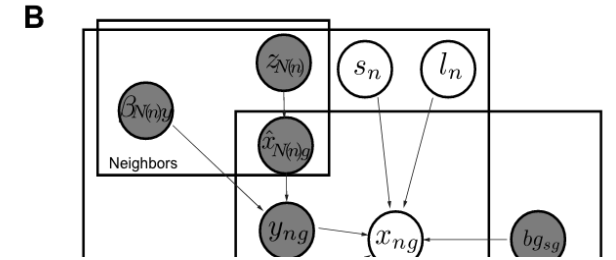
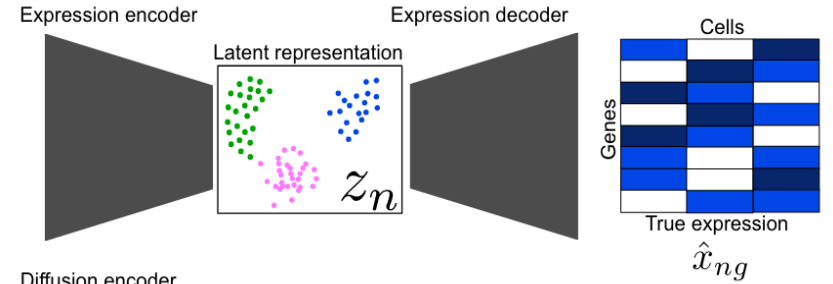
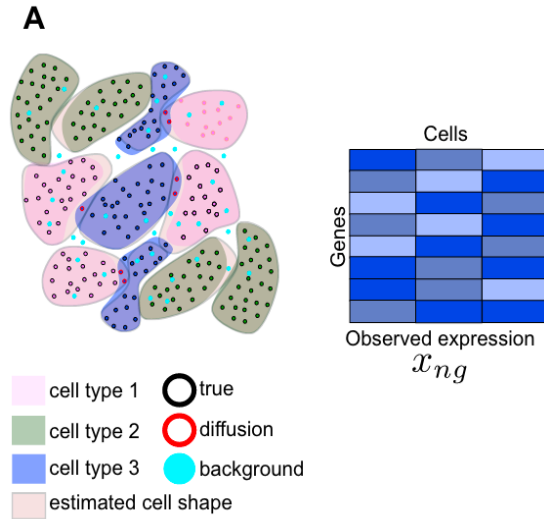
10x Spaceranger + QC

SMURF segmentation
bins to cells

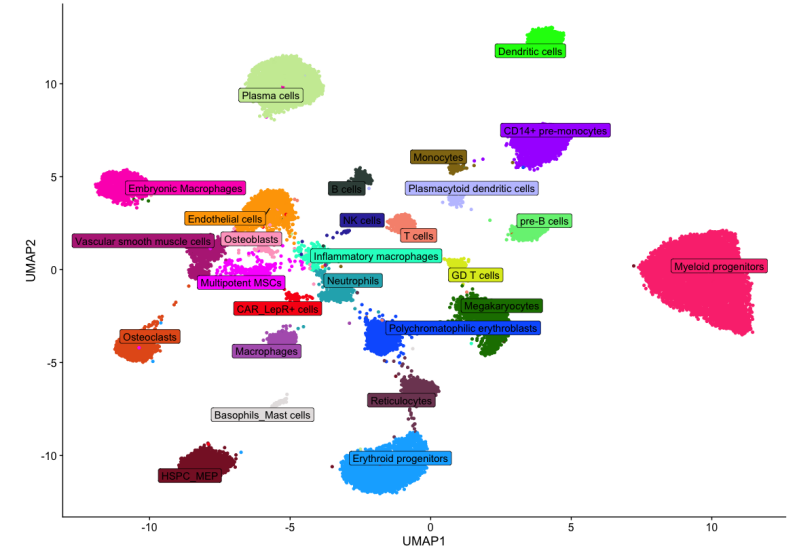
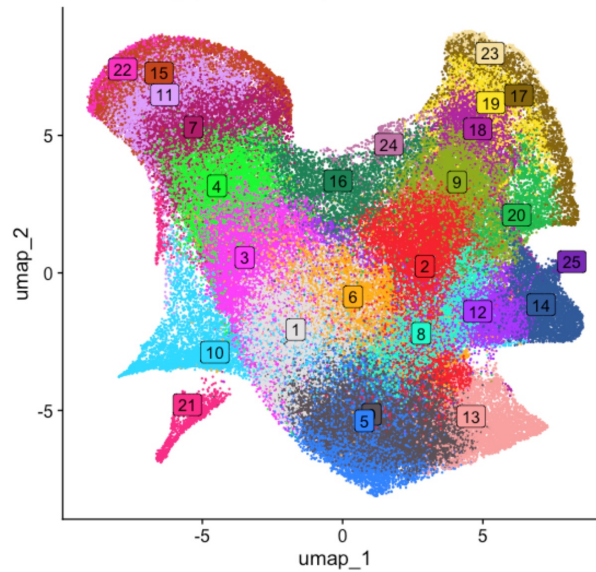
RCTD with BM atlas
similarity to celltypes

Supervised resotVI
only high confident cells

Seurat/SFE processing



Use supervised resotVI with confident set of cells
(primary – secondary > 0.5) ~10-15% of cells



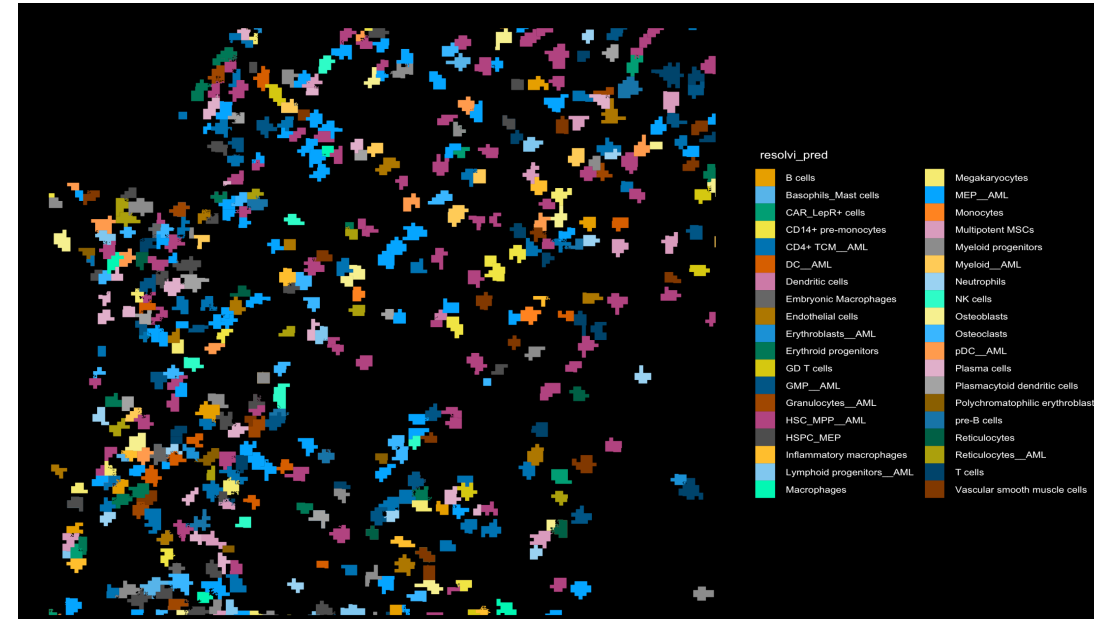
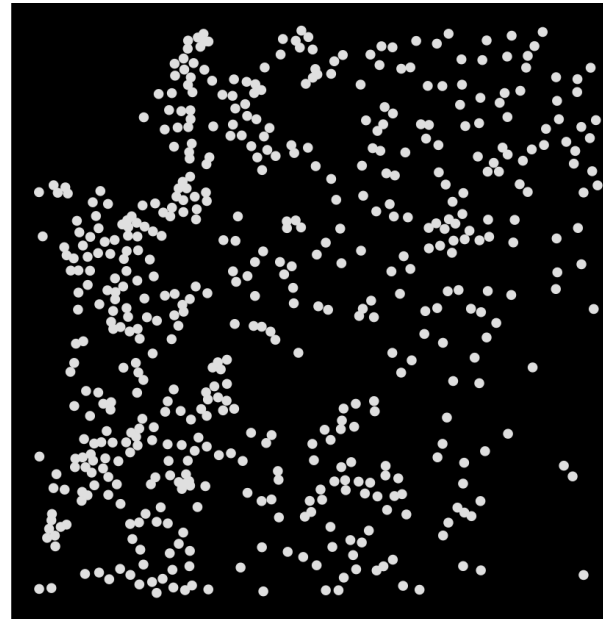
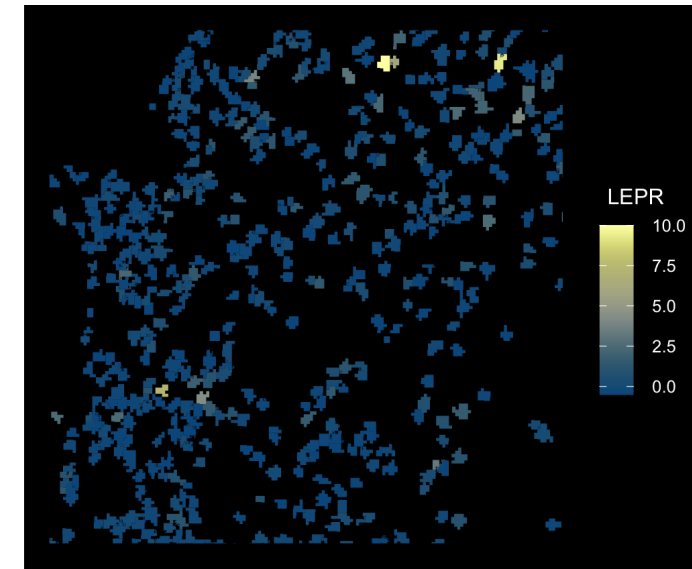
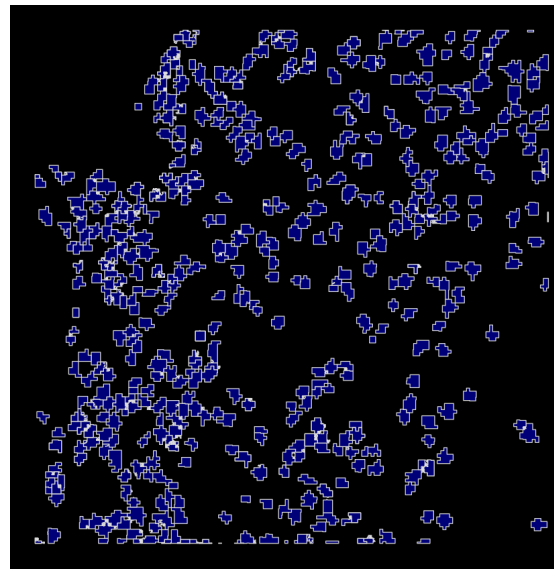
10x Spaceranger + QC

SMURF segmentation
bins to cells

RCTD with BM atlas
similarity to celltypes

Supervised resolVI
only high confident cells

Seurat/SFE processing



Make Seurat/SpatialFeatureExperiment objects for downstream processing

Snakemake Xenium pipeline

Using initial output from Xenium machine followed by resegmentation, additional segmentation with *Proseg* and output in the *SpatialFeatureExperiment* or *spatialdata* format.

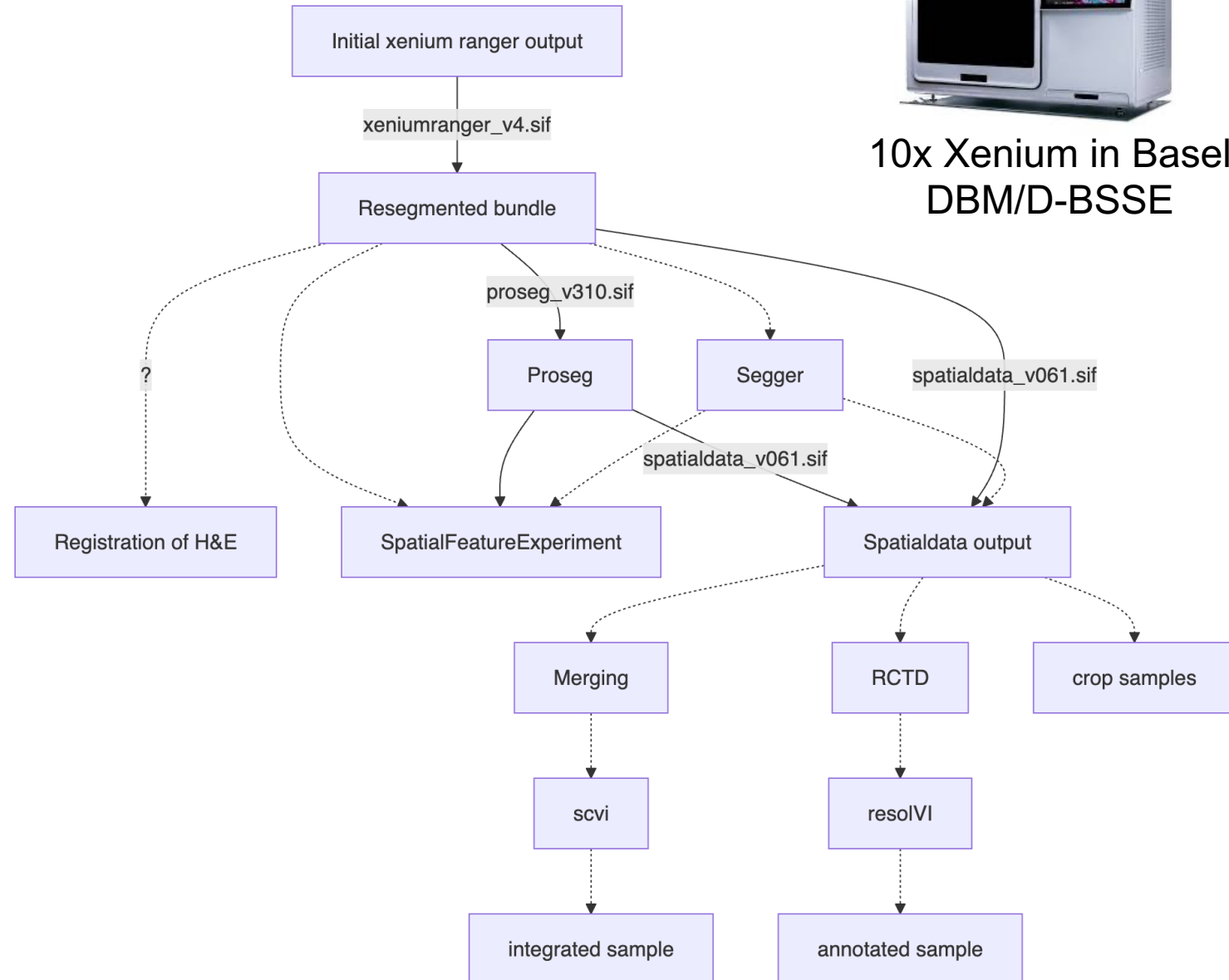
Runtime: ~1day for the whole Xenium slide

Downstream steps:

- RCTD + resolVI annotation based on the reference/matching atlas
- Cropping samples (if not cropped by Xenium, too tight on the slide) based on polygons from Xenium Explorer
- Integrating samples with scvi?
- SPLIT correction and annotation?



10x Xenium in Basel
DBM/D-BSSE



Conclusions

Cell segmentation is **very important** (however almost **never perfect**). Should be done at the very beginning (i.e. part of the Xenium onboarding analysis) and influence all downstream steps.

Comprehensive up-to-now list of sequencing-based and imaging-based spatial transcriptomics cell segmentation tools.

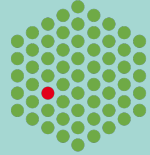
Spatial transcriptomics can enhance just image-based cell segmentation, however introduce certain technology-specific biases (spillover, diffusion).

For the best result you almost always need to combine tools (i.e. SMURF + RCTD + resolVI). Still computational tools lacking interoperability between R and Python ecosystems.



University
of Basel

EMBL



Thank you for your attention.

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