



Statistical methods for spatial omics data

- Overview on the technologies (review)
- Finding spatially-variable genes
- Deconvoluting low-resolution (or aggregating high-resolution) spatial omics data
- Spatially-aware dimension reduction / clustering
- Cell-cell communication —> co-localization
- Classical spatial statistics
 - Point patterns: random, clustered, intensity/correlation
 - Lattice data: useful summaries / functions
 - Models with spatially correlated errors



Slide from Helena Crowell

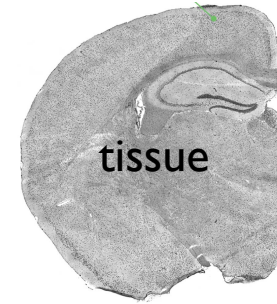
bulk



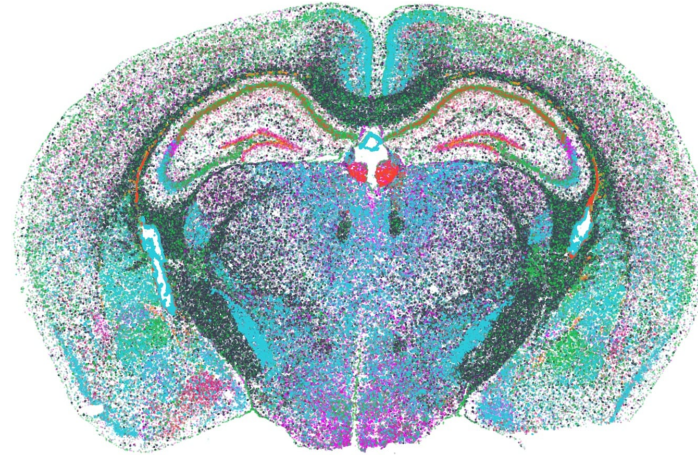
single-cell



spatial

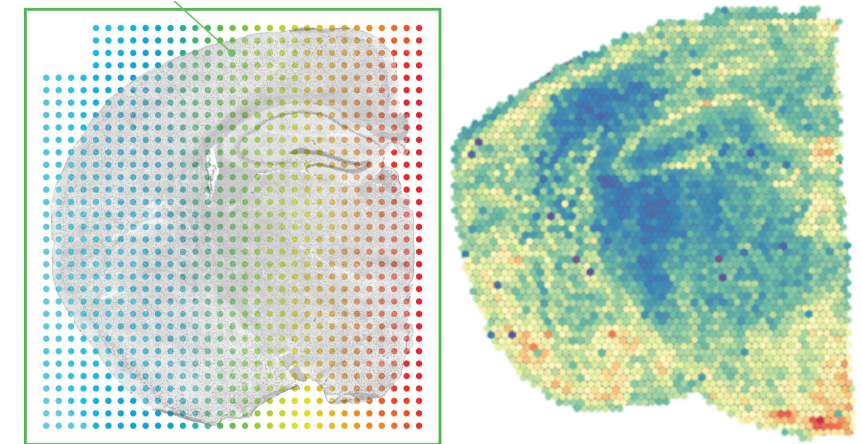


imaging-based



- molecule-level data
- targeted panel (100s of features; >2024: 1000s)
- single-cell resolution requires segmentation

sequencing-based



- spot-level data
- whole transcriptome (10,000s of features)
- single-cell resolutions requires aggregation or deconvolution

Technology choices: expression table + coordinates

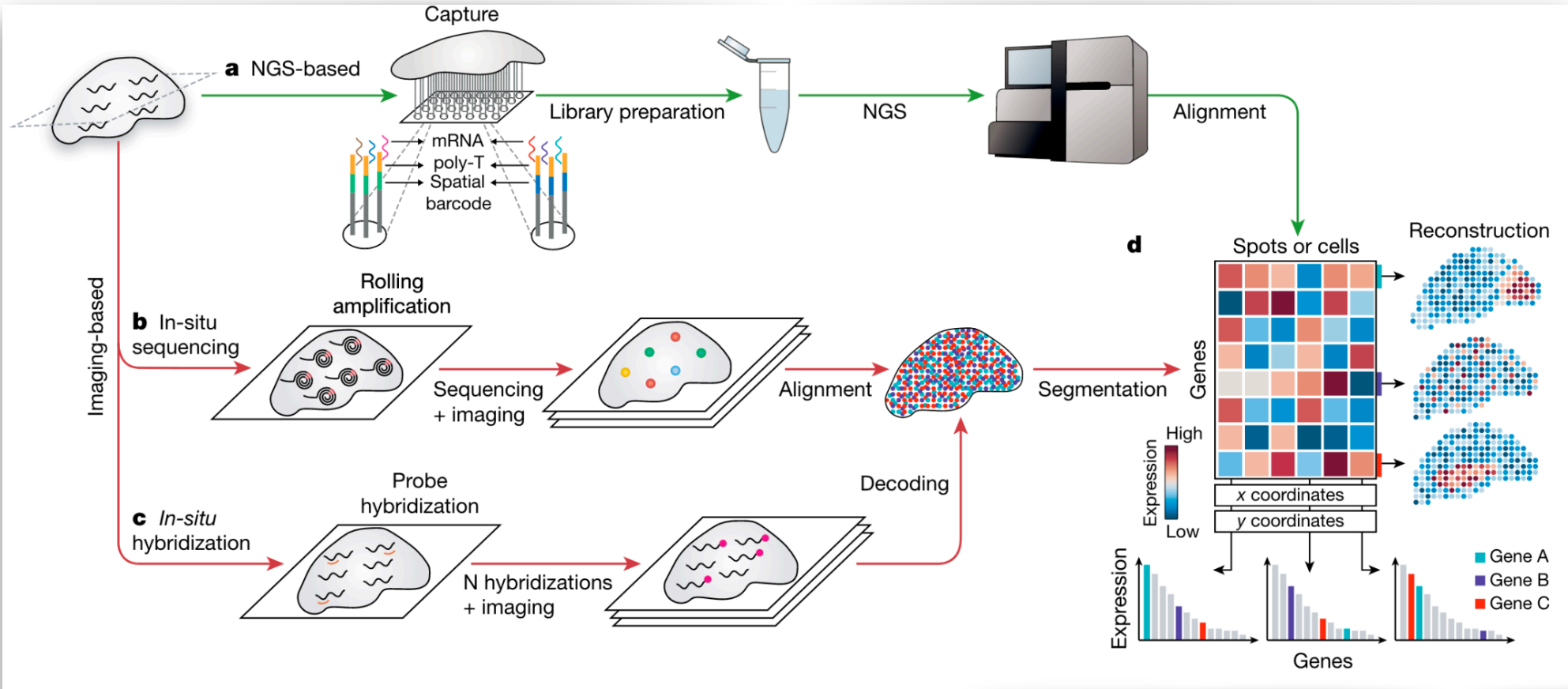
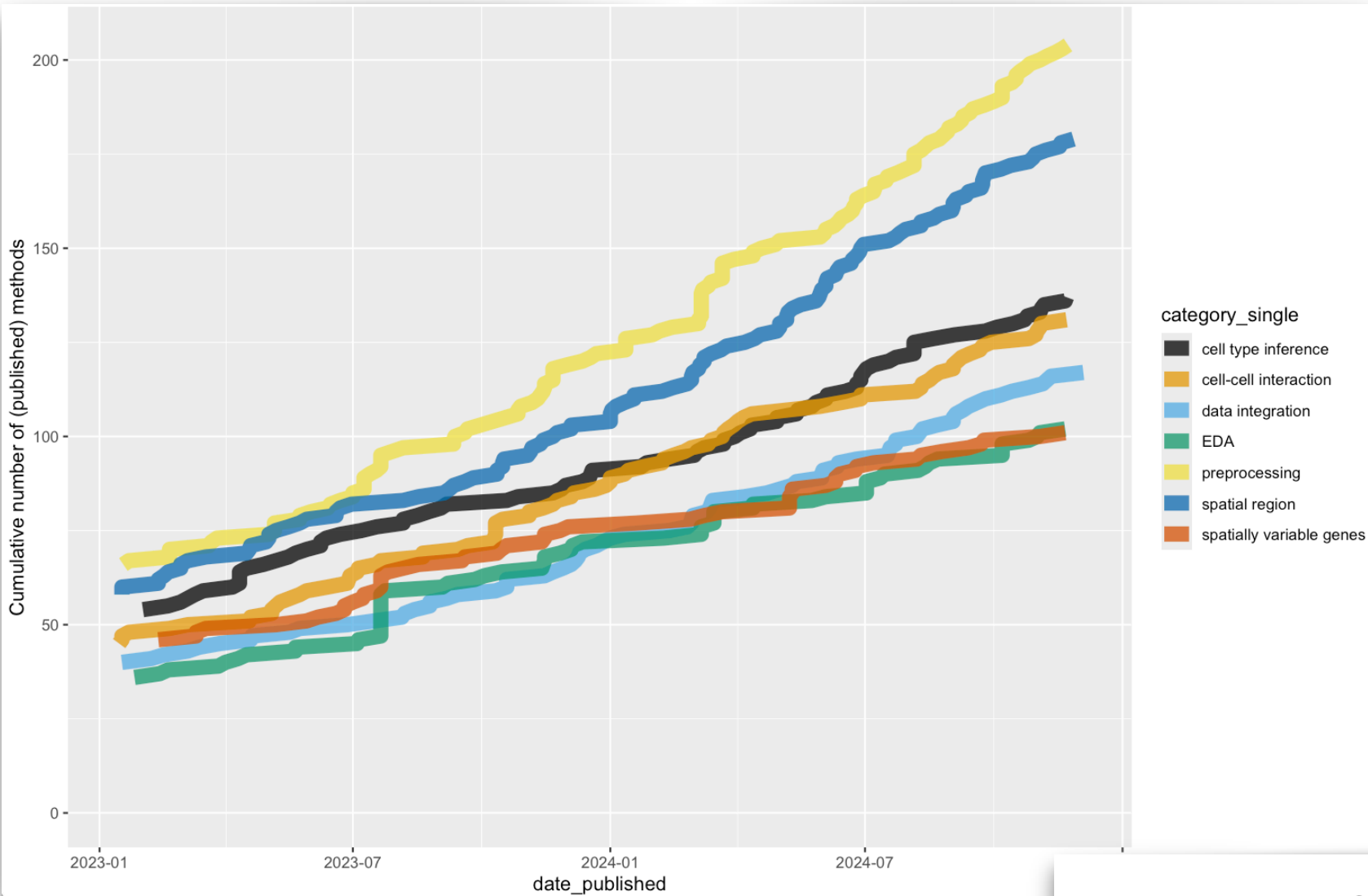


Fig. 1 | The technologies of spatial transcriptomics provide a gene-expression matrix. a, NGS-based spatial transcriptomic methods barcode transcripts according to their location in a lattice of spots. **b,** ISS approaches directly read out the transcript sequence within the tissue. **c,** ISH

methods detect fluorescent probe gene-expression in genes and location

Review

Exploring tissue architecture using spatial transcriptomics



(Spatial omics)
 computational
 method
 explosion —>

—> Google sheet

Museum of spatial transcriptomics

Lambda Moses ¹ and Lior Pachter ^{1,2}



SpatialDE: identification of spatially variable genes

Valentine Svensson^{1,2} , Sarah A Teichmann^{1,3}
& Oliver Stegle^{2,4} 

Finding spatially-variable genes: SpatialDE

- SpatialDE: response = normal distribution with covariance with two components: i) based on distance b/w points - exponential decay; ii) constant non-spatial variance
- Null model: fit just the non-spatial variance (i.e., without sigma)
- Fit 2 models, likelihood ratio test

SpatialDE model. SpatialDE models gene expression profiles $y = (y_1, \dots, y_N)$ for a given gene across spatial coordinates $X = (x_1, \dots, x_N)$, using a multivariate normal model of the form

$$P(y | \mu, \sigma_s^2, \delta, \Sigma) = N(y | \mu \cdot \mathbf{1}, \sigma_s^2 \cdot (\Sigma + \delta \cdot I)) \quad (1)$$

The fixed effect $\mu_g \cdot \mathbf{1}$ accounts for the mean expression level, and Σ denotes a spatial covariance matrix defined on the basis of the input coordinates of pairs of cells. SpatialDE uses the so-called squared exponential covariance function to define Σ :

$$\Sigma_{i,j} = k(x_i, x_j) = \exp\left(-\frac{|x_i - x_j|^2}{2 \cdot l^2}\right) \quad (2)$$

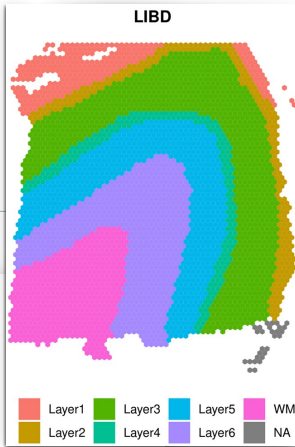


nnSVG for the scalable identification of spatially variable genes using nearest-neighbor Gaussian processes

Received: 15 June 2022

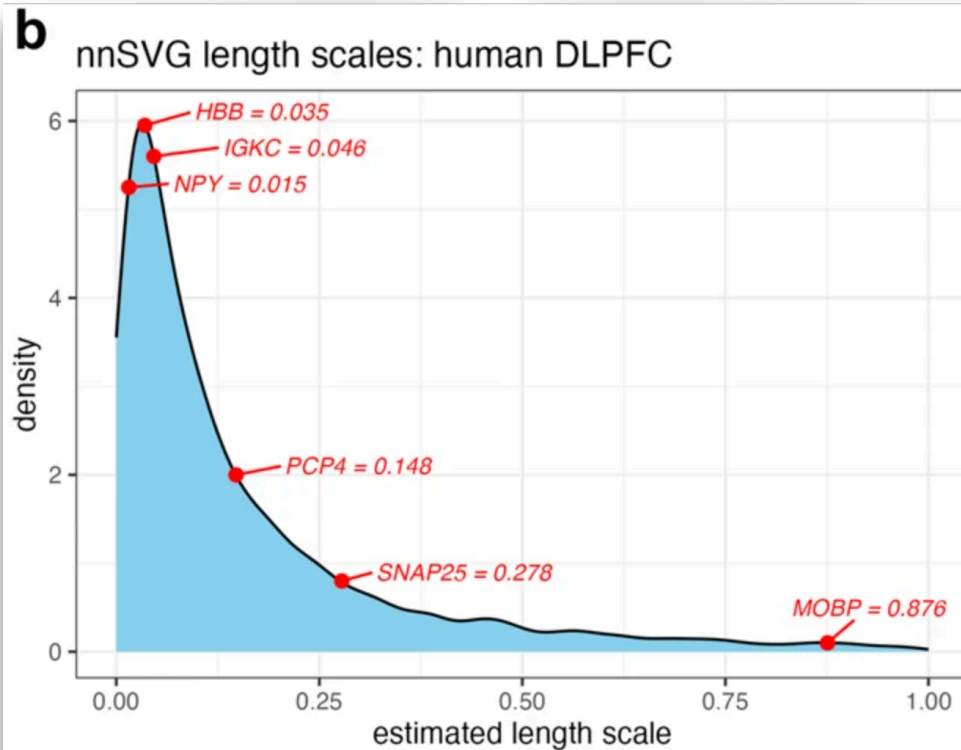
Lukas M. Weber¹, Arkajyoti Saha², Abhirup Datta¹, Kasper D. Hansen¹ & Stephanie C. Hicks¹✉

Accepted: 23 June 2023

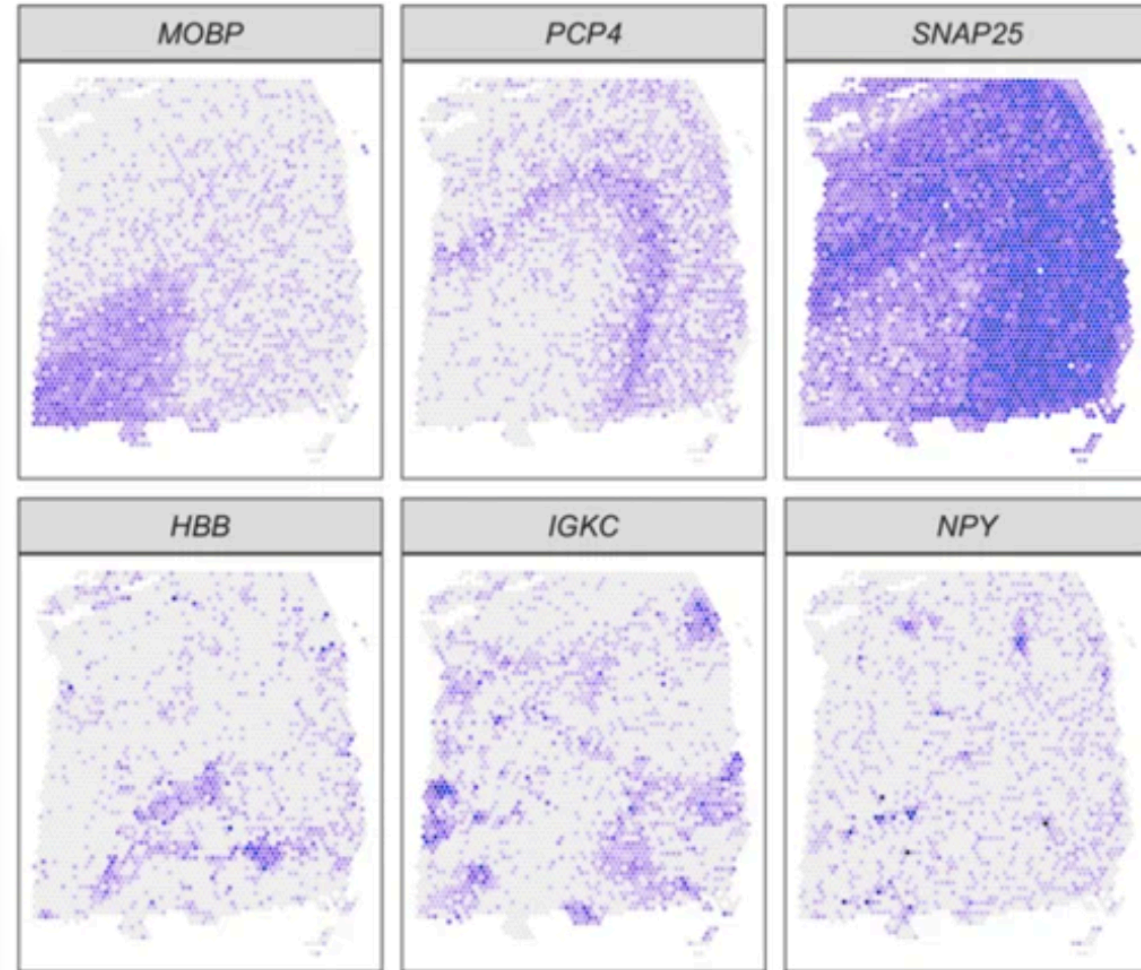


Spatially variable genes → scale

$$C_{ij}(\theta) = \sigma^2 \exp\left(\frac{-\|s_i - s_j\|}{l}\right)$$



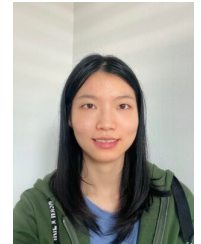
Selected SVGs: human DLPFC



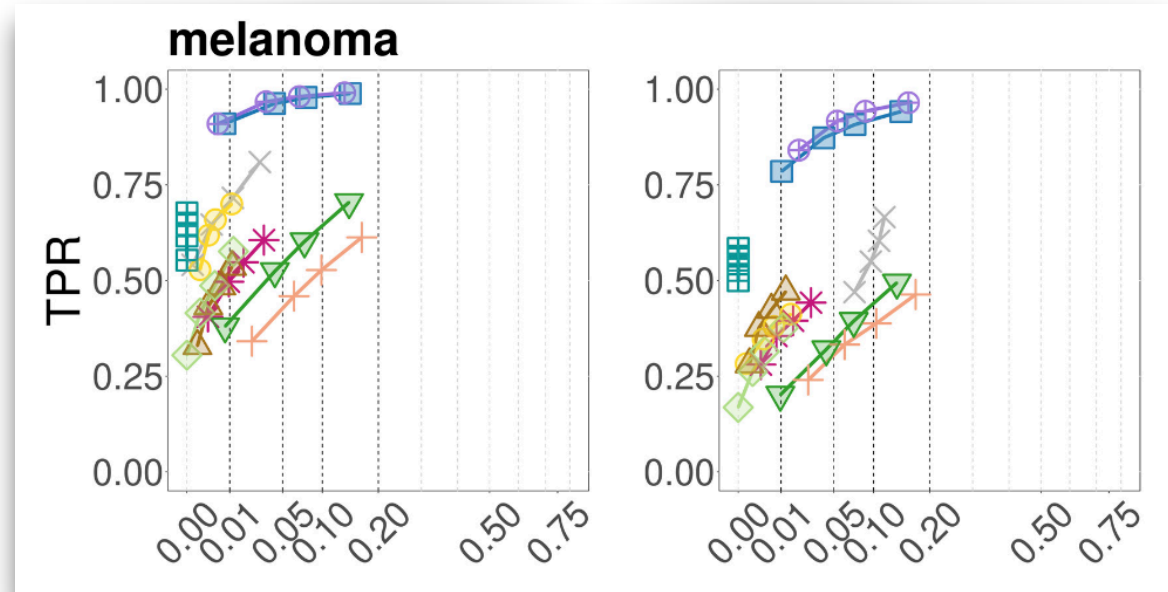
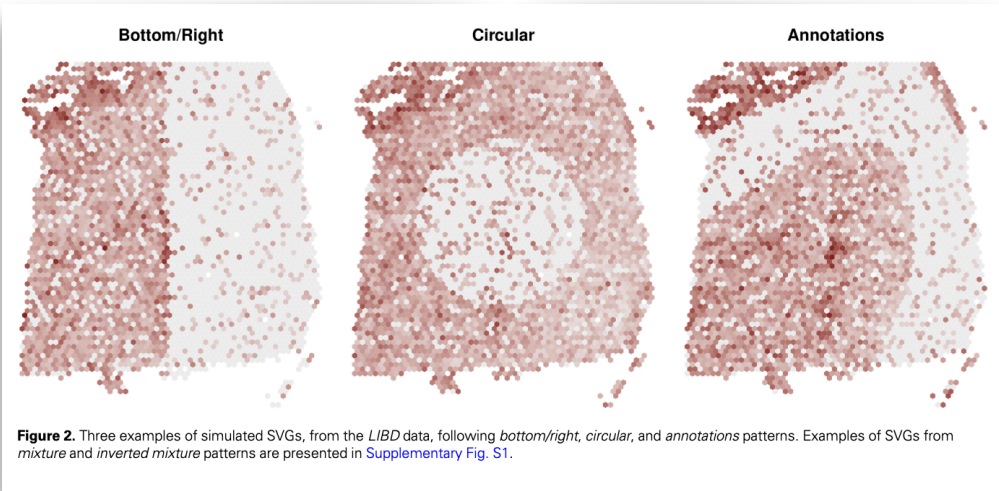
Alternatively, spatially variable features = DE between domains



Simone Tiberi



Peiying Cai



To find spatially variable genes (SVGs); spatial clustering + classical statistical method works quite well

- BayesSpace_DESpace
- ▲ SPARK-X
- + MERINGUE
- BayesSpace_findMarkers
- StLearn_DESpace
- ▼ SpatialDE
- * SpaGCN
- StLearn_findMarkers
- StLearn_FindAllMarkers
- SPARK
- ▼ SpatialDE2
- * nnSVG
- BayesSpace_FindAllMarkers

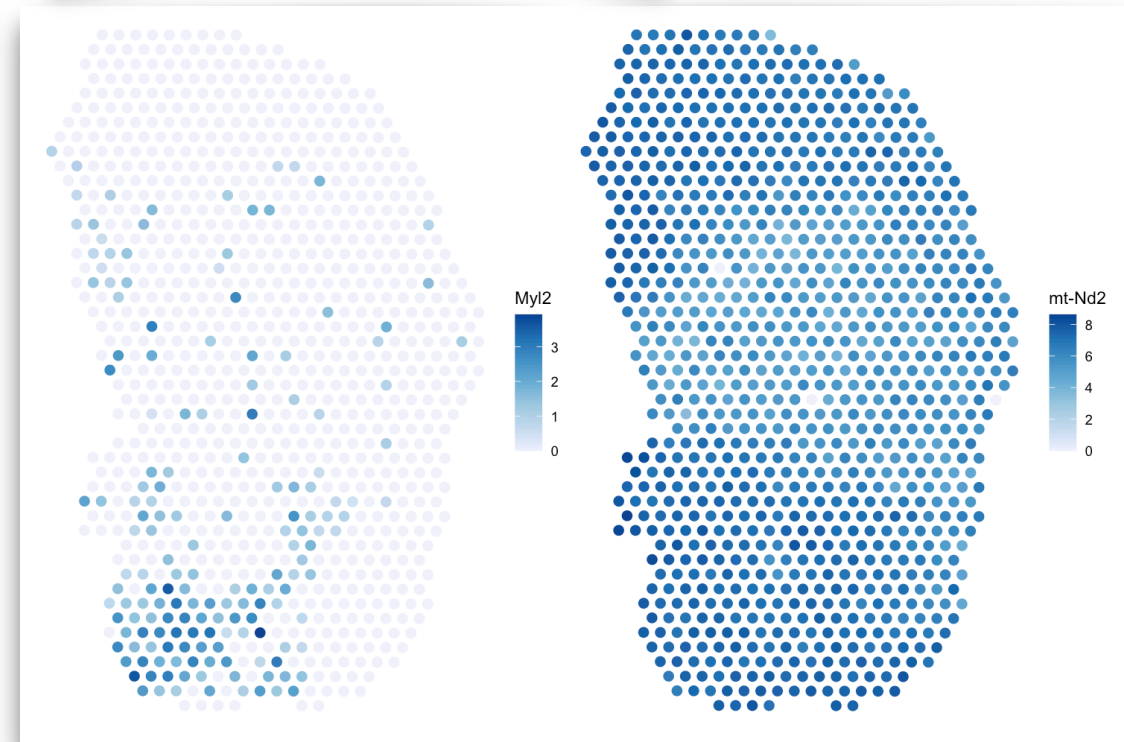
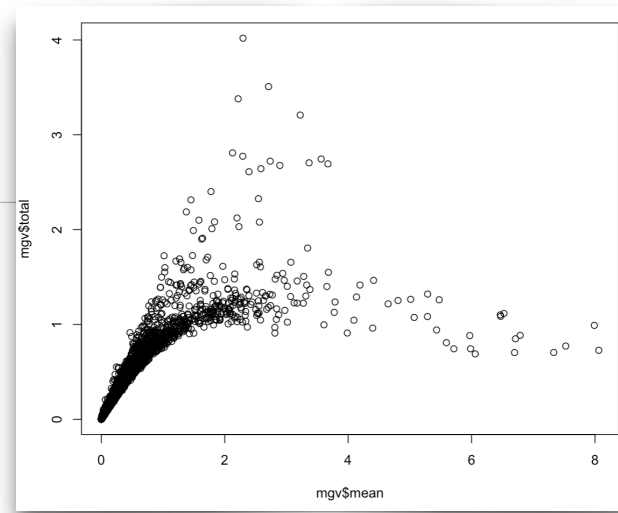
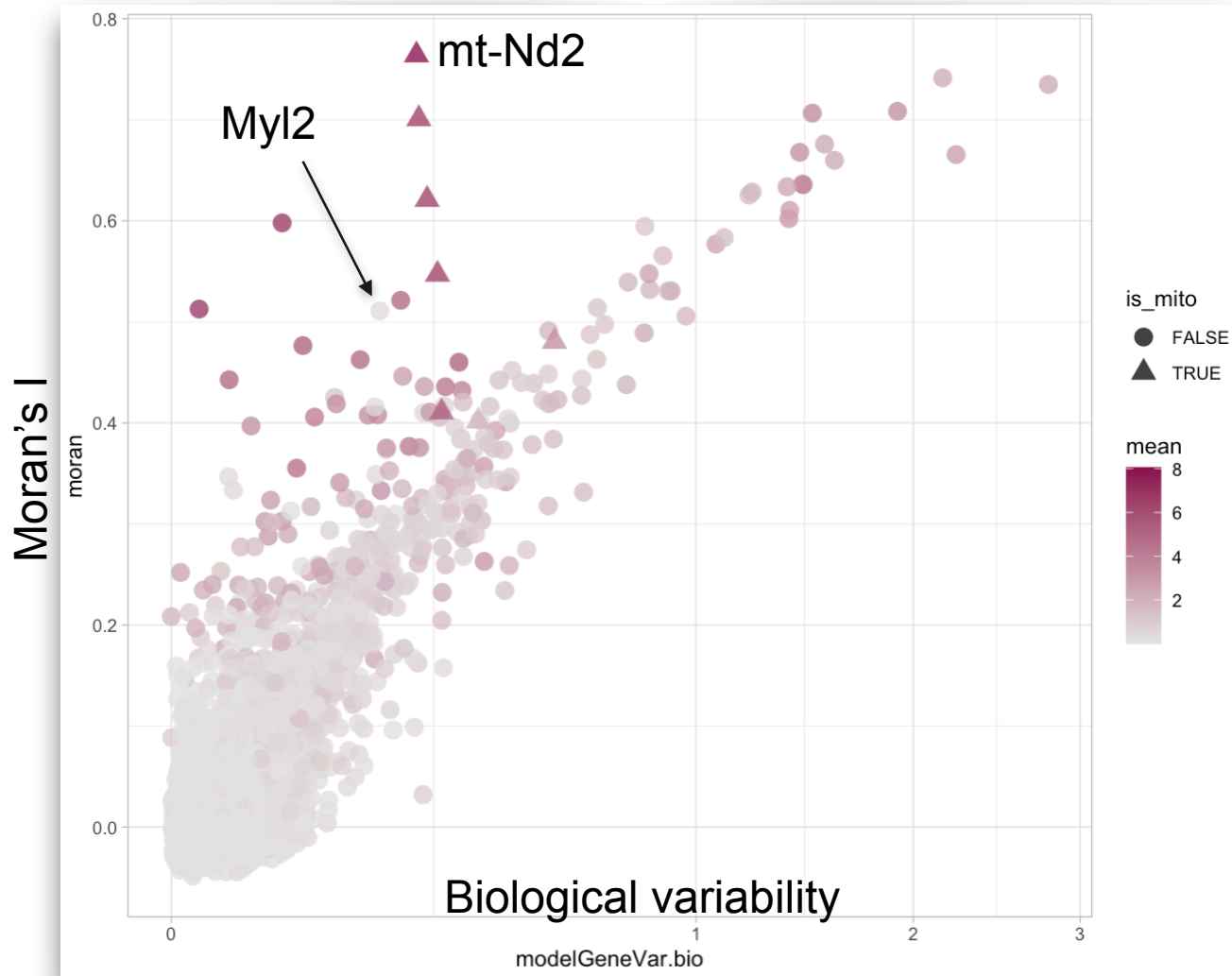
JOURNAL ARTICLE

***DESpace*: spatially variable gene detection via differential expression testing of spatial clusters**

Peiying Cai, Mark D Robinson, Simone Tiberi



Spatially variable versus highly variable



(More mathematical details on Moran's I below)

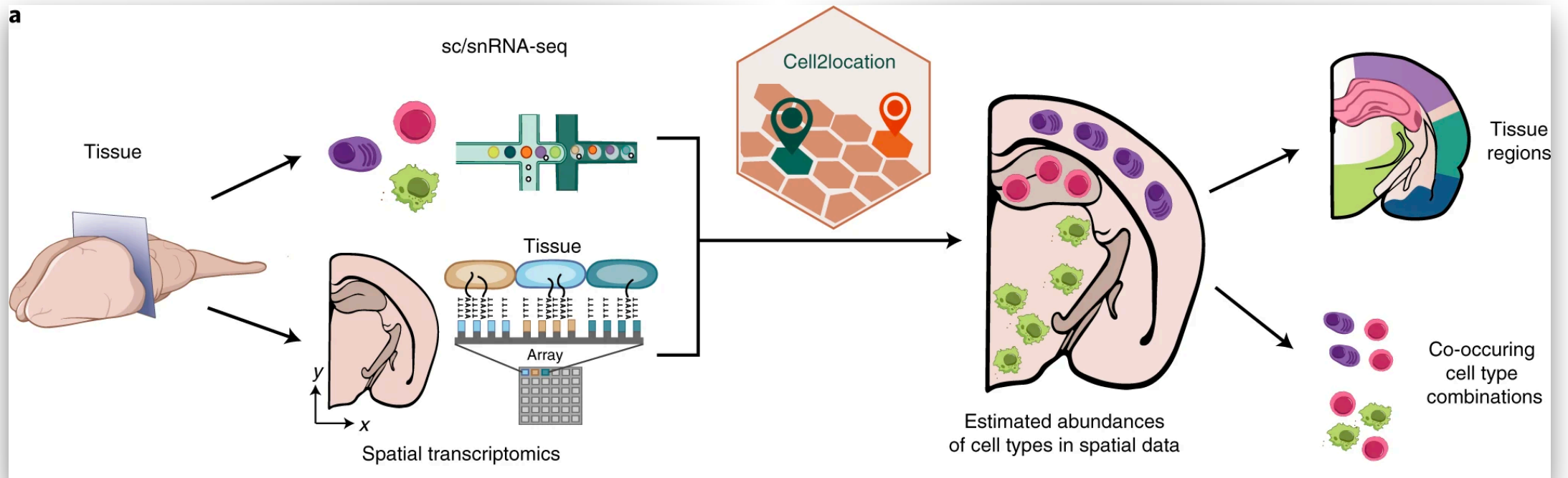


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Deconvoluting low-resolution spatial omics (sequencing) data

- Cell2location: negative binomial regression for reference cell type signatures; decompose spot-level mRNA counts into reference cell types





Deconvoluting low-resolution spatial omics data

- Cell2location: negative binomial regression for reference cell type signatures; decompose spot-level mRNA counts into reference cell types

Cell2location model. Cell2location models the elements of the spatial expression count matrix $d_{s,g}$ as negative binomial distributed, given an unobserved gene expression level (rate) $\mu_{s,g}$ and gene- and batch-specific over-dispersion $\alpha_{e,g}$:

$$d_{s,g} \sim NB(\mu_{s,g}, \alpha_{e,g}).$$

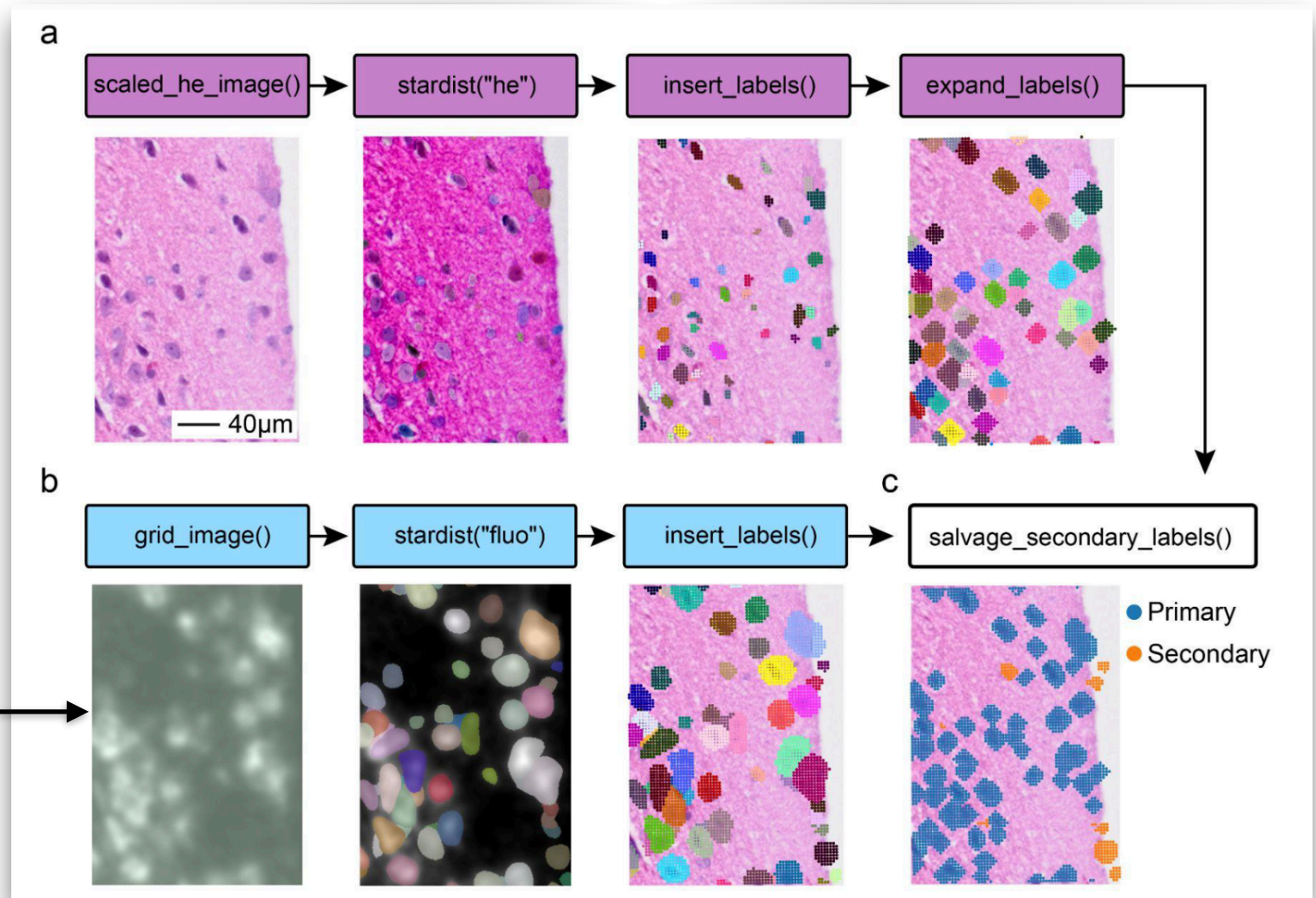
The expression rate of genes g at location s , $\mu_{s,g}$ in the mRNA count space is modeled as a linear function of reference cell types signatures $g_{f,g}$:

$$\mu_{s,g} = \left(\underbrace{m_g}_{\text{technology sensitivity}} \cdot \underbrace{\sum_f w_{s,f} g_{f,g}}_{\text{cell type contributions}} + \underbrace{s_{e,g}}_{\text{additive shift}} \right) \cdot \underbrace{\gamma_s}_{\text{per-location sensitivity}}.$$

Aggregating high-resolution spatial omics (sequencing) data

- bin2cell: combines segmentation on H&E/IF and segmentation on gene expression counts

Image of counts per spot (smoothed)





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BANKSY unifies cell typing and tissue domain segmentation for scalable spatial omics data analysis

Received: 3 April 2023

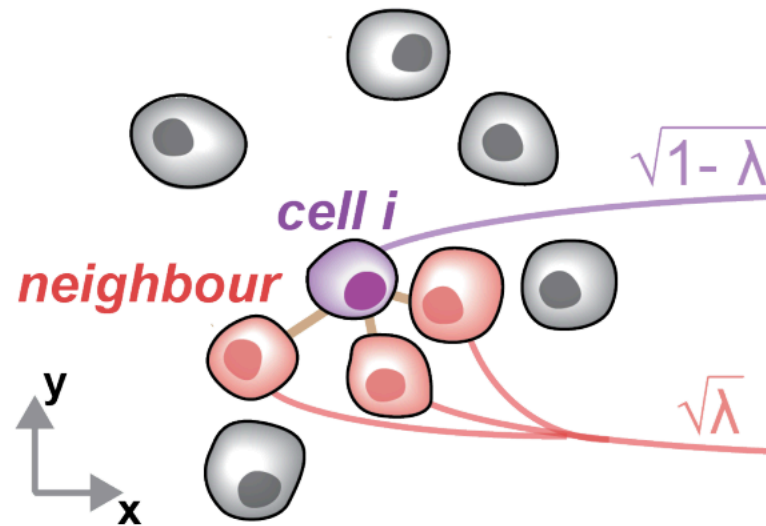
Accepted: 16 January 2024

Published online: 27 February 2024

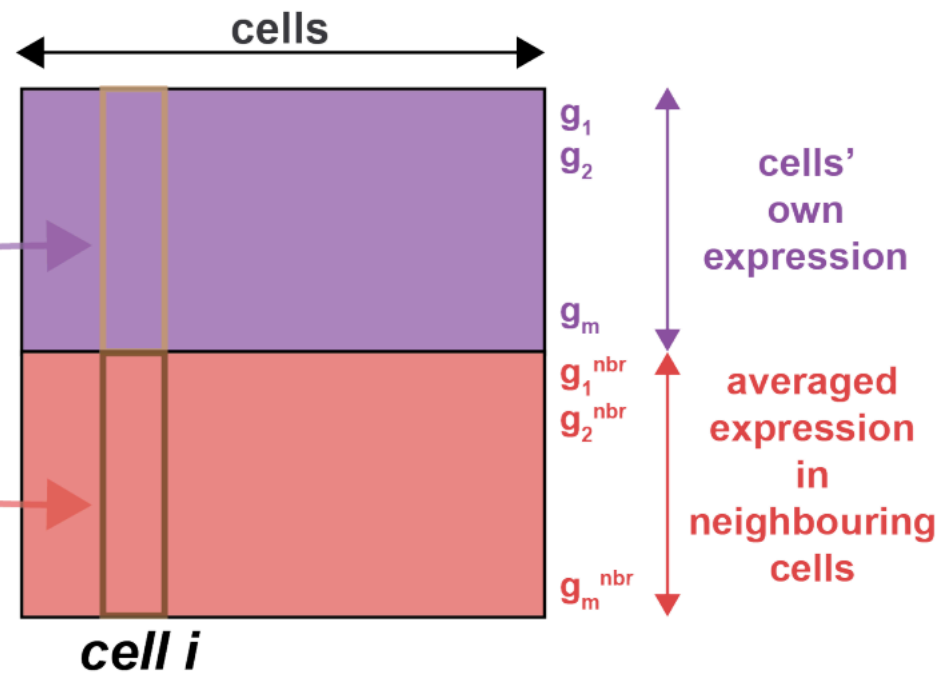
Vipul Singhal^{1,13}, Nigel Chou^{1,13}, Joseph Lee², Yifei Yue³, Jinyue Liu¹, Wan Kee Chock¹, Li Lin⁴, Yun-Ching Chang⁵, Erica Mei Ling Teo⁵, Jonathan Aow¹, Hwee Kuan Lee^{4,6,7,8,9,10}, Kok Hao Chen¹✉ & Shyam Prabhakar^{1,11,12}✉

Spatial clustering / domain detection (BANKSY)
→ combine transcription and spatial information

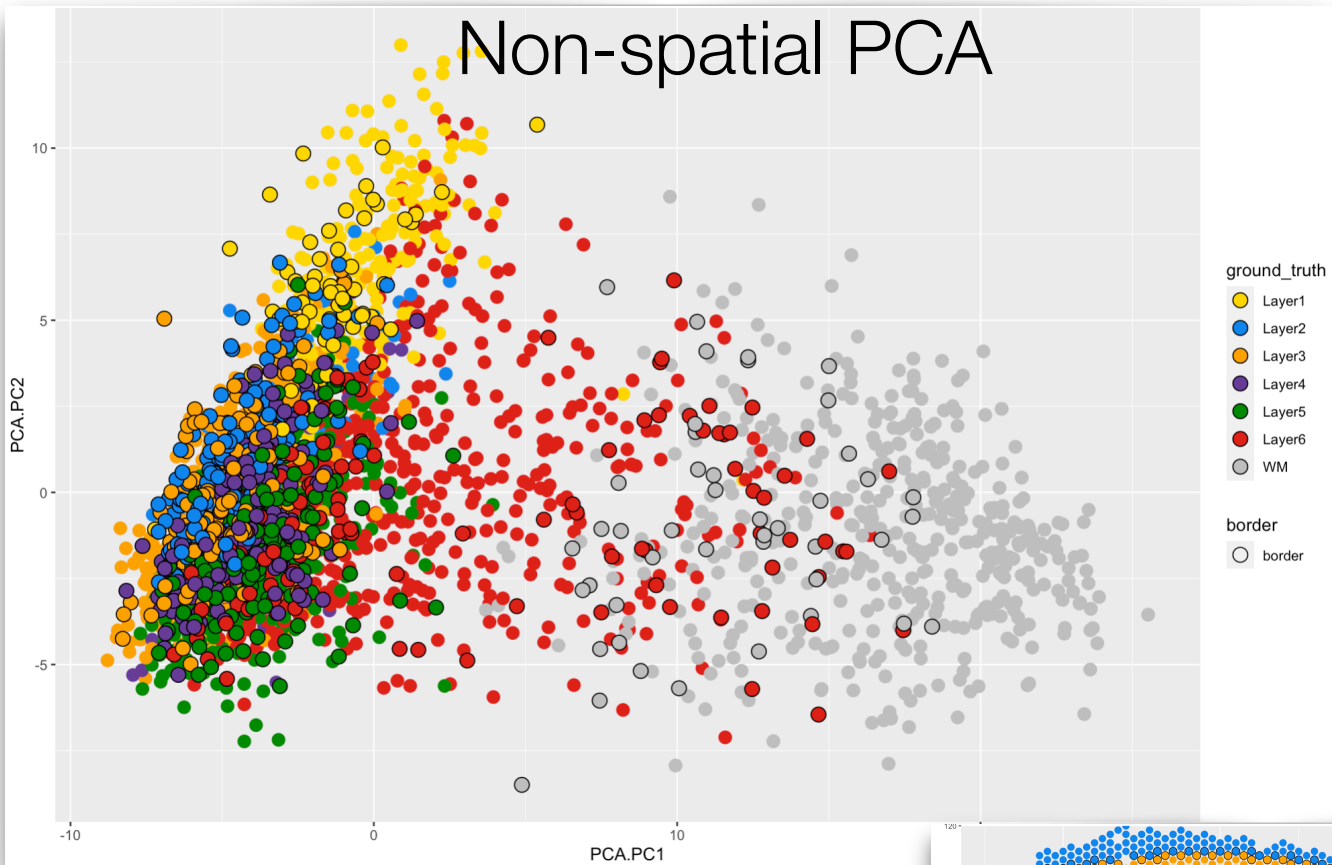
a Cells in physical space



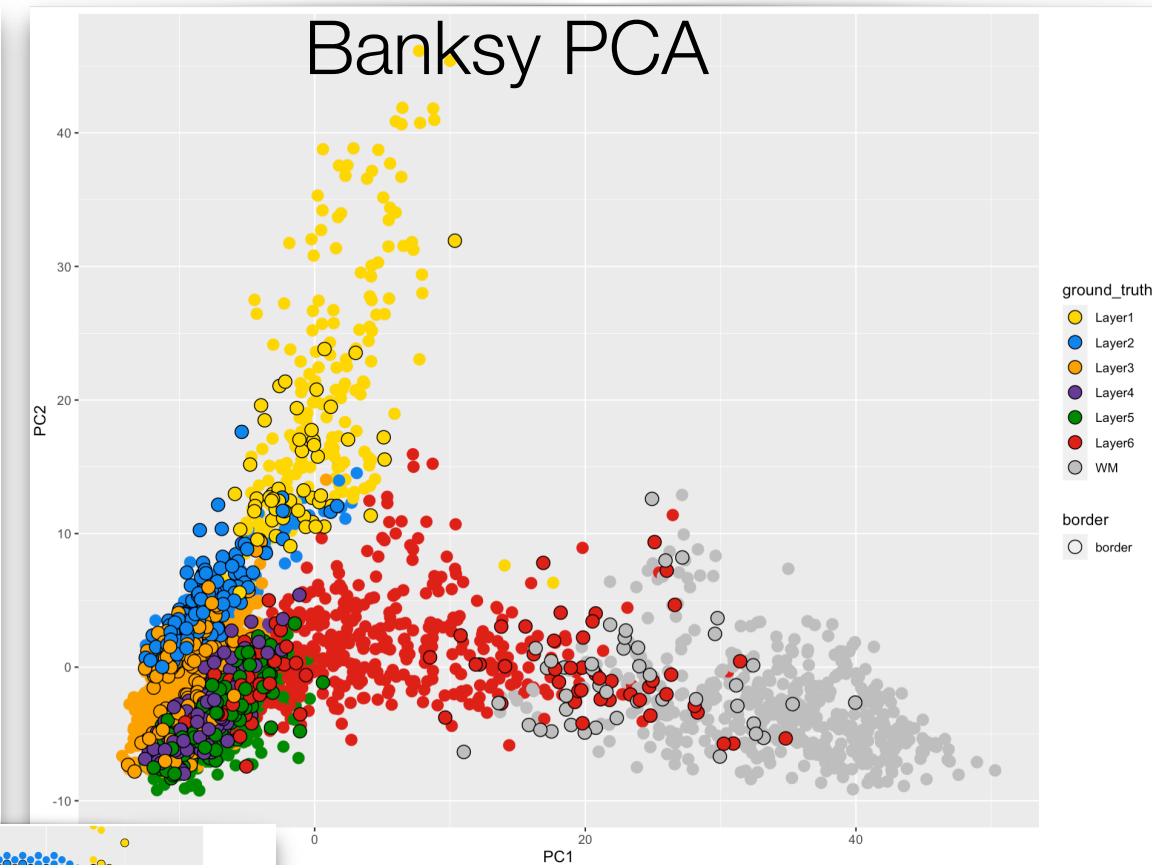
Neighbour-augmented expression matrix



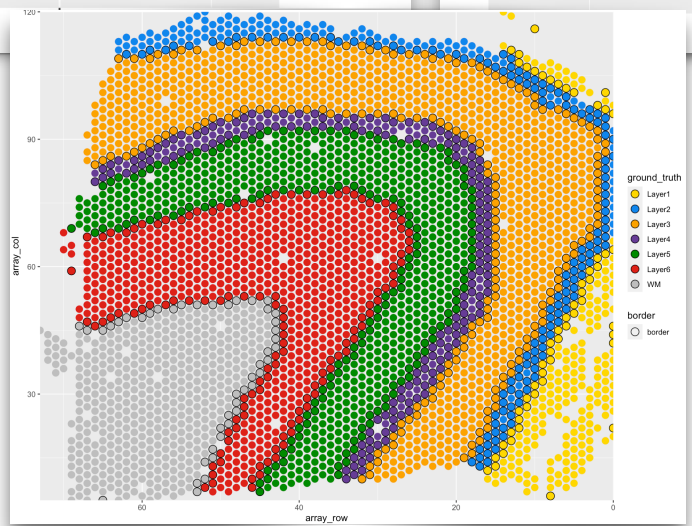
Non-spatial PCA



Banksy PCA



Sample 151673



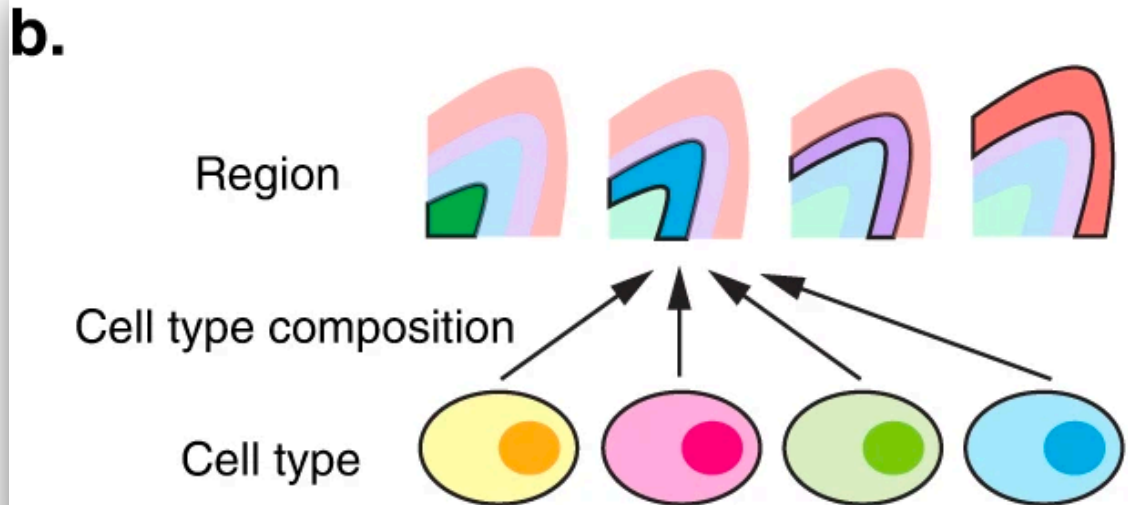






Spatially aware dimension reduction for spatial transcriptomics

Received: 10 March 2022

Lulu Shang ^{1,2} & Xiang Zhou ^{1,2} ✉

Spatial domain detection ~ spatially homogeneous regions ~ spatial niches
—> needs a definition!

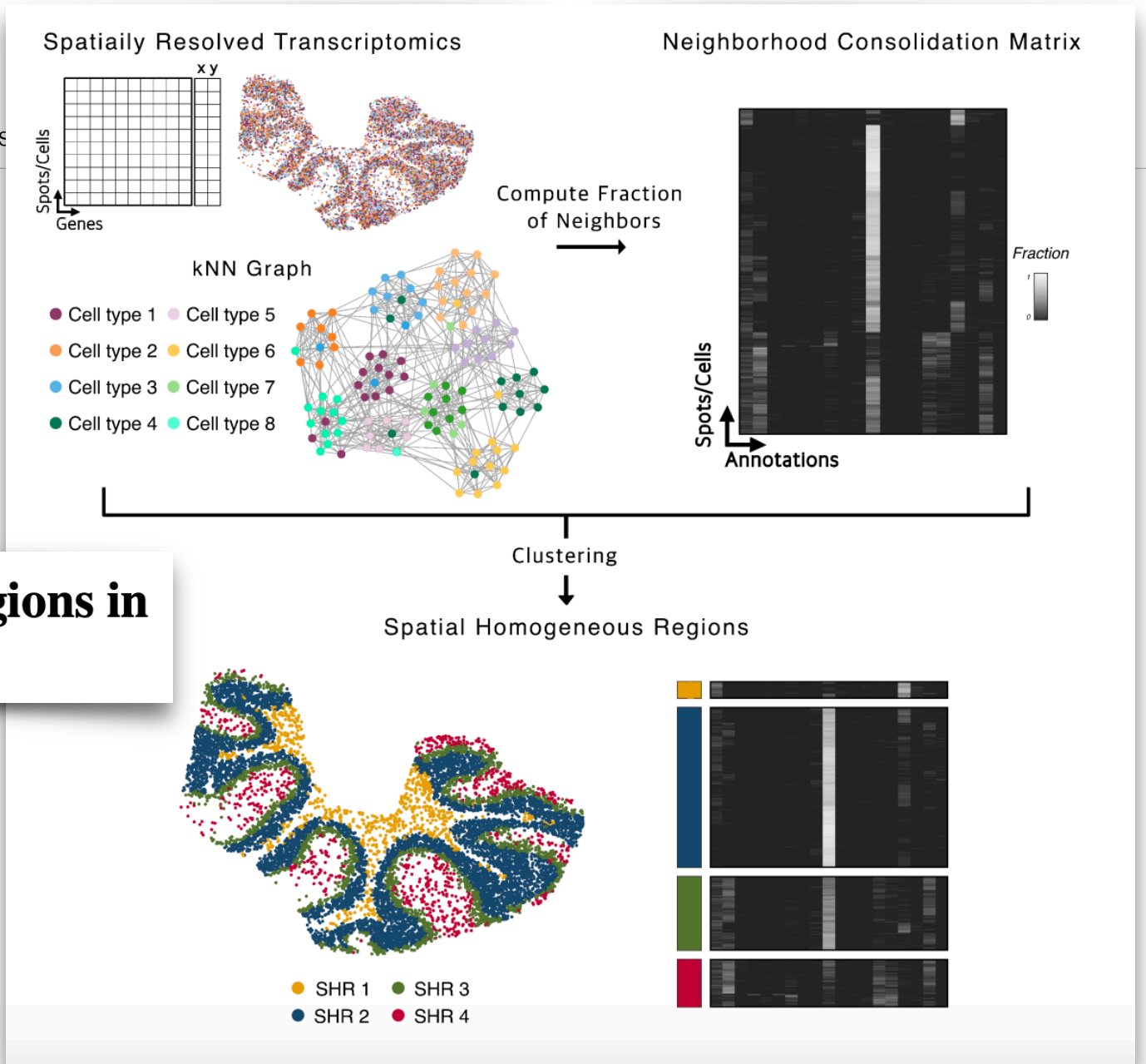


				
Scenario 1	70%	10%	10%	10%
Scenario 2	45%	45%	5%	5%
Scenario 3	60%	30%	5%	5%
Scenario 4	35%	30%	30%	5%



Spatial domain detection ~ spatially homogeneous regions

Identification of spatial homogeneous regions in tissues with concordex





Statistical methods for spatial omics data

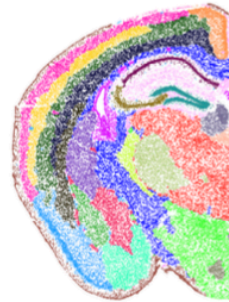
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pasta: Data representations determine spatial statistics options

A

Imaging-based

- Targeted
- Higher resolution



STARmap



10X Visium

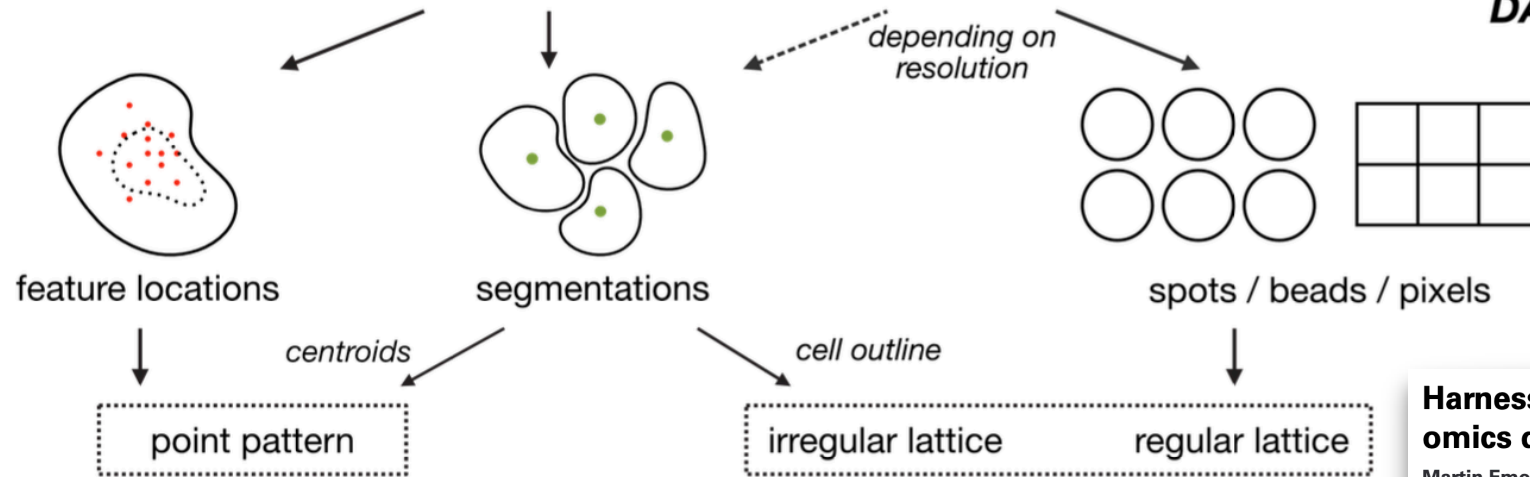
TECHNOLOGY

HTS-based

- Untargeted
- Lower resolution

B

DATA MODALITY



Samuel



Martin

Harnessing the potential of spatial statistics for spatial omics data with *pasta*

Martin Emons^{1,†}, Samuel Gunz^{1,†}, Helena L. Crowell², Izaskun Mallona¹, Malte Kuehl^{3,4}, Reinhard Furrer⁵, Mark D. Robinson^{1,*}

¹Department of Molecular Life Sciences and SIB Swiss Institute of Bioinformatics, University of Zurich, 8057 Zurich, Switzerland

²Centro Nacional de Análisis Genómico (CNAG), 08028 Barcelona, Spain

³Department of Clinical Medicine, Aarhus University, 8200 Aarhus N, Denmark

⁴Department of Pathology, Aarhus University Hospital, 8200 Aarhus N, Denmark

⁵Department of Mathematical Modeling and Machine Learning, University of Zurich, 8057 Zurich, Switzerland

[†]To whom correspondence should be addressed. Email: mark.robinson@mls.uzh.ch

^{*}The first two authors should be regarded as Joint First Authors.

pasta: Data representations determine spatial statistics options

Categorical (e.g., cell types)

Uni-
variate

Colocalization of one cell type and at which scale?

K, L and G functions

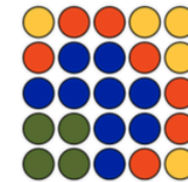


- cell type A
- cell type B
- cell type C
- cell type D

not common

How often are spots of the same cluster neighbouring each other?

Join count statistics



- cluster A
- cluster B
- cluster C
- cluster D

Bi-
variate

Colocalization between two cell types and at which scale?

Cross K, L and G functions

Multi-
variate

Colocalization of one cell to a set of other cell types?

Dot functions

Which clusters are found more frequently neighbouring each other?

Multivariate join count statistics

Numerical (e.g., gene expression)

Uni-
variate

At which scale is there (spatial) correlation of gene expression?

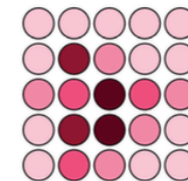
Mark correlation function



not common

Spatial autocorrelation of a gene?

Moran's I and relatives



Bi-
variate

Spatial correlation of two genes?

Bivariate Moran's I and relatives

Multi-
variate

not common

Spatial correlation of a set of genes?

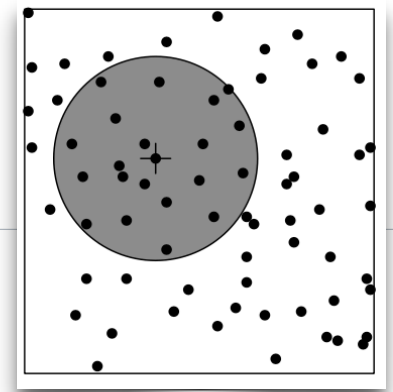
Multivariate Geary's C



Samuel



Martin



Correlation for **point patterns**

- Ripley's K function
- mathematical definition:

$$K(r) = \frac{1}{\lambda} \mathbb{E} [\text{number of } r\text{-neighbours of } u \mid \mathbf{X} \text{ has a point at location } u]$$

$$t(u, r, \mathbf{X}) = \sum_{j=1}^{n(\mathbf{X})} \mathbf{1} \{0 < \|u - x_j\| \leq r\}$$

Definition 7.1. If \mathbf{X} is a stationary point process, with intensity $\lambda > 0$, then for any $r \geq 0$

$$K(r) = \frac{1}{\lambda} \mathbb{E} [t(u, r, \mathbf{X}) \mid u \in \mathbf{X}] \quad (7.6)$$

does not depend on the location u , and is called the K-function of \mathbf{X} .

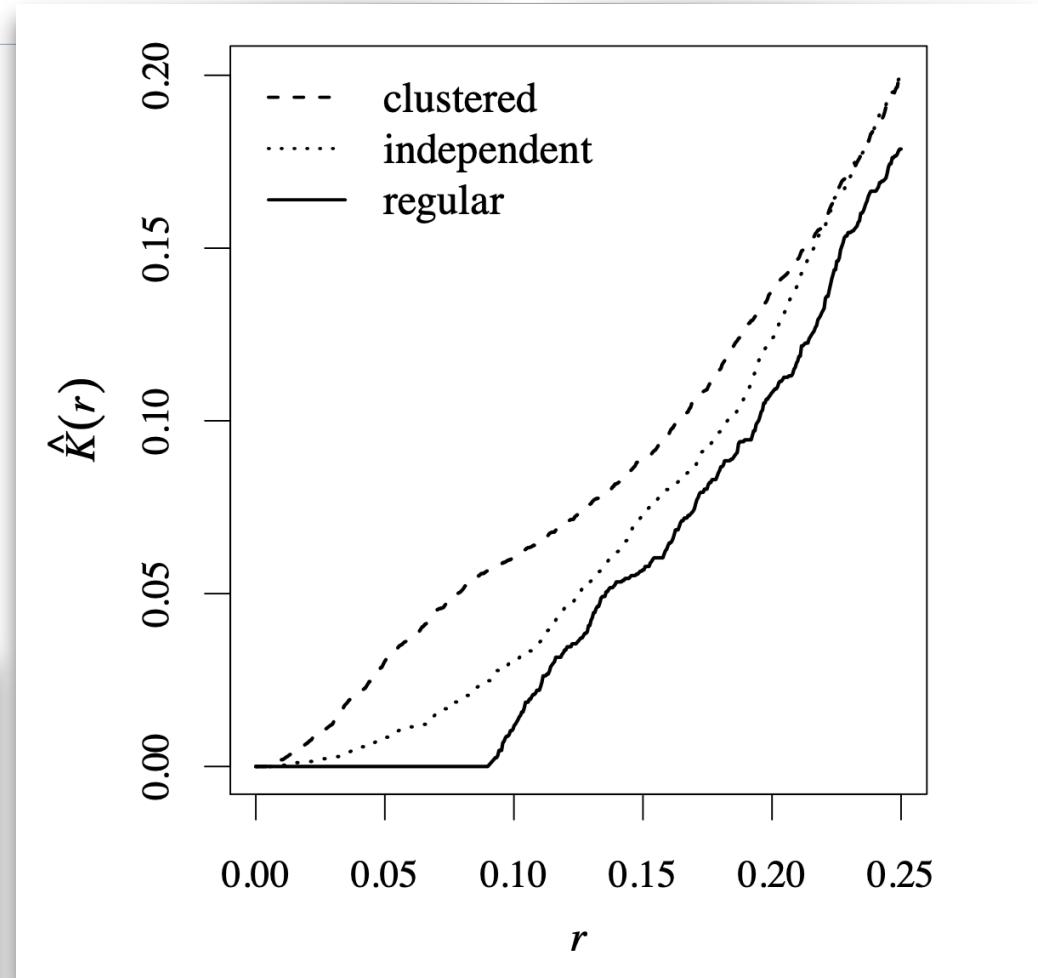
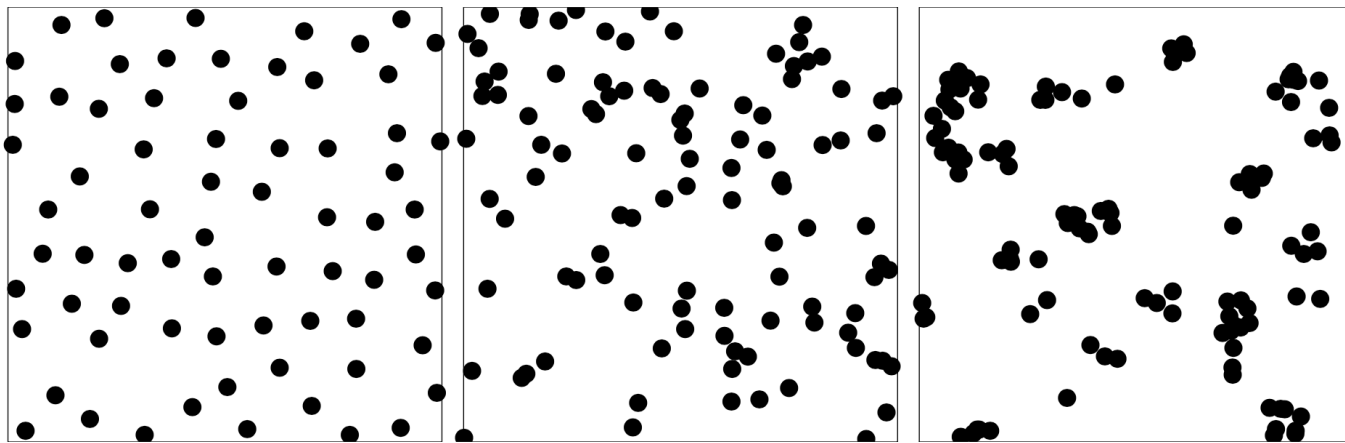
Correlation for **point patterns**

- Ripley's K function
- words definition: *the empirical K-function $K(r)$ is the cumulative average number of data points lying within a distance r of a typical data point*

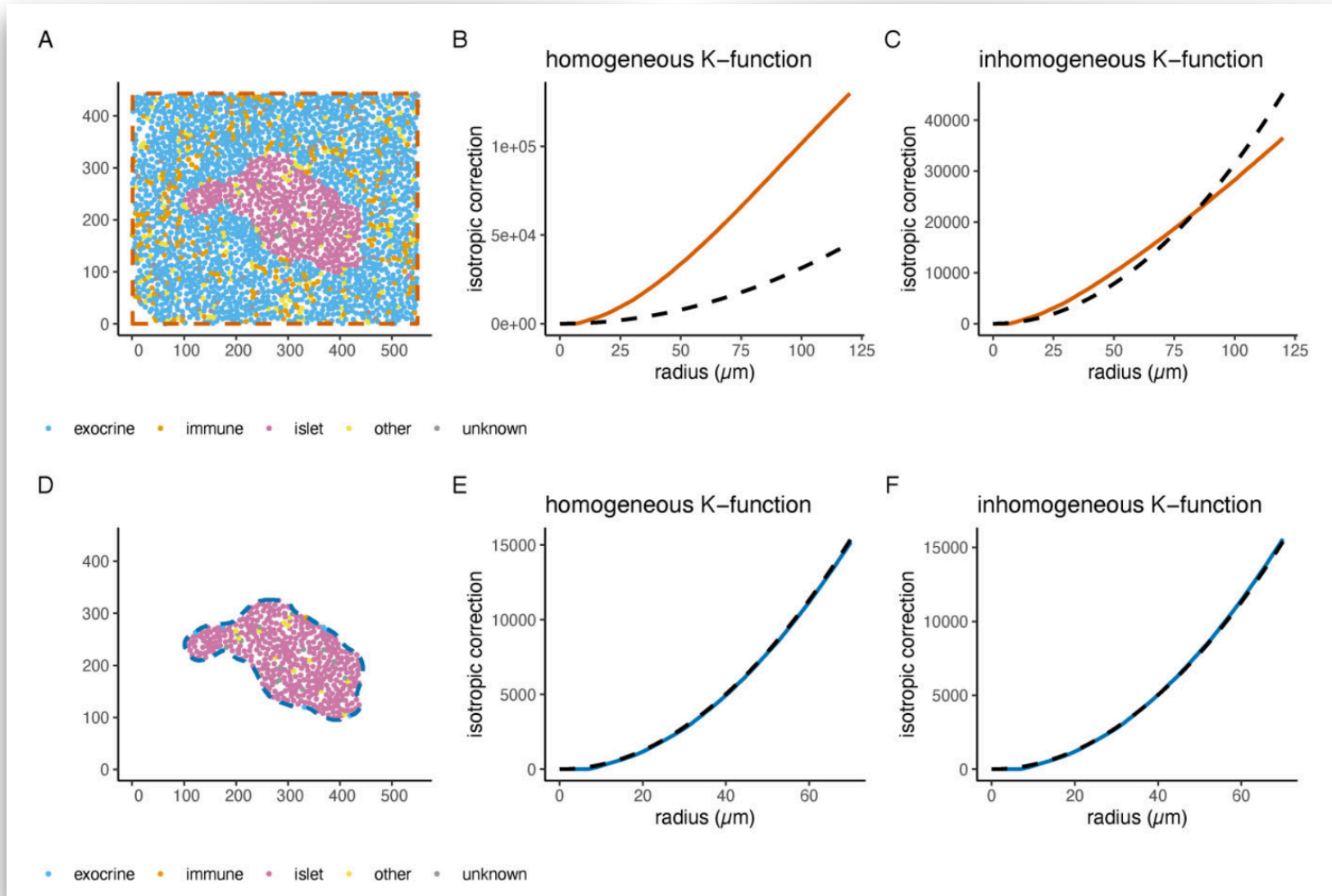
regular

independent

clustered



pasta: the 'gotcha' of spatial statistics — is it clustering or intensity?



Samuel



Martin

K-functions here:
clustering / intensity of
pink cells (islets).



Spatial autocorrelation: Global Moran's I

- Global measure of auto-correlation (correlation to signal nearby in space); assume homogeneity!
- Alternative: Geary's C

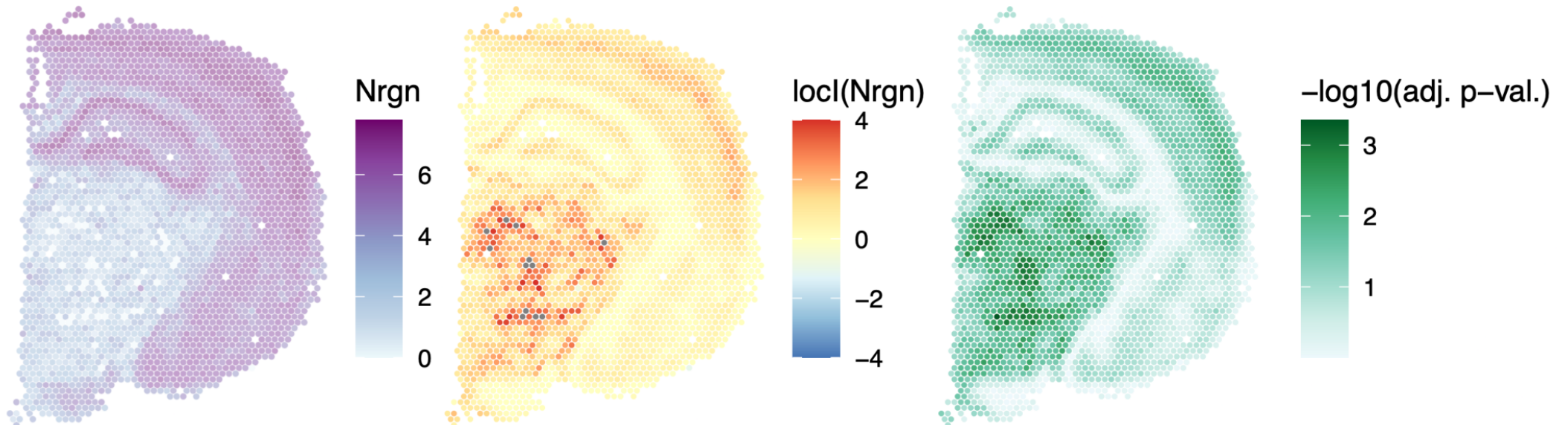
$$I = \frac{1}{\sum_{ij} w_{ij}} \frac{\sum_{ij} w_{ij} (X_i - \bar{X})(X_j - \bar{X})}{N^{-1} \sum_i (X_i - \bar{X})^2}$$

$$C = \frac{(N - 1) \sum_i \sum_j w_{ij} (x_i - x_j)^2}{2W \sum_i (x_i - \bar{x})^2}$$

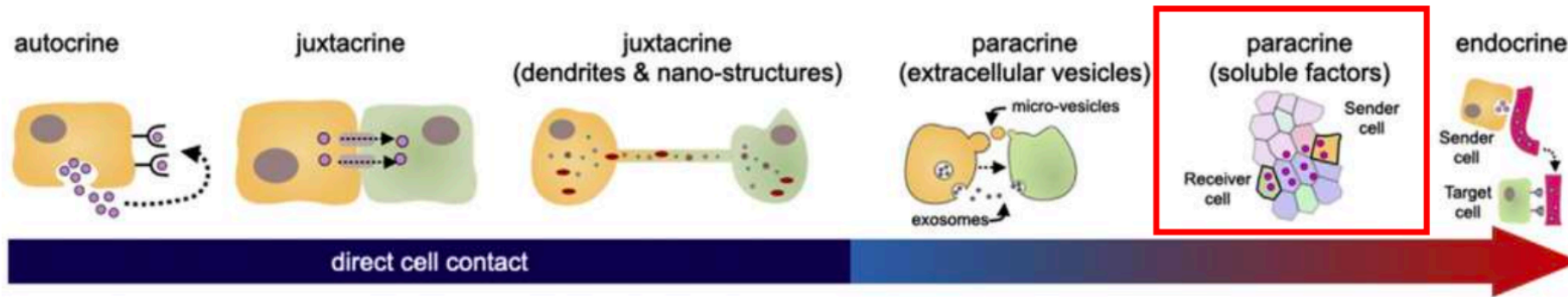
Spatial autocorrelation: Local Moran's I

- Local measure of auto-correlation (correlation to signal nearby in space)

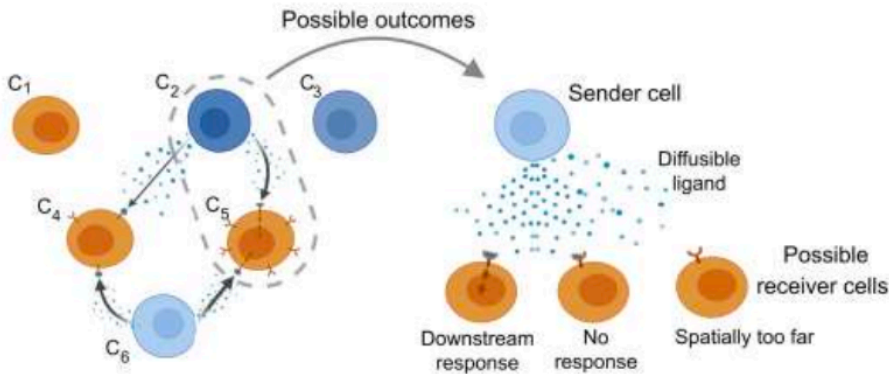
$$I_i = \frac{x_i - \bar{x}}{\sum_{k=1}^n (x_k - \bar{x})^2 / (n - 1)} \sum_{j=1}^n w_{ij} (x_j - \bar{x})$$



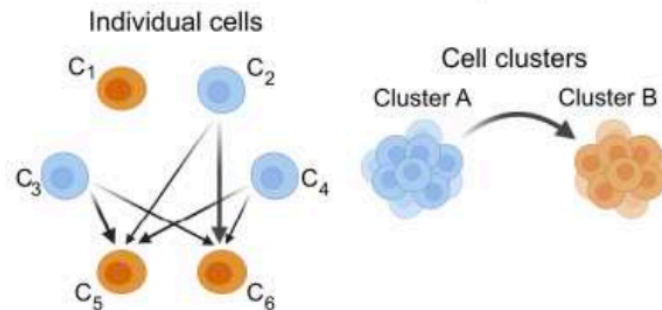
Various modes of CCC exist



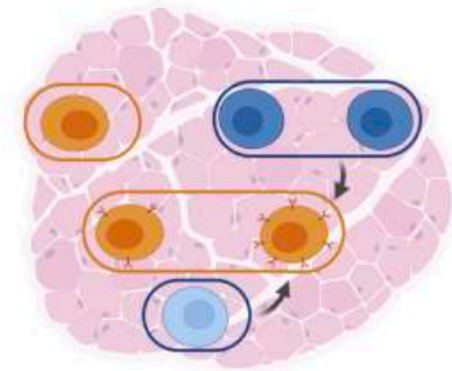
Cell-cell communication in tissue



Inference in scRNA-seq



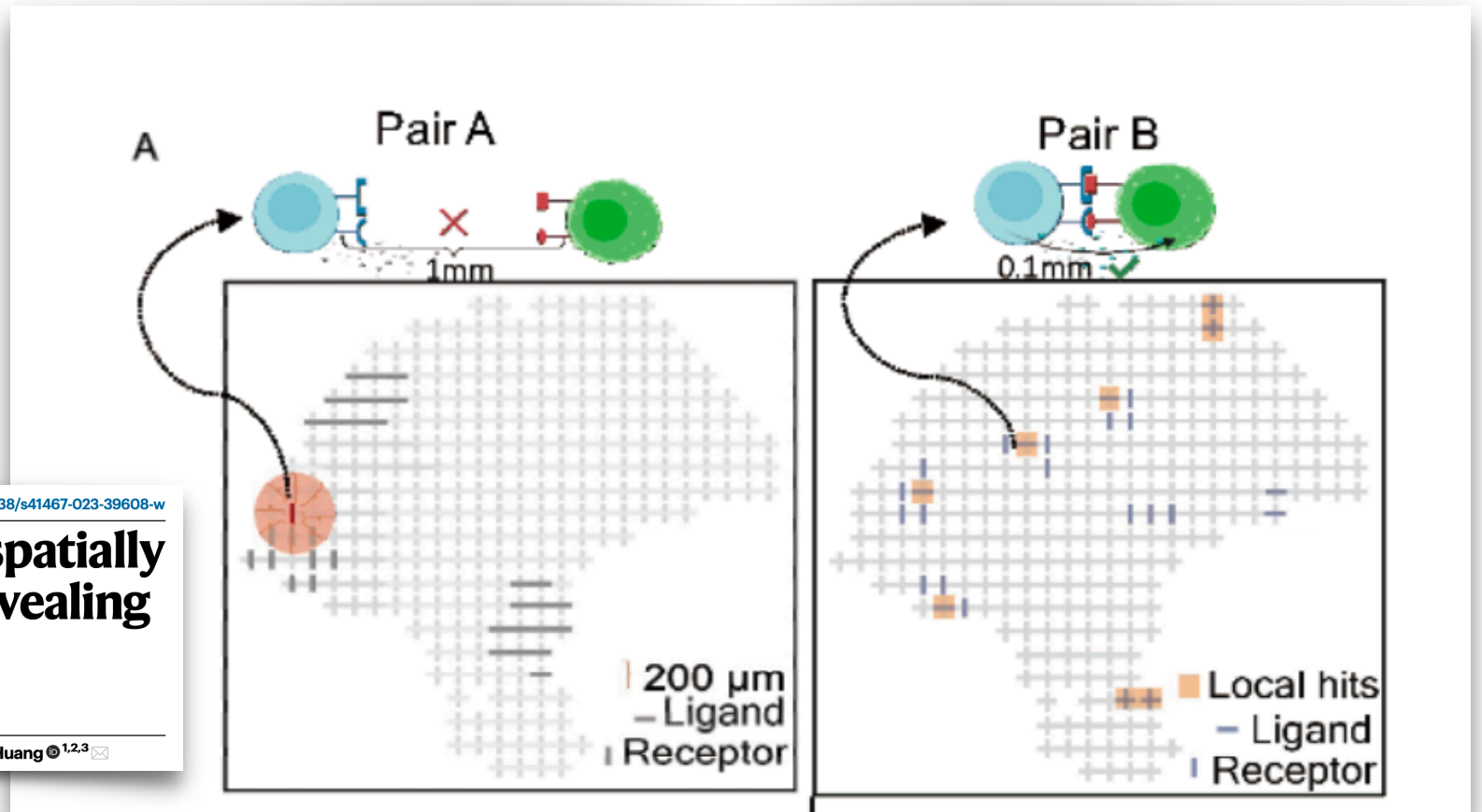
Inference in spatial transcriptomics



$$\text{Global Moran's } R = \frac{\sum_i \sum_j w_{ij} (x_i - \bar{x})(y_j - \bar{y})}{\sqrt{\sum_i (x_i - \bar{x})^2} \sqrt{\sum_i (y_i - \bar{y})^2}}$$

Cell-cell communication

- SpatialDM: Global Moran's R, which is a bivariate version of Moran's I



$$\text{Global Moran's } R = \frac{\sum_i \sum_j w_{ij} (x_i - \bar{x})(y_j - \bar{y})}{\sqrt{\sum_i (x_i - \bar{x})^2} \sqrt{\sum_i (y_i - \bar{y})^2}},$$

Cell-cell communication

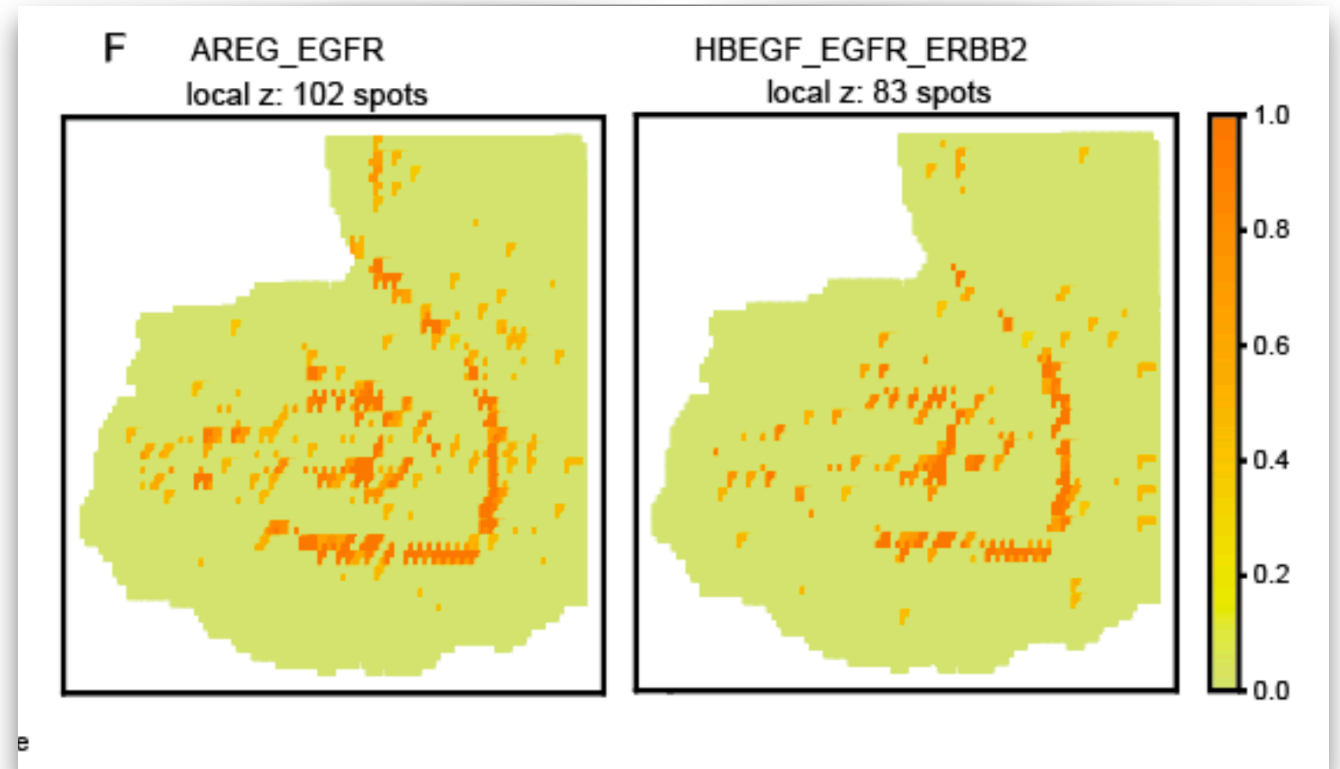
- SpatialDM: Global Moran's R, which is a bivariate version of Moran's I

SpatialDM Non-significant due to spatial range

↓

Global Moran's R = $\frac{N}{W} \sum_i \sum_j w_{ij} \bar{x}_i \bar{y}_j$

Local Moran's R = $\sum_j w_{ij} \bar{x}_i \bar{y}_j + \sum_j w_{ij} \bar{y}_i \bar{x}_j$

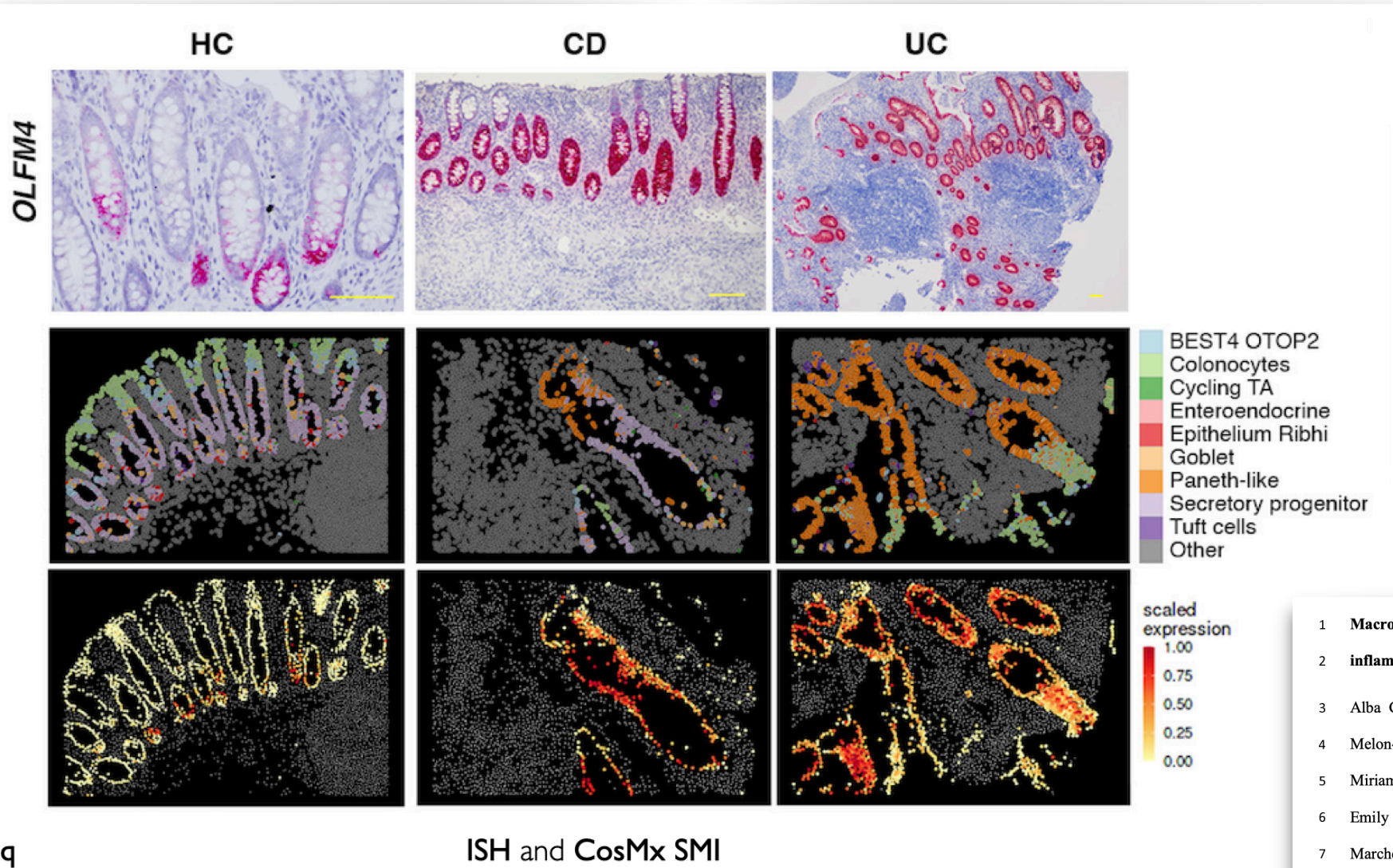




Research

- `spatialFDA`: Flexible modeling of point pattern summaries —> Martin
- `DESpace2`: DE beyond markers/SVGs: “differential spatial patterns” —> Peiying
- `sosta`: “Spatial structure”-focused analyses —> Samuel
- `OSTA`: Orchestrating spatial transcriptomics analysis with Bioconductor
- `SpaceHack`: using consensus clustering to consolidate domain detection

Tissue “structures” are often visible



- healthy control (HC)
- Crohn's disease (CD)
- ulcerative colitis (UC)

- 1 Macrophage and neutrophil heterogeneity at single-cell spatial resolution in inflammatory bowel disease
- 2
- 3 Alba Garrido-Trigo^{1,2}, Ana M. Corraliza^{1,2}, Marisol Veny^{1,2}, Isabella Dotti^{1,2}, Elisa Melon-Ardanaz^{1,2}, Aina Rill³, [Helena L. Crowell](#)⁴, Ángel Corbí⁵, Victoria Gudiño^{1,2}, Miriam Esteller^{1,2}, Iris Álvarez-Teubel^{1,2}, Daniel Aguilar^{1,2}, M Carme Masamunt^{1,2}, Emily Killingbeck⁶, Youngmi Kim⁶, Michael Leon⁶, Sudha Visvanathan⁷, Domenica Marchese⁸, Ginevra Caratù⁸, Albert Martin-Cardona^{2,9}, Maria Esteve^{2,9}, Julian Panés,^{1,2} Elena Ricart^{1,2}, Elisabetta Mereu^{3,*}, Holger Heyn^{8,10,*}, Azucena Salas^{1,2}
- 4
- 5
- 6
- 7
- 8



Research

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Orchestrating Spatial Transcriptomics Analysis with Bioconductor

- <https://bioconductor.org/books/OSTA>

Orchestrating Spatial Transcriptomics Analysis with Bioconductor

Helena L. Crowell^{1,*}, Yixing Dong^{2,3,*}, Ilaria Billato⁴, Peiying Cai^{5,6}, Martin Emons^{5,6}, Samuel Gunz^{5,6}, Boyi Guo⁷, Mengbo Li^{8,9,10}, Alexandru Mahmoud¹¹, Artür Manukyan¹², Hervé Pagès¹³, Pratibha Panwar^{14,15,16}, Shreya Rao^{14,15,17}, Callum J. Sargeant⁸, Lori Shepherd Kern¹⁸, Marcel Ramos^{19,20}, Jieran Sun^{2,3}, Michael Totty²¹, Vincent J. Carey¹¹, Yunshun Chen^{8,9,10}, Leonardo Collado-Torres^{21,22,23}, Shila Ghazanfar^{14,15,16}, Kasper D. Hansen^{21,24,25}, Keri Martinowich^{22,26,27,28}, Kristen R. Maynard^{22,26,27}, Ellis Patrick^{14,15,16,17}, Dario Righelli²⁹, Davide Risso^{30,31}, Simone Tiberi³², Levi Waldron^{19,20}, Raphael Gottardo^{2,3,33,†}, Mark D. Robinson^{5,6,†}, Stephanie C. Hicks^{21,25,34,35,†}, and Lukas M. Weber^{36,†}

Book is available. Preprint on bioRxiv.

(Successor of the OSCA book: <https://bioconductor.org/books/OSCA/>)

6 Example datasets

7 Python interoperability

Sequencing-based platforms

8 Introduction

9 Reads to counts

10 Quality control

11 Intermediate processing

12 Deconvolution

13 Workflow: Visium DLPFC

14 Workflow: Visium CRC

15 Workflow: Visium HD

Imaging-based platforms

16 Introduction

17 Segmentation

18 Quality control

19 Intermediate processing

20 Neighborhood analysis

21 Cell-cell communication

22 Sub-cellular analysis

23 Workflow: Xenium

24 Workflow: CosMX

Platform-independent analyses

25 Normalization



Research

- `spatialFDA`: Flexible modeling of point pattern summaries —> Martin
- `DESpace2`: DE beyond markers/SVGs: “differential spatial patterns” —> Peiying
- `sosta`: “Spatial structure”-focused analyses —> Samuel
- `OSTA`: Orchestrating spatial transcriptomics analysis with Bioconductor
- **SpaceHack: using consensus clustering to consolidate domain detection**

Meta-benchmark

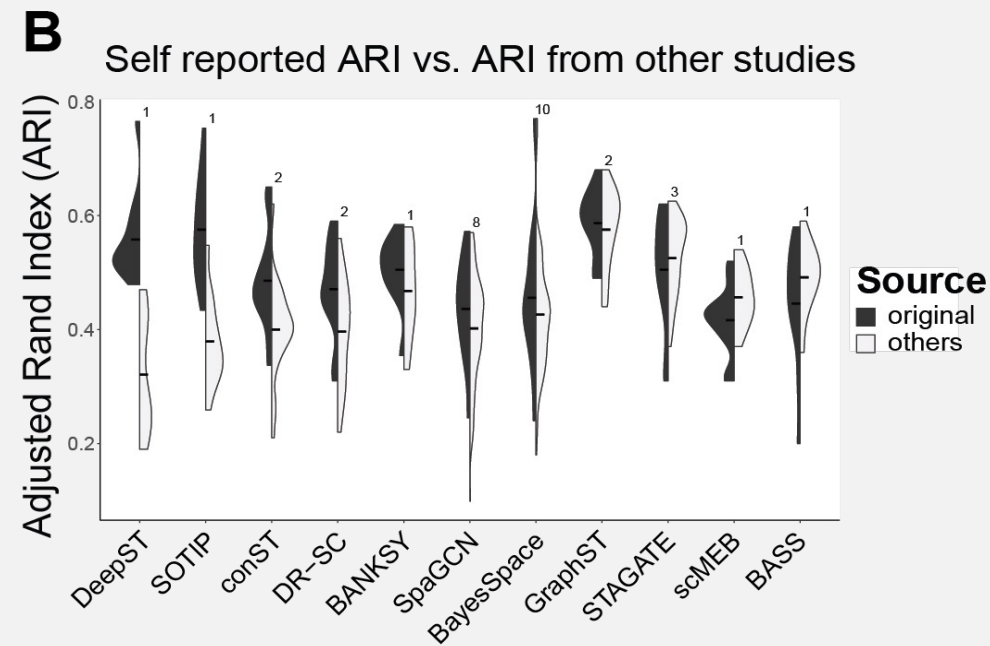
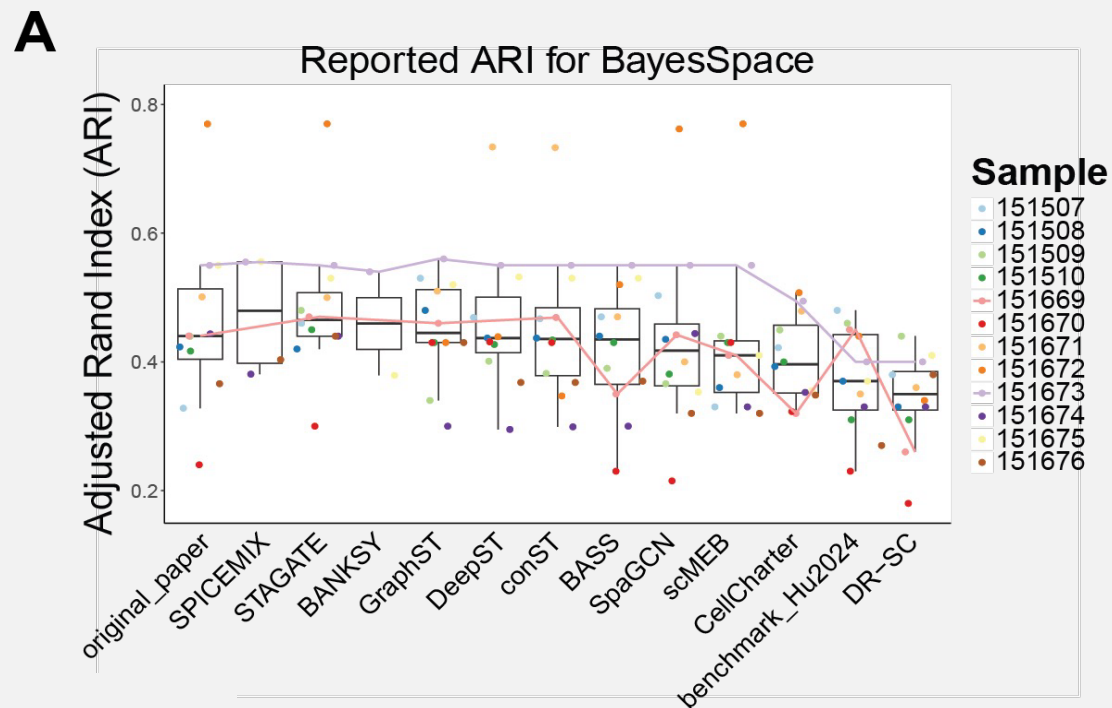
Reported method performances are inconsistent across studies

Beyond benchmarking: an expert-guided consensus approach to spatially aware clustering

Jieran Sun^{1†}, Kirti Biharie^{2,3†}, Peiyang Cai^{4†}, Niklas Müller-Böttcher^{5†}, Paul Kiessling^{6†}, Meghan A. Turner^{7†}, Søren H. Dam^{8,9†}, Florian Heyl^{10,11†}, Sarusan Kathirchelvan⁴, Martin Emons⁴, Samuel Gunz⁴, Sven Twardziok⁵, Amin El-Heliebi¹², Martin Zacharias¹³, SpaceHack 2.0 participants, Roland Eils³, Marcel Reinders³, Raphael Gottardo¹, Christoph Kuppe⁶, Brian Long^{7*}, Ahmed Mahfouz^{2,3*}, Mark D. Robinson^{4*}, Naveed Ishaque^{5*}



Peiyang Cai



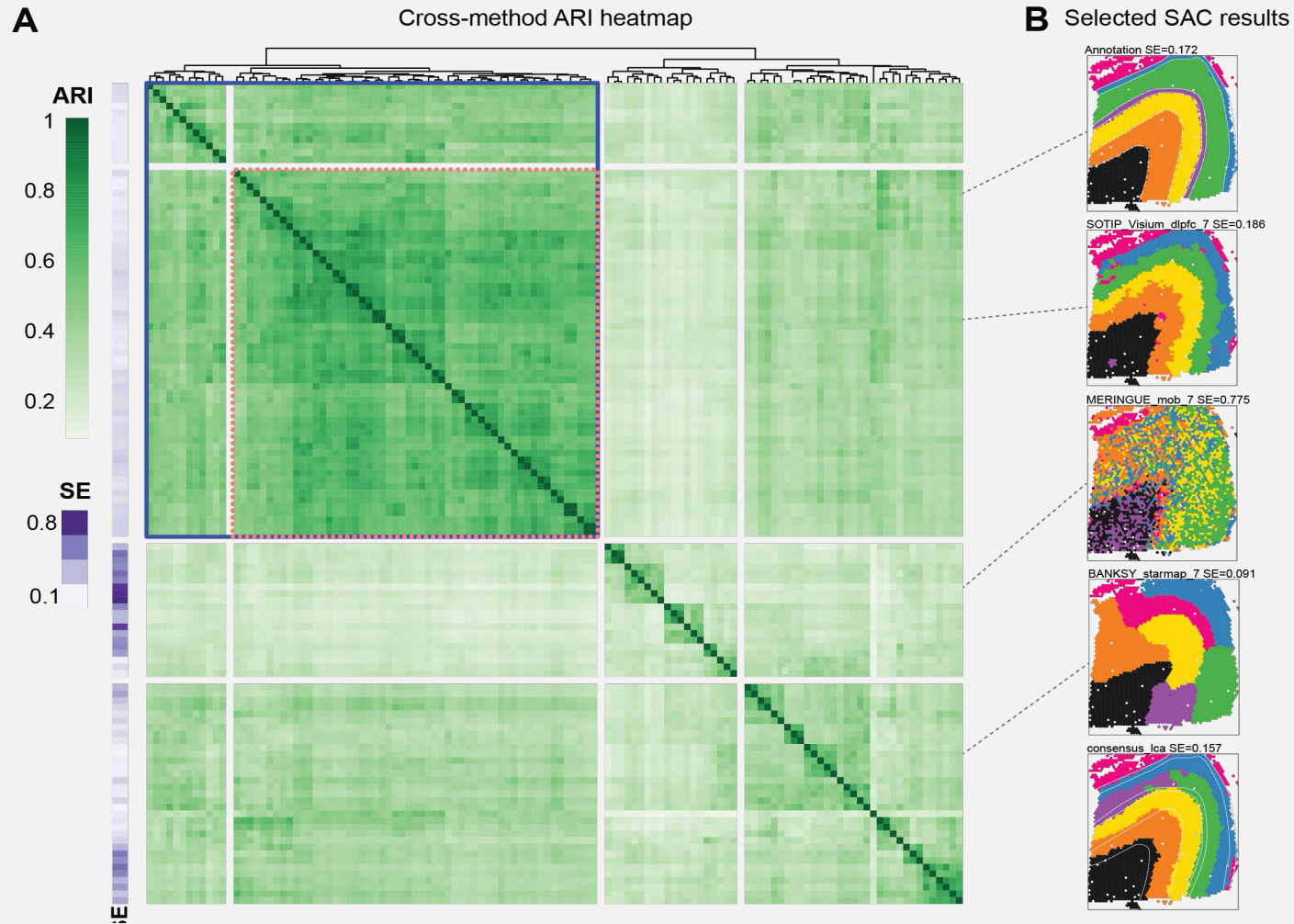
Ensemble clustering

Methods are often more similar to each other than to the ground truth.

Beyond benchmarking: an expert-guided consensus approach to spatially aware clustering

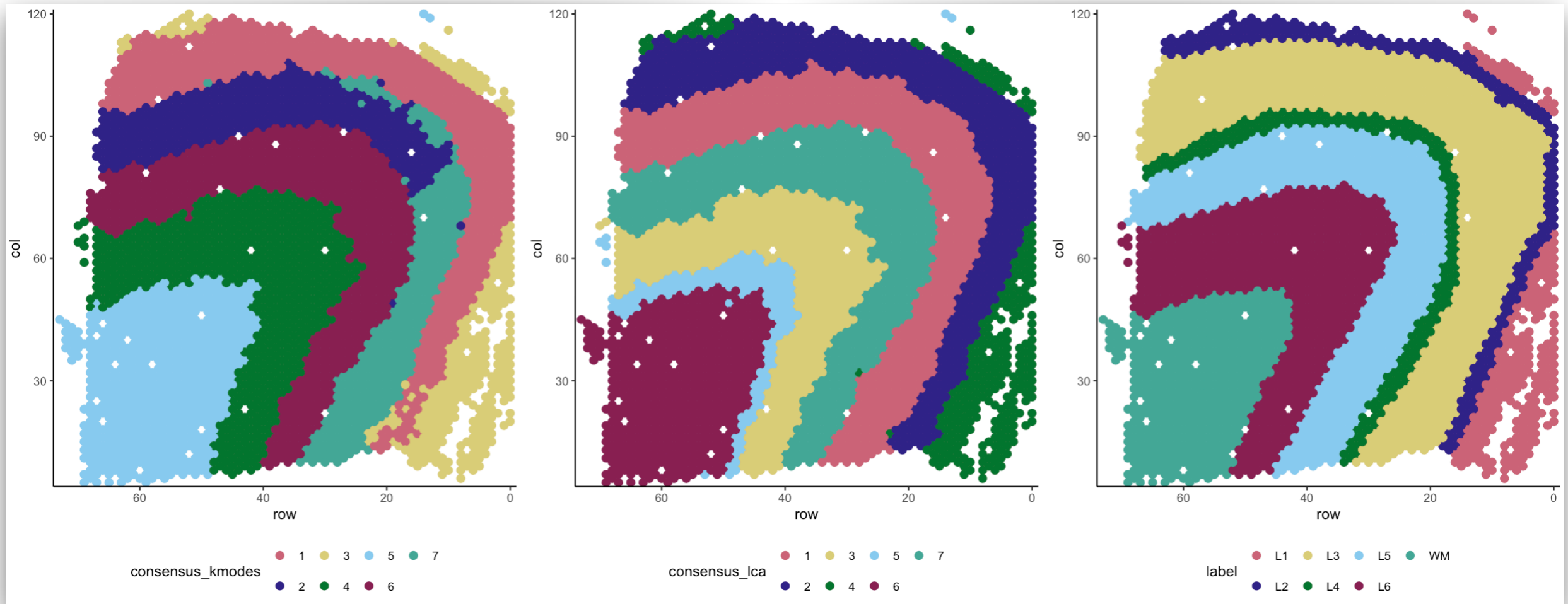
Jieran Sun^{1†}, Kirti Biharie^{2,3†}, Peiyong Cai^{4†}, Niklas Müller-Böttcher^{5†}, Paul Kiessling^{6†}, Meghan A. Turner^{7†}, Søren H. Dam^{8,9†}, Florian Heyl^{10,11†}, Sarusan Kathirchelvan⁴, Martin Emons⁴, Samuel Gunz⁴, Sven Twardziok⁵, Amin El-Heliebi¹², Martin Zacharias¹³, SpaceHack 2.0 participants, Roland Eils⁵, Marcel Reinders³, Raphael Gottardo¹, Christoph Kuppe⁶, Brian Long^{7*}, Ahmed Mahfouz^{2,3*}, Mark D. Robinson^{4*}, Naveed Ishaque^{5*}

Smoothness Entropy (Low = smooth) →



Consensuses

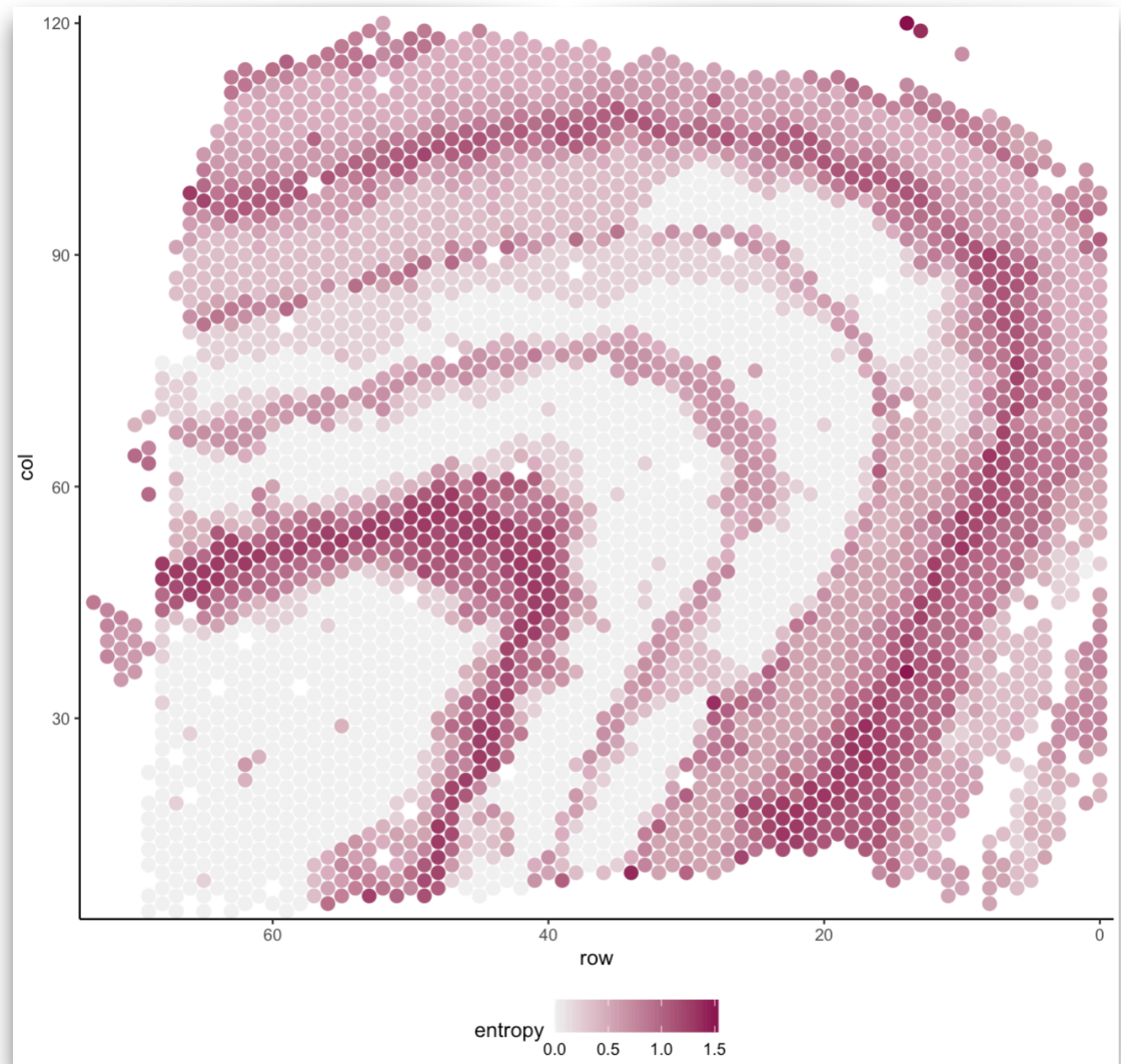
“ground truth”



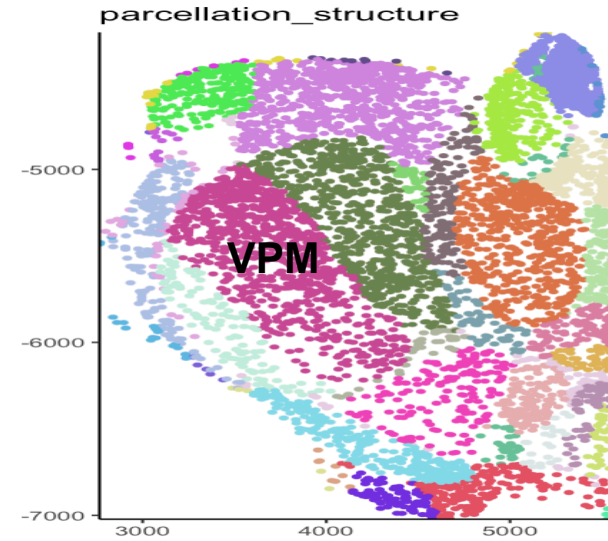
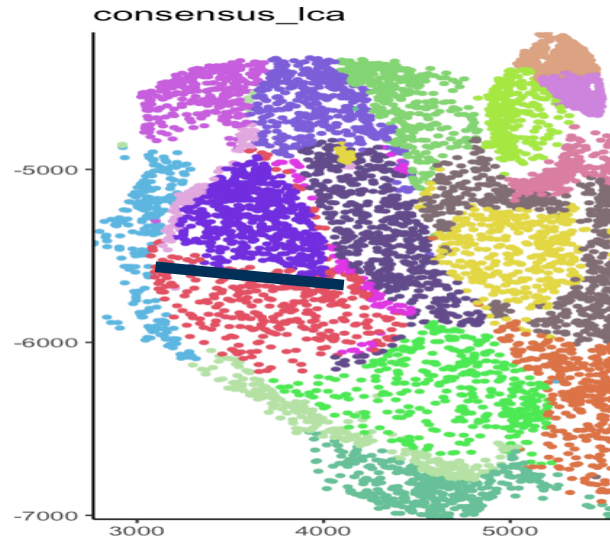
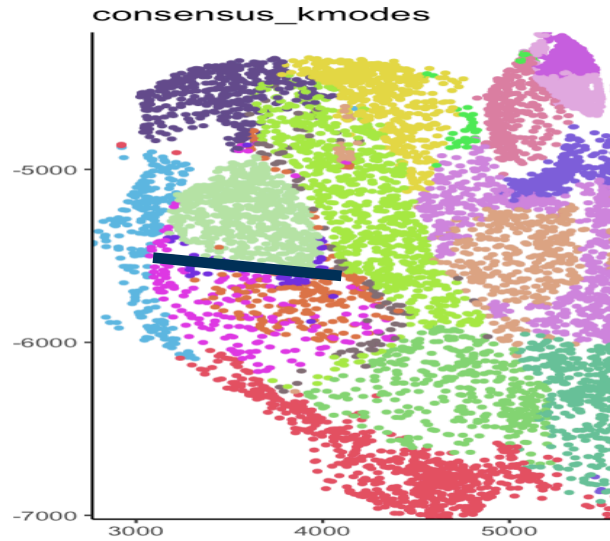
Entropy #2: Understanding spot-level uncertainty (across methods)

Entropy in the sense of how stable across algorithms

(align the spot-wise cluster labels across methods, entropy across label proportions)

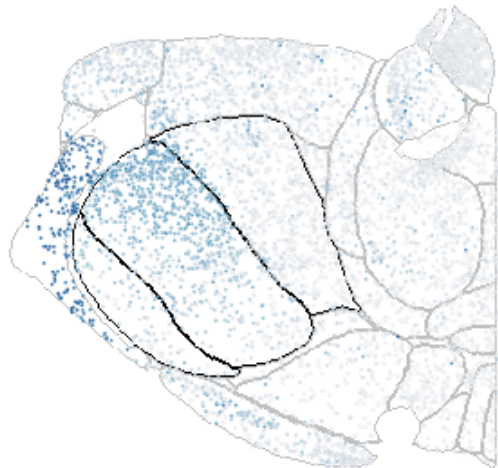


VPM

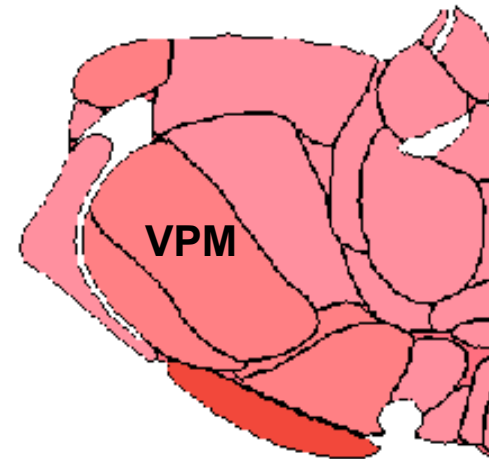
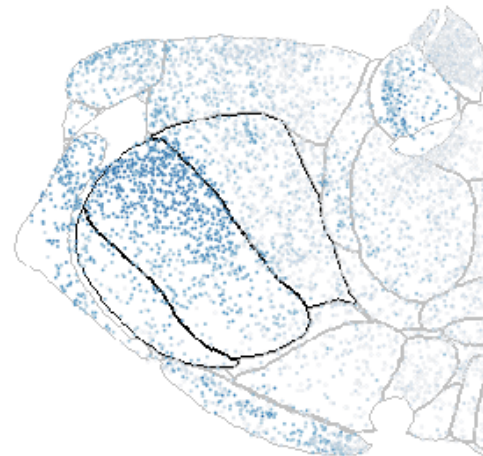


Slide from
Meghan Turner

Section 6.8
Pvalb



Section 6.8
Kcnab3



Concluding remarks

- You are collecting/analyzing spatial data: what **spatial features** do you want to quantify?
- A few places where (classical) spatial statistics might be useful; data determines what you can do: point patterns versus lattice
- Tools available for various task: Functional data analysis (Martin), multi-cellular structure-based analyses (Samuel),