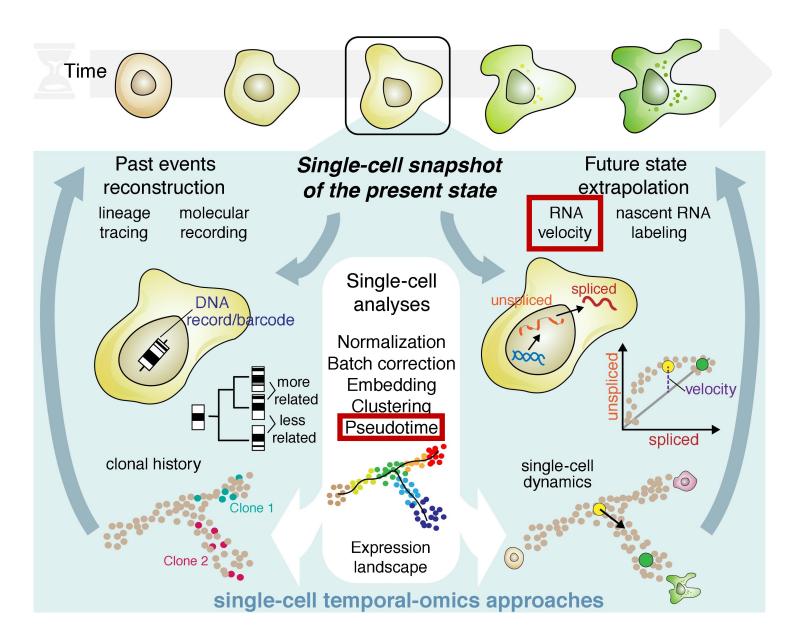
RNA velocity

Single Cell Transcriptomics in Python Alex Lederer

Emergence of single-cell temporal-omics approaches



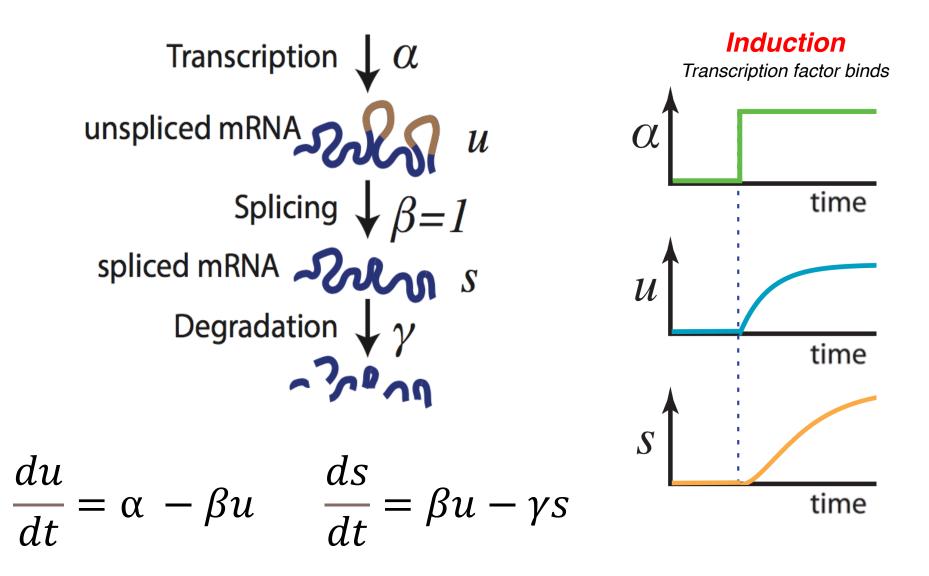
Lederer and La Manno 2020

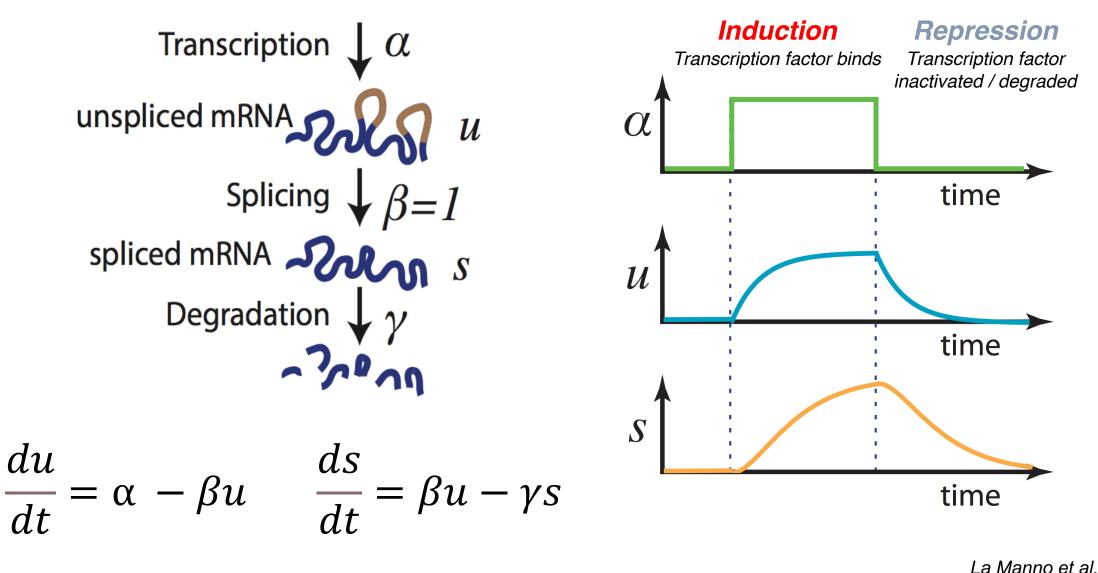
```
Transcription \downarrow \alpha
unspliced mRNA \beta = 1
Splicing 4\beta = 1
spliced mRNA \gamma
Degradation 4\gamma
```

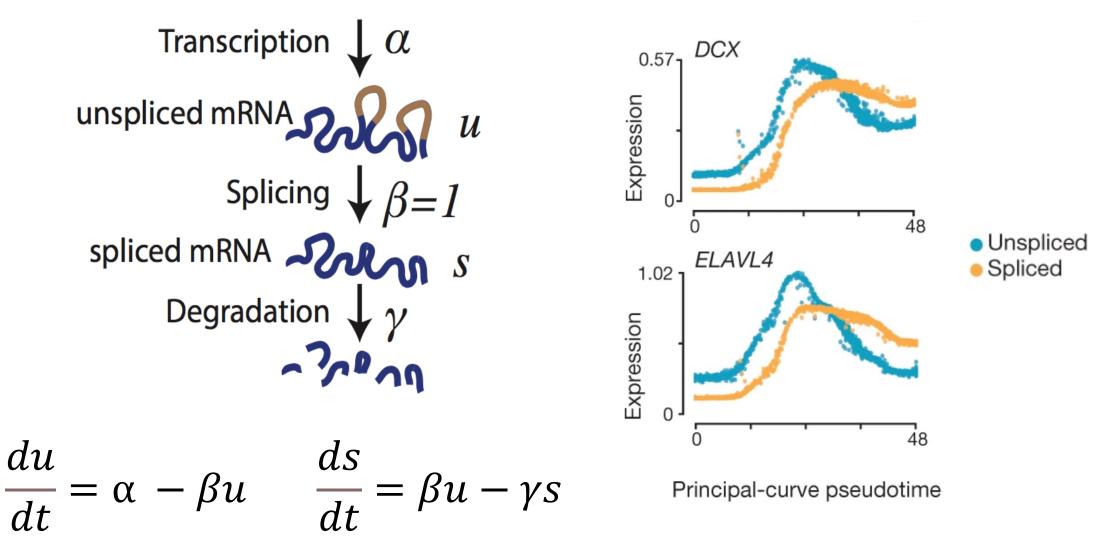
Transcription
$$\downarrow \alpha$$

unspliced mRNA $\downarrow \beta = 1$
spliced mRNA $\downarrow \beta = s$
Degradation $\downarrow \gamma$

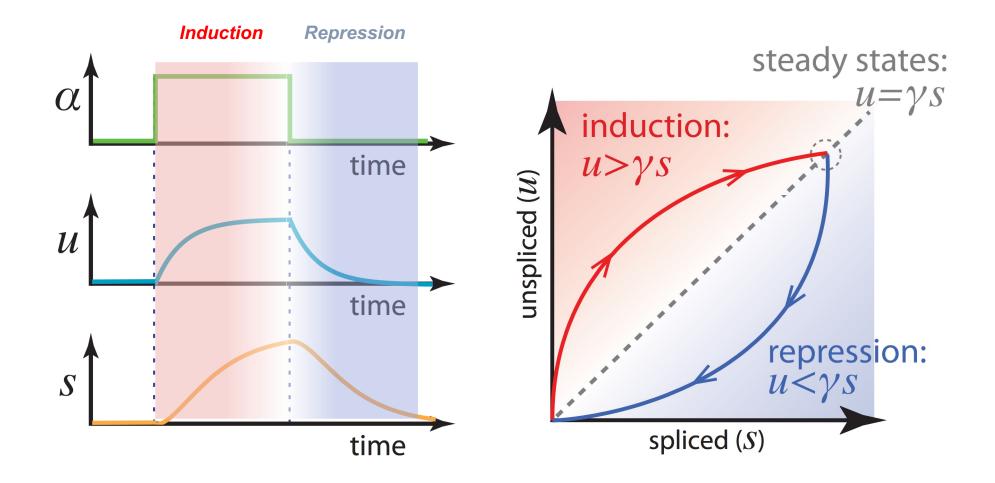
 $\frac{du}{dt} = \alpha - \beta u \qquad \frac{ds}{dt} = \beta u - \gamma s$





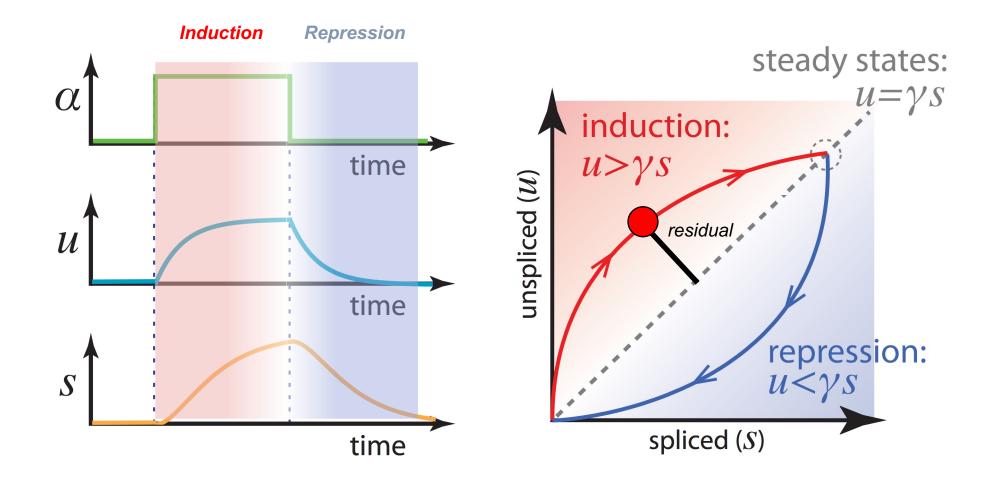


Phase portraits show changes to relative unspliced and spliced RNA abundances



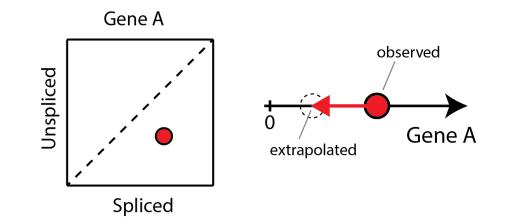
La Manno et al. 2018

Phase portraits show changes to relative unspliced and spliced RNA abundances

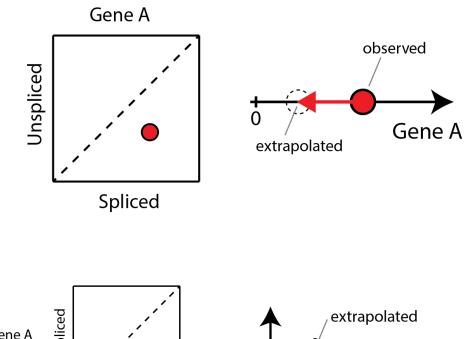


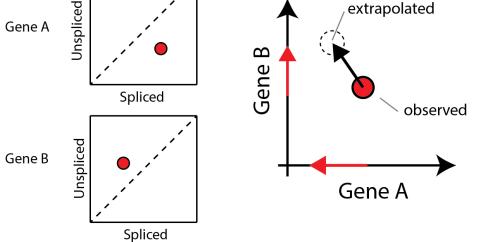
La Manno et al. 2018

RNA velocity is a gene-specific estimate



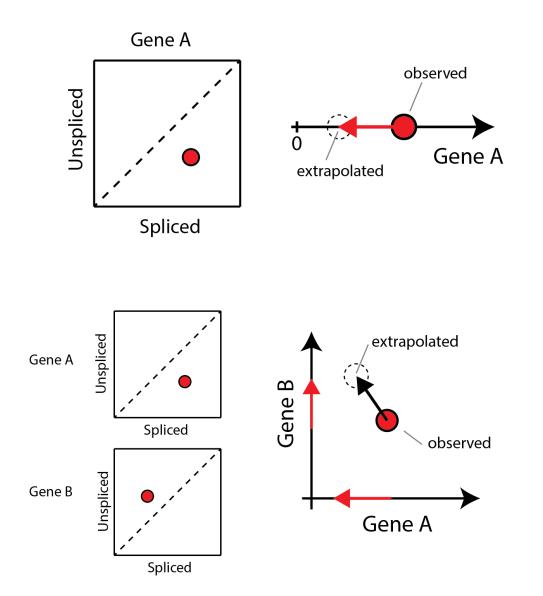
RNA velocity is a gene-specific estimate





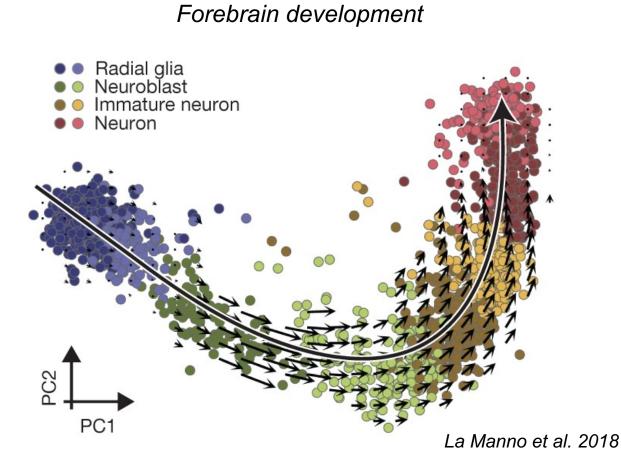
La Manno et al. 2018

RNA velocity is a gene-specific estimate

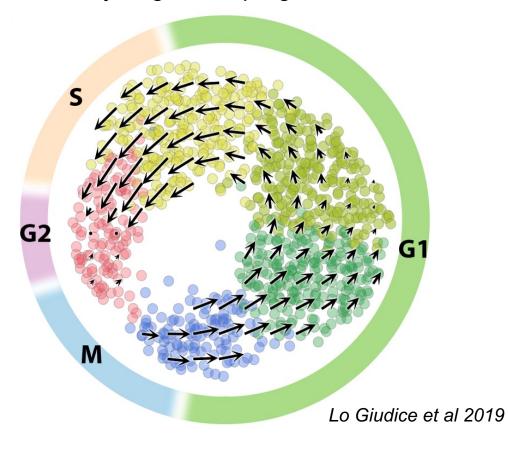


Problem: each gene is on a different time scale!

Examples of RNA velocity applied to trajectories



Cycling retinal progenitors

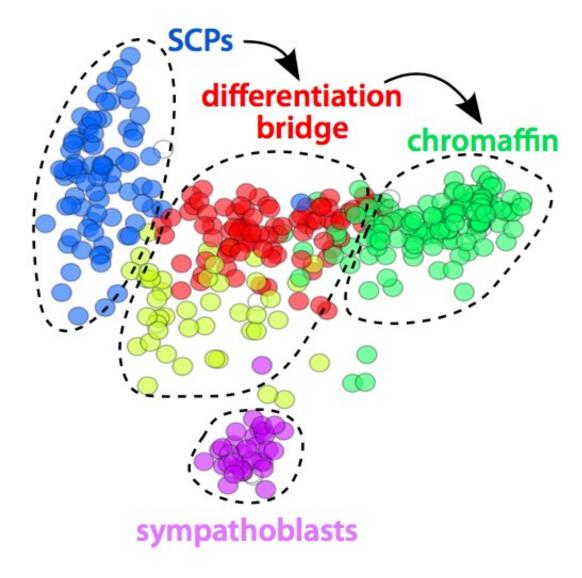


When should I use RNA velocity?

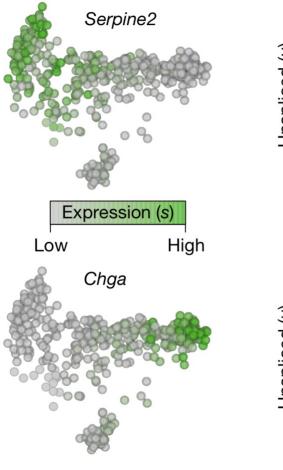
- I expect there to be a trajectory in my single-cell dataset, but:
 - I do not know the direction along which cells should change
 - I expect the trajectories to change for a similar population of cells but in different dataset conditions (in response to gene perturbations, environmental stimuli, etc)

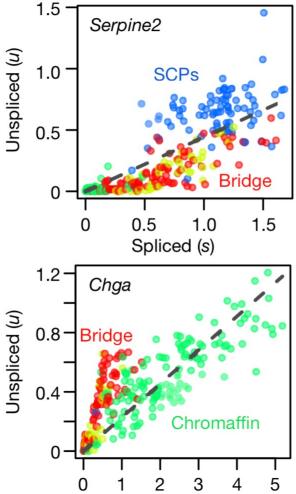
Note: the changes between cell states should unfold on a relatively short time scale (hours to days) and not a long time scale (weeks to months).

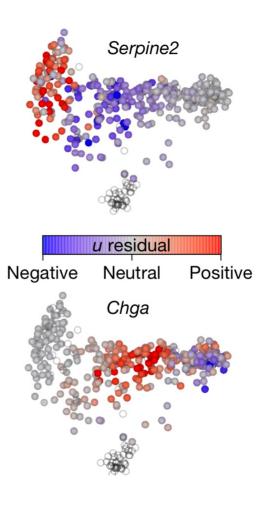
RNA velocity to describe a simple differentiation process



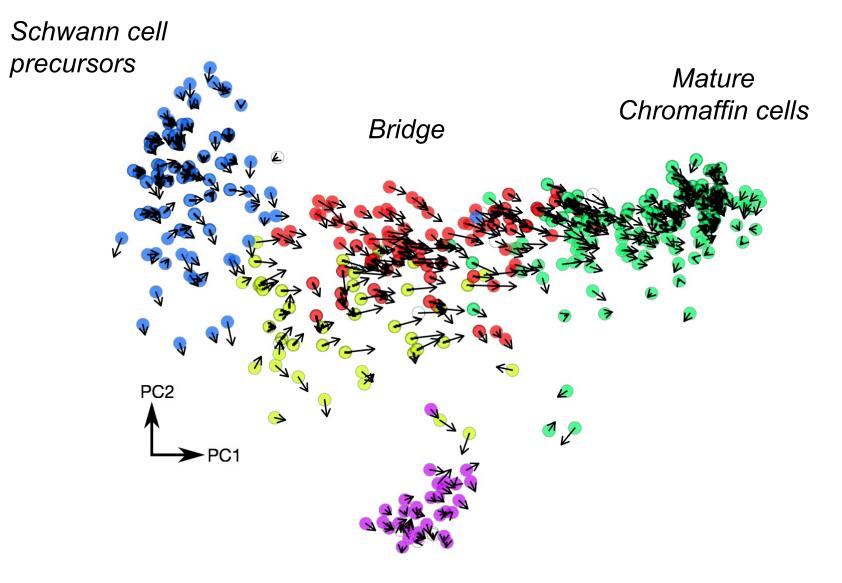
RNA velocity to describe a simple differentiation process



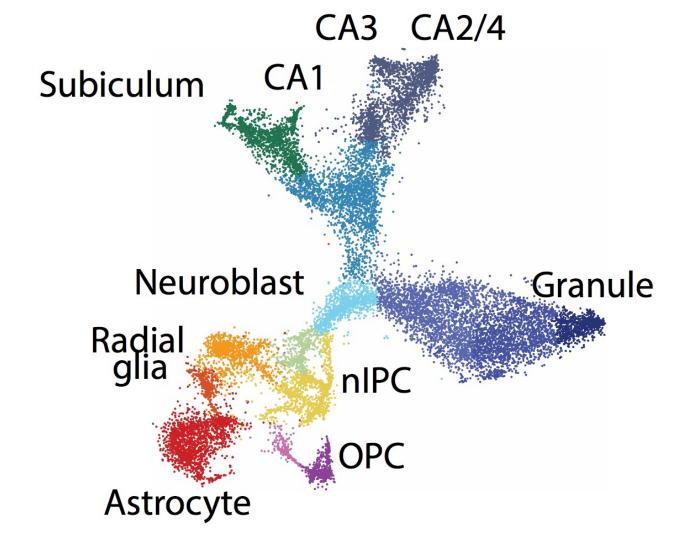




RNA velocity to describe a simple differentiation process



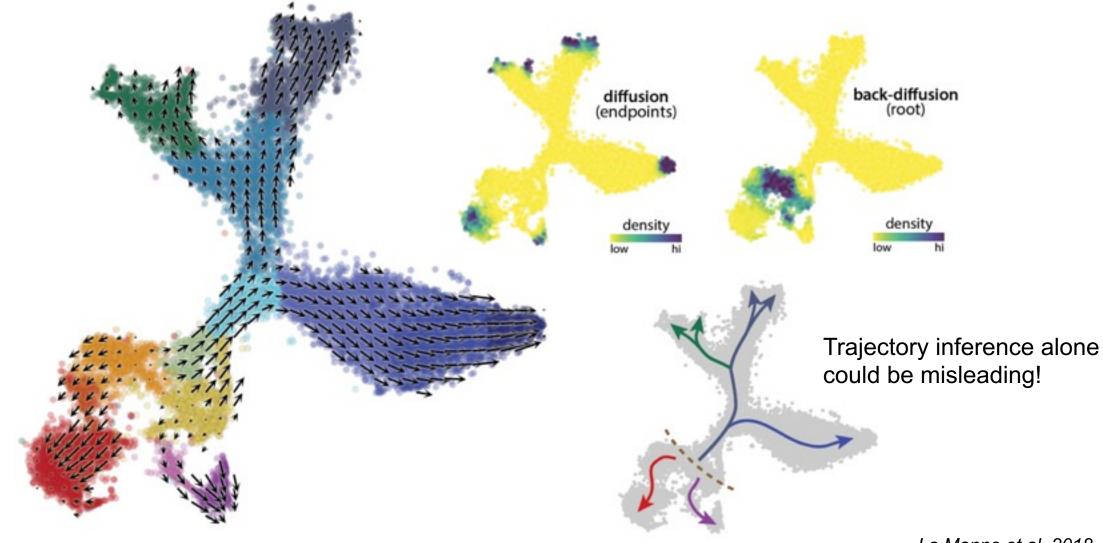
A more complex application of RNA velocity



Pseudotime analysis could be misleading!

La Manno et al. 2018

A more complex application of RNA velocity



Other recent velocities!

Generalizing RNA velocity to transient cell states through dynamical modeling

Chromatin Potential Identified by Shared Single-Cell Profiling of RNA and Chromatin

UniTVelo: temporally unified RNA velocity reinforces single-cell trajectory inference

Protein velocity and acceleration from single-cell multiomics experiments

Representation learning of RNA velocity

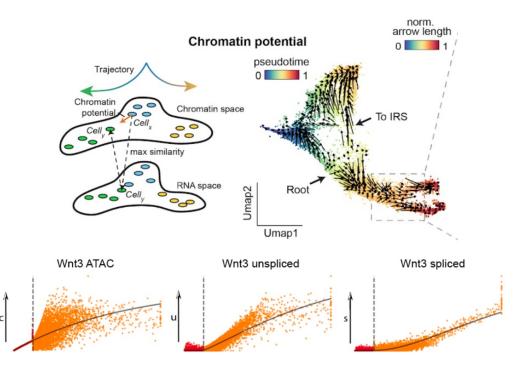
reveals robust cell transitions

A relay velocity model infers cell-dependent RNA velocity

DeepVelo: Single-cell transcriptomic deep velocity field learning with neural ordinary differential equations

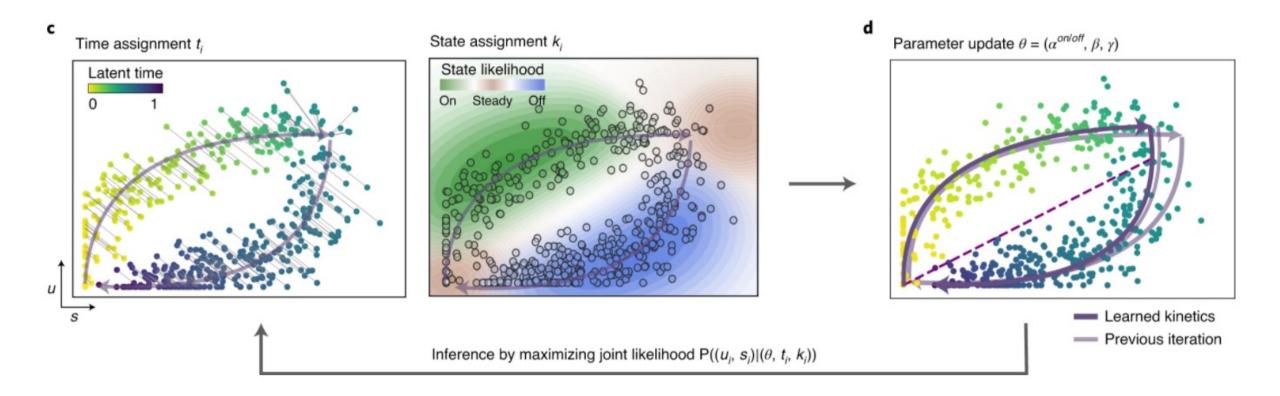
Single-cell multi-omic velocity infers dynamic and decoupled gene regulation

Multi-omic single-cell velocity models epigenometranscriptome interactions and improves cell fate prediction Time assignment t_i Latent time t_i State assignment k_i State assi



+ about 40-50 additional velocity methods! (it's not yet a category on scrna-tools.org)

Using expectation-maximization to estimate velocity and RNA kinetics with scvelo



Bergen et al. Nat Biotech. 2020

Should you perform RNA velocity analysis on your data?

RNA velocity unraveled:

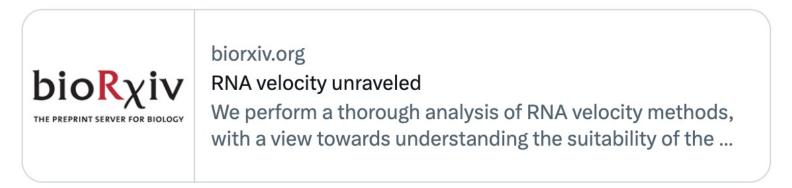
https://journals.plos.org/ploscompbiol/article?id=10.1371/journal.pcbi.1010492



@lpachter

If you work w/ single-cell RNA-seq & are performing RNA velocity analyses, you might find this @GorinGennady et al. preprint w/ Meichen Fang & Tara Chari of interest. It's a deep dive into the method, and navigation of the 67 pages may be aided w/ this 1/

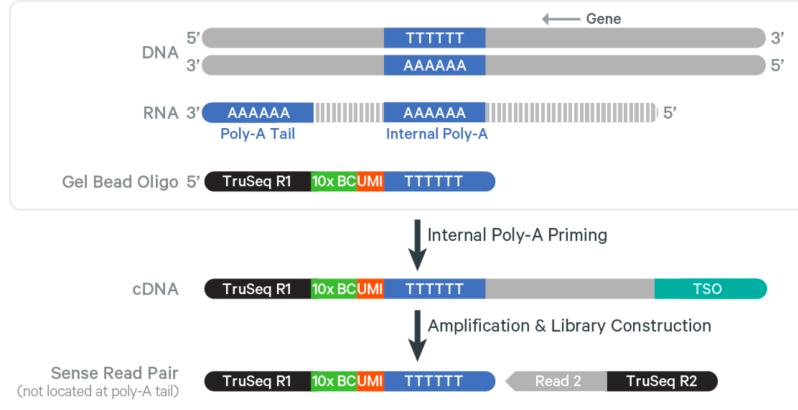
...



Should you perform RNA velocity analysis on your data?

- Do I expect a temporal axis of variation in your data?
- Do I expect the time scale of temporal change to unfold in hours/days or weeks?
- Which single-cell technology do you use (single cell vs single nuclei) and how much detection of intronic reads do you expect?

Internal priming and measurement of additional intronic regions

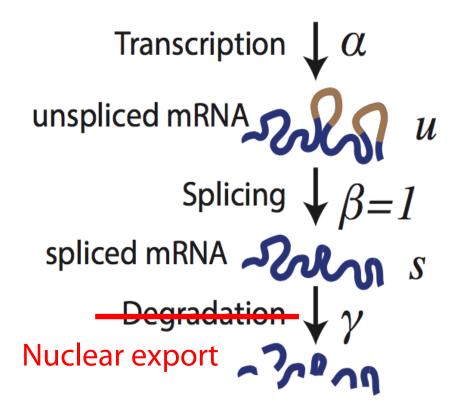


10X Genomics Technical Note, CG000376 (2021)

- The poly(dT) primer can prime to an internal poly-A stretch
- In humans, there are 21x more poly-A stretches found in introns than exons

Conclusion: there may be intron detection biases between different genes in the data!

The RNA life cycle captured by single nucleus RNA-seq is different



In single-nuclei data, it is not possible to measure the degradation rate for a gene.

The nuclear export rate may be instead described by changes to spliced abundance.

Nuclear export occurs much more quickly than the degradation – be careful!

Two steps for RNA velocity analysis

Step 1. Intron/exon counting

- Command line tool that takes the output bam files from CellRanger and generates separate count matrices for spliced and unspliced mRNA
- Original tool (*velocyto*): <u>https://velocyto.org/velocyto.py/tutorial/cli.html</u>
- Also possible with STARsolo and alevin-fry methods for read alignment

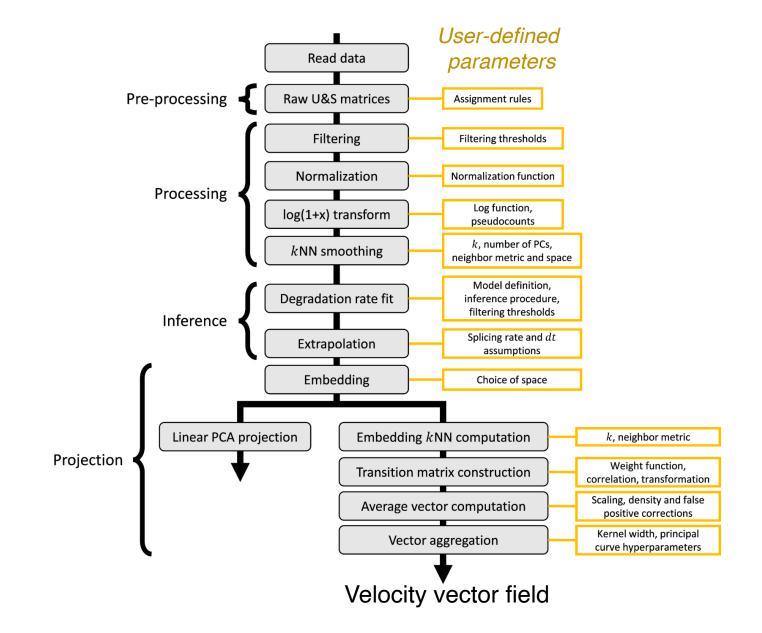
Step 2. Velocity estimation and visualization

Most widely-used tools are:

- velocyto (<u>https://velocyto.org/velocyto.py/tutorial/analysis.html</u>)
- scvelo (<u>https://scvelo.readthedocs.io/</u>)

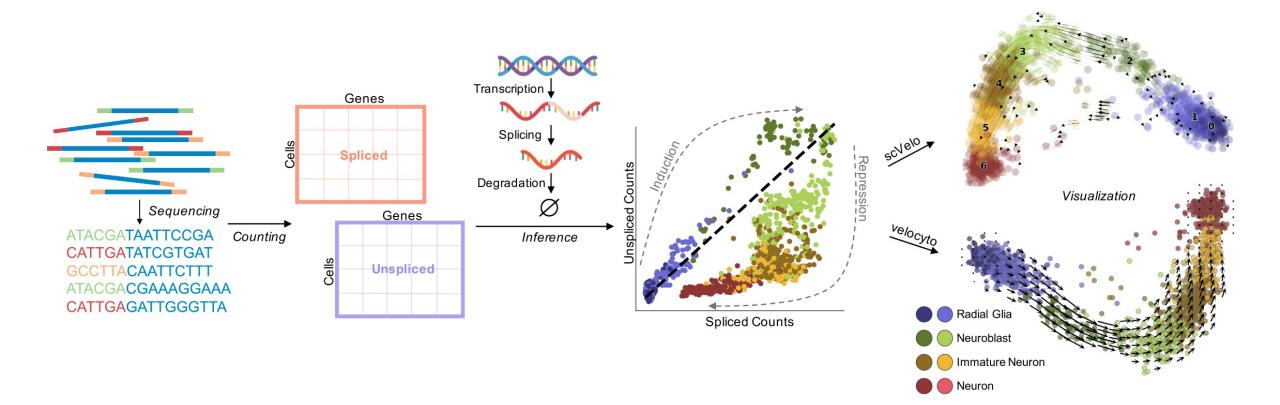
Most RNA velocity tools are implemented and better-supported in Python.

The RNA velocity workflow is complex!



"RNA velocity unraveled" Gorin et al PLoS Comp Biol 2022

A recap of RNA velocity analysis



"RNA velocity unraveled" Gorin et al PLoS Comp Biol 2022